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(54) Title: NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION

(57) Abstract: The present invention encompasses isolated nucleic acids containing transcriptional units which encode a signal sequence of one flavivirus and an immunogenic flavivirus antigen of a second flavivirus or of a chimeric immunogenic flavivirus antigen comprising sequence from more than one flavivirus. The invention further encompasses a nucleic acid and protein vaccine and the use of the vaccine to immunize a subject against flavivirus infection. The invention also provides antigens encoded by nucleic acids of the invention, antibodies elicited in response to the antigens and use of the antigens and/or antibodies in detecting flavivirus or diagnosing flavivirus infection.

NUCLEIC ACID VACCINES FOR PREVENTION OF FLAVIVIRUS INFECTION

This application is a continuation-in-part of, and claims the benefit of, U.S. application Serial No. 09/826,115, filed April 4, 2001, which status is pending and
5 which is continuation-in-part of, and claims the benefit of U.S. application Serial No. 09/701,536, filed November 29, 2000, which status is pending and which is a national stage application of international application Serial No. PCT/US99/12298, filed June 3, 1999 from, and which claims the benefit of, U.S. provisional application Serial No. 60/087,908, filed June 4, 1998, which applications are hereby incorporated herein in
10 their entirety by reference.

Field of the Invention

This invention relates to novel vaccines, diagnostics and methods of using both in the treatment and prevention of the diseases caused by flaviviruses. In particular,
15 the vaccines are recombinant nucleic acids which contain genes for structural proteins of flaviviruses, such as Japanese encephalitis virus (JEV), West Nile virus (WNV) or related flaviviruses. These vaccines serve as a transcriptional unit for the biosynthesis of the virus protein antigens when administered *in vivo*. The diagnostics are compositions containing antigens produced from the recombinant nucleic acids that can
20 be used to detect flavivirus infection.

Background of the Invention

Flaviviruses are members of the genus *Flavivirus*, which is classified within the family Flaviviridae. The flaviviruses are largely pathogenic to humans and other mammals. Flaviviruses that inflict disease upon humans and animals include Alfuy,
25 Apoi, Aroa, Bagaza, Banzi, Batu Cave, Bouboui, Bukalasa bat, Bussuquara, Cacipacore, Carey Island, Cowbone Ridge, Dakar bat, Dengue (serotypes 1, 2, 3 and 4), Edge Hill, Entebbe bat, Gadgets Gully, Iguape, Ilheus, Israel turkey

meningoencephalitis, Japanese encephalitis, Jugra, Jutiapa, Kadam, Karshi, Kedougou, Kokobera, Koutango, Kunjin, Kyasanur Forest disease, Langat, Meaban, Modoc, Montana myotis leukoencephalitis, Murray Valley encephalitis, Naranjal, Negishi, Ntaya, Omsk hemorrhagic fever, Phnom Penh bat, Potiskum, Powassan, Rio Bravo, Rocio, Royal Farm, Russian spring summer encephalitis, Saboya, Sal Vieja, San Perlita, Saumarez Reef, Sepik, Sokuluk, Spondweni, St. Louis encephalitis, Stratford, Tick-borne encephalitis - central European subtype, Tick-borne encephalitis - far eastern subtype, Tembusu, THCAr, Tyuleny, Uganda S, Usutu, West Nile, Yaounde, Yellow fever, Yokose, Ziki, Cell fusing agent and other related flaviviruses, as listed in Kuno et al. (*J. Virol.* 72: 73-83 (1998)).

The flaviviruses contain the following three structural proteins: prM/M, the premembrane and membrane protein; E, the envelope protein; and C, the capsid protein. (Monath, in Virology (Fields, ed.), Raven Press, New York, 1990, pp. 763-814; Heinz and Roehrig, in Immunochemistry of Viruses II: The Basis for Serodiagnosis and Vaccines (van Regenmortel and Neurath, eds.), Elsevier, Amsterdam, 1990, pp. 289-305). M has a molecular weight (MW) of about 7-8 kilodaltons (kDa) and E has a MW of about 55-60 kDa. M is synthesized as a larger precursor termed prM. The pr portion of prM is removed when prM is processed to form M protein in mature virions. M and E are located in the membrane of the flavivirus particle, and so have long been considered to constitute important immunogenic components of the viruses.

The flaviviruses are RNA viruses comprising single stranded RNA having a length, among the various species, of about 10 kilobases (kb). The C protein, with a MW of 12-14 kDa, complexes with the RNA to form a nucleocapsid complex. Several nonstructural proteins are also encoded by the RNA genome which are termed NS1, NS2A, NS2B, NS3, NS4A, NS4B and NS5. The genome is translated within the host cell as a polyprotein, then processed co- or post-translationally into the individual gene products by viral- or host-specific proteases (Figure 1).

The nucleotide sequences of the genomes of several flaviviruses are known, as summarized in U. S. Patent No. 5,494,671. That for JEV is provided by Sumiyoshi et al. (*Virology* 161: 497-510 (1987)) and Hashimoto et al. (*Virus Genes* 1: 305-317 (1988)). The nucleotide sequences of the virulent strain SA-14 of JEV and the
5 attenuated strain SA-14-14-2, used as a vaccine in the People's Republic of China, are compared in the work of Nitayaphan et al. (*Virology* 177: 541-552 (1990)).

Nucleotide sequences encoding the structural proteins of other flavivirus species are also known. In many cases, the sequences for the complete genomes have been reported. The sequences available include dengue serotype 1 virus, dengue serotype 2
10 virus (Deubel et al., *Virology* 155: 365-377 (1986); Gruenberg et al., *J. Gen. Virol.* 69: 1391-1398 (1988); Hahn et al. *Virology* 162: 167-180 (1988)), dengue serotype 3 virus (Osatomi et al., *Virus Genes* 2: 99-108 (1988)), dengue serotype 4 virus (Mackow et al., *Virology* 159: 217-228 (1987), Zhao et al., *Virology* 155: 77-88 (1986)), West Nile virus (Lanciotti et al., *Science* 286: 2331-2333 (1999)), Powassan virus (Mandl et al.,
15 *Virology* 194: 173-184 (1993)) and yellow fever virus (YFV) (Rice et al., *Science* 229: 726-733 (1985)).

Many flaviviruses, including St. Louis encephalitis virus (SLEV), WNV and JEV, are transmitted to humans and other host animals by mosquitoes. They therefore occur over widespread areas and their transmission is not easily interrupted or
20 prevented.

West Nile fever is a mosquito-borne flaviviral infection that is transmitted to vertebrates primarily by various species of *Culex* mosquitoes. Like other members of the Japanese encephalitis (JE) antigenic complex of flaviviruses, including JE, SLE and Murray Valley encephalitis (MVE) viruses, WNV is maintained in a natural cycle
25 between arthropod vectors and birds. The virus was first isolated from a febrile human in the West Nile district of Uganda in 1937 (Smithburn et al., *Am. J. Trop. Med. Hyg.* 20: 471-492 (1940)). It was soon recognized as one of the most widely distributed flaviviruses, with its geographic range including Africa, the Middle East, Western Asia, Europe and Australia (Hubalek et al., *Emerg. Infect. Dis.* 5: 643-50 (1999)). Clinically,

West Nile fever in humans is a self-limited acute febrile illness accompanied by headache, myalgia, polyarthropathy, rash and lymphadenopathy (Monath and Tsai, in Clinical Virology, (Richman, Whitley and Hayden eds.), Churchill-Livingstone, New York, 1997, pp. 1133-1186). Acute hepatitis or pancreatitis has been reported on
5 occasion and cases of WNV infection in elderly patients are sometimes complicated by encephalitis or meningitis (Asnis et al., *Clin. Infect. Dis.* 30: 413-418 (2000)). Thus, infection by WNV is a serious health concern in many regions of the world.

The geographical spread of the disease, particularly the introduction of WNV into the U.S. in 1999, has greatly increased awareness of the human and animal health
10 concerns of this disease. Between late August and early September 1999, New York City and surrounding areas experienced an outbreak of viral encephalitis, with 62 confirmed cases, resulting in seven deaths. Concurrent with this outbreak, local health officials observed increased mortality among birds (especially crows) and horses. The outbreak was subsequently shown to be caused by WNV, based on monoclonal
15 antibody (Mab) mapping and detection of genomic sequences in human, avian and mosquito specimens (Anderson et al., *Science* 286: 2331-2333 (1999); Jia et al., *Lancet* 354: 1971-1972 (1999); Lanciotti et al., *Science* 286: 2333-2337 (1999)). Virus activity detected during the ensuing winter months indicated that the virus had established itself in North America (*Morb. Mortal. Wkly. Rep.* 49: 178-179 (2000); Asnis et al., *Clin.*
20 *Infect. Dis.* 30: 413-418 (2000); Garmendia et al., *J. Clin. Micro.* 38: 3110-3111 (2000)). Surveillance data reported from the northeastern and mid-Atlantic states during the year 2000 confirmed an intensified epizootic/epidemic transmission and a geographic expansion of the virus with documentation of numerous cases of infection in birds, mosquitoes and horses, as well as cases in humans (*Morb. Mortal. Wkly. Rep.*
25 49: 820-822 (2000)).

Currently, no human or veterinary vaccine is available to prevent WNV infection and mosquito control is the only practical strategy to combat the spread of the disease.

Japanese encephalitis virus (JEV) infects adults and children and there is a high mortality rate among infants, children and the elderly in areas of tropical and subtropical Asia (Tsai et al., in Vaccines (Plotkin, ed.) W.B. Saunders, Philadelphia, Pa, 1999, pp. 672-710). Among survivors, there are serious neurological consequences, related to the symptoms of encephalitis, that persist after infection. In more developed countries of this region, such as Japan, the Republic of China (Taiwan) and Korea, JEV has been largely controlled by use of a vaccine of inactivated JEV. Nevertheless, it is still prevalent in other countries of the region.

Vaccines available for use against JEV infection include live virus inactivated by such methods as formalin treatment, as well as attenuated virus (Tsai et al., in Vaccines (Plotkin, ed.) W.B. Saunders, Philadelphia, Pa, 1994, pp. 671-713). Whole virus vaccines, although effective, do have certain problems and/or disadvantages. The viruses are cultivated in mouse brain or in cell culture using mammalian cells as the host. Such culture methods are cumbersome and expensive. Furthermore, there is the attendant risk of incorporating antigens from the host cells, i.e., the brain or other host, into the final vaccine product, potentially leading to unintended and undesired allergic responses in the vaccine recipients. There is also the risk of inadvertent infection among workers involved in vaccine production. Finally, there is the risk that the virus may not be fully or completely inactivated or attenuated and thus, the vaccine may actually cause disease.

Dengue fever and dengue hemorrhagic fever (DF/DHF) are caused by dengue virus, which is also a mosquito-borne flavivirus. There are four antigenically related, but distinct, dengue virus serotypes, (DEN-1, DEN-2, DEN-3 and DEN-4), all of which can cause DF/DHF. Symptoms of DF, the mild form of dengue-related disease, include fever, rash, severe headache and joint pain. Mortality among those subjects suffering from DF is low; however, among those subjects suffering from DHF, mortality can be as high as 5%. From available evidence, more than 3 million cases of DHF and 58,000 deaths have been attributed to DHF over the past 40 years, making DHF a major emerging disease (Halstead, in Dengue and Dengue Hemorrhagic Fever (Gubler and Kuno, eds.) CAB International, New York, NY, (1997) pp 23-44). Nevertheless,

despite decades of effort, safe and effective vaccines to protect against dengue virus infection are not yet available.

Yellow fever is prevalent in tropical regions of South America and sub-Saharan Africa and is transmitted by mosquitos. Infection leads to fever, chills, severe headache and other pains, anorexia, nausea and vomiting, with the emergence of jaundice. A live virus vaccine, 17D, grown in infected chicken embryos, is considered safe and effective. Nevertheless, there remains a need for a vaccine that is stable under adverse conditions, such as are commonly encountered in the tropical regions of Africa and the Americas where the vaccine is most needed.

10 A recombinant flavivirus which is a chimera between two flaviviruses is disclosed in PCT publication WO 93/06214. The chimera is a construct fusing non-structural proteins from one "type," or serotype, of dengue virus or a flavivirus, with structural proteins from a different "type," or serotype, of dengue virus or other flavivirus.

15 Several recombinant subunit and viral vaccines have been devised in recent years. U.S. Patent No. 4,810,492 describes the production of the E glycoprotein of JEV for use as the antigen in a vaccine. The corresponding DNA is cloned into an expression system in order to express the antigen protein in a suitable host cell such as *E. coli*, yeast, or a higher organism cell culture. U. S. Patent No. 5,229,293 discloses
20 recombinant baculovirus harboring the gene for JEV E protein. The virus is used to infect insect cells in culture such that the E protein is produced and recovered for use as a vaccine.

U.S. Patent No. 5,021,347 discloses a recombinant vaccinia virus genome into which the gene for JEV E protein has been incorporated. The live recombinant
25 vaccinia virus is used as the vaccine to immunize against JEV. Recombinant vaccinia viruses and baculoviruses in which the viruses incorporate a gene for a C-terminal truncation of the E protein of dengue serotype 2, dengue serotype 4 and JEV are disclosed in U.S. Patent 5,494,671. U.S. Patent 5,514,375 discloses various

recombinant vaccinia viruses which express portions of the JEV open reading frame extending from prM to NS2B. These pox viruses induced formation of extracellular particles that contain the processed M protein and the E protein. Two recombinant viruses encoding these JEV proteins produced high titers of neutralizing and

5 hemagglutinin-inhibiting antibodies, and protective immunity, in mice. The extent of these effects was greater after two immunization treatments than after only one. Recombinant vaccinia virus containing genes for the prM/M and E proteins of JEV conferred protective immunity when administered to mice (Konishi et al., *Virology* 180: 401-410 (1991)). HeLa cells infected with recombinant vaccinia virus bearing genes

10 for prM and E from JEV were shown to produce subviral particles (Konishi et al., *Virology* 188: 714-720 (1992)). Dmitriev et al. reported immunization of mice with a recombinant vaccinia virus encoding structural and certain nonstructural proteins from tick-borne encephalitis virus (*J. Biotechnology* 44: 97-103 (1996)).

Recombinant virus vectors have also been prepared to serve as virus vaccines

15 for dengue fever. Zhao et al. (*J. Virol.* 61: 4019-4022 (1987)) prepared recombinant vaccinia virus bearing structural proteins and NS1 from dengue serotype 4 and achieved expression after infecting mammalian cells with the recombinant virus. Similar expression was obtained using recombinant baculovirus to infect target insect cells (Zhang et al., *J. Virol.* 62: 3027-3031(1988)). Bray et al. (*J. Virol.* 63: 2853-2856

20 (1989)) also reported a recombinant vaccinia dengue vaccine based on the E protein gene that confers protective immunity to mice against dengue encephalitis when challenged. Falgout et al. (*J. Virol* 63: 1852-1860 (1989)) and Falgout et al. (*J. Virol.* 64: 4356-4363 (1990)) reported similar results. Zhang et al. (*J. Virol* 62: 3027-3031 (1988)) showed that recombinant baculovirus encoding dengue E and NS1 proteins

25 likewise protected mice against dengue encephalitis when challenged. Other combinations in which structural and nonstructural genes were incorporated into recombinant virus vaccines failed to produce significant immunity (Bray et al., *J. Virol.* 63: 2853-2856 (1989)). Also, monkeys failed to develop fully protective immunity to dengue virus challenge when immunized with recombinant baculovirus expressing the

30 E protein (Lai et al. (1990) pp. 119-124 in F. Brown, R. M. Chancock, H. S. Ginsberg

and R. Lerner (eds.) Vaccines 90: Modern approaches to new vaccines including prevention of AIDS, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY).

Immunization using recombinant DNA preparations has been reported for SLEV and dengue-2 virus, using weanling mice as the model (Phillpotts et al., *Arch. Virol.* 141: 743-749 (1996); Kochel et al., *Vaccine* 15: 547-552 (1997)). Plasmid DNA encoding the prM and E genes of SLEV provided partial protection against SLEV challenge with a single or double dose of DNA immunization. In these experiments, control mice exhibited about 25% survival and no protective antibody was detected in the DNA-immunized mice (Phillpotts et al., *Arch. Virol.* 141: 743-749 (1996)). In mice that received three intradermal injections of recombinant dengue-2 plasmid DNA containing prM, 100% developed anti-dengue-2 neutralizing antibodies and 92% of those receiving the corresponding E gene likewise developed neutralizing antibodies (Kochel et al., *Vaccine* 15: 547-552 (1997)). Challenge experiments using a two-dose schedule, however, failed to protect mice against lethal dengue-2 virus challenge.

The vaccines developed to date for immunizing against infection by JEV, SLEV, dengue virus and other flaviviruses have a number of disadvantages and problems attending their use. Inactivated vaccine is costly and inconvenient to prepare. In addition, any such vaccine entails the risk of allergic reaction originating from proteins of the host cell used in preparing the virus. Furthermore, such vaccines present considerable risk to the workers employed in their production. Candidate attenuated JEV vaccines are undergoing clinical trials, but as of 1996 have not found wide acceptance outside of the People's Republic of China (Hennessy et al., *Lancet* 347: 1583-1586 (1996)).

Recombinant vaccines based on the use of only certain proteins of flaviviruses, such as JEV, produced by biosynthetic expression in cell culture with subsequent purification or treatment of antigens, do not induce high antibody titers. Also, like the whole virus preparations, these vaccines carry the risk of adverse allergic reaction to antigens from the host or to the vector. Vaccine development against dengue virus and

WNV is less advanced and such virus-based or recombinant protein-based vaccines face problems similar to those alluded to above.

There is therefore a need for vaccines or improved vaccines directed against flaviviruses such as yellow fever virus, dengue virus, JEV, SLEV and WNV which are inexpensive to prepare, present little risk to workers involved in their manufacture, carry minimal risk of adverse immunological reactions due to impurities or adventitious immunogenic components and are highly effective in eliciting neutralizing antibodies and protective immunity. There is furthermore a need for a vaccine against JEV, WNV and related flaviviruses that minimizes the number of immunizing doses required.

10

Many of the shortcomings of the current art as described in detail for the production of vaccines also apply to the production of antigens and antibodies to be used for the production of immunodiagnostics. Particularly, the concurrent risks and costs involved in the production of antigens from viruses and the failure of most currently available recombinantly expressed antigens to elicit effective immune responses are paralleled in the field of immunodiagnostics by the same risks, high costs and a corresponding lack of sensitivity. Thus, because of the high costs, risk of accidental infection with live virus and the lower than desired levels of sensitivity of the previously available tests, there exists a need for rapid, simple and highly sensitive diagnostic tests for detecting flavivirus infection and/or contamination.

20

The present invention meets these needs by providing highly immunogenic recombinant antigens for use in diagnostic assays for the detection of antibodies to selected flaviviruses. The present invention further provides for the use of recombinant antigens derived from flaviviruses, flavivirus genes or mimetics thereof in immunodiagnostic assays for the detection of antibodies to flavivirus proteins.

25

Summary of the Invention

The present invention provides a nucleic acid molecule which contains a transcriptional unit (TU) for an immunogenic flavivirus antigen. The TU directs a host

cell, after being incorporated within the cell, to synthesize the antigen. In an important aspect of the invention, the flavivirus can be yellow fever virus (YFV), dengue serotype 1 virus (DEN-1), dengue serotype 2 virus (DEN-2), dengue serotype 3 virus (DEN-3), dengue serotype 4 virus (DEN-4), St. Louis encephalitis virus (SLEV), Japanese encephalitis virus (JEV), West Nile virus (WNV), Powassan virus or any other flavivirus. In important embodiments of the present invention, the antigen can be the flavivirus prM/M protein, the E protein, or both. In important embodiments of the present invention, the antigen can be a chimeric flavivirus protein. In particular, when the TU includes both the prM/M and E proteins, the host cell secretes subviral particles containing the prM/M and E antigens. In a further important aspect of the invention, the nucleic acid is a DNA molecule. In additional significant embodiments, the nucleic acid TU includes a control sequence disposed appropriately such that it operably controls the expression of the prM/M and E antigens and this control sequence can be the cytomegalovirus immediate early promoter. In an additional embodiment, the nucleotide sequence of the TU is engineered to optimize eukaryotic translation by minimizing large hairpin structures in the 5'-end untranslated region of an mRNA produced by the TU and/or the inclusion of a Kozak consensus sequence at the translational start site of an mRNA produced by the TU. In an additional embodiment, the transcriptional unit also includes a poly-A terminator.

20 The present invention further provides a host cell comprising a nucleic acid molecule which includes a transcriptional unit for an immunogenic flavivirus antigen that directs the host cell to synthesize the immunogenic antigen. The flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. In important embodiments, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins. In the latter case, the cell secretes subviral particles containing the prM/M and E antigens.

30 Additionally, the invention provides a composition for vaccinating a subject against a flavivirus containing a nucleic acid molecule that includes a transcriptional unit for an immunogenic flaviviral antigen. The transcriptional unit directs a cell within the body of the subject, after being incorporated therein, to synthesize the

immunogenic antigen. The composition further includes a pharmaceutically acceptable carrier. In significant embodiments, the flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. Furthermore, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins.

- 5 In the latter instance, the cell secretes subviral particles comprising the flavivirus prM/M and E antigens. These subviral particles are also referred to as noninfectious recombinant antigen (NRA). In important embodiments, the nucleic acid molecule is a DNA molecule. In further significant embodiments, the transcriptional unit additionally contains a control sequence disposed appropriately such that it operably
- 10 controls the synthesis of the prM/M and E antigens when the nucleic acid is introduced into the cell of the subject. This control sequence can be the cytomegalovirus immediate early promoter. In a still further embodiment, the transcriptional unit can also include a poly-A terminator.

- 15 The compositions provided by the present invention for vaccinating a subject against a flavivirus can include a nucleic acid molecule, or molecules, that include transcriptional units for more than one immunogenic flaviviral antigen. The more than one immunogenic flaviviral antigen can be from different flavivirus species, strains or isolates in any combination. In significant embodiments, the flaviviruses included can
- 20 be two or more, three or more, four or more, five or more, or seven or more flaviviruses. Examples of such flaviviruses include, but are not limited to YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flaviviruses. Combination vaccines can be formulated to confer immunity to flavivirus disease common to particular geographical regions. In a particular embodiment directed
- 25 toward tropical and subtropical Asia, DEN-1, DEN-2, DEN-3, DEN-4, WN, and JE viruses can be selected. In a particular embodiment directed to Africa, DEN-1, DEN-2, DEN-3, DEN-4, WN and YF can be selected. In a particular embodiment directed to Latin America, DEN-1, DEN-2, DEN-3, DEN-4, Rocio and YF viruses can be selected.

- The invention provides still further a method of immunizing a subject against
- 30 infection by a flavivirus. The method involves administering to the subject an effective amount of a vaccinating composition that contains a nucleic acid molecule which

includes a transcriptional unit for an immunogenic flavivirus antigen. The transcriptional unit directs a cell within the body of the subject, after being taken up by the cell, to synthesize the immunogenic antigen. The composition additionally includes a pharmaceutically acceptable carrier. In significant embodiments of the method, the
5 flavivirus may be YFV, DEN-1, DEN-2, DEN-3, DEN-4, SLEV, JEV, WNV, Powassan virus or other flavivirus. In yet other important aspects of the method, the antigen may be the prM/M protein, the E protein, or both the prM/M and the E proteins. When the antigen is both the prM/M and the E proteins, the cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles
10 comprising the flaviviral prM/M and E antigens. Additionally, in significant embodiments of the method, the vaccinating composition is administered to the subject in a single dose, via a parenteral route. In yet a further aspect of the method, the nucleic acid is a DNA molecule. In yet additional embodiments of the method, the transcriptional unit further includes a control sequence disposed appropriately such that
15 it operably controls the synthesis of the prM/M and E antigens and in a significant aspect of this embodiment, the control sequence is the cytomegalovirus immediate early promoter. Furthermore, the transcriptional unit may include a poly-A terminator.

These aspects and embodiments of the invention are the basis for its distinct attributes and advantages. Being a nucleic acid construct involving only portions of the
20 flavivirus genome rather than the sequence encompassing the complete genome, the nucleic acid TU-containing vaccine is completely nonviable. It therefore poses no danger of infection by the flavivirus to those involved in its manufacture or to subjects receiving the vaccine. The nucleic acid vaccine is easy to prepare and easy to administer and is stable in storage prior to use. Unexpectedly it has been found that the
25 nucleic acid vaccine of the invention is essentially 100% successful in conferring protective immunity in mammals after administering only a single dose. A further unexpected result is that the nucleic acid TU is able to engender immunity to a flavivirus in a female mammal which can be transmitted to its progeny through the milk. Without wishing to be limited by theory, the inventor believes that a possible
30 mechanism for the success of the nucleic acid in conferring protective immunity is that a host cell harboring the nucleic acid, such as the cell of a subject to whom the vaccine

is administered, produces subviral particles containing the flaviviral prM/M and E antigens. These particles mimic the immunogenic attributes of native flavivirus virions.

The present invention also provides noninfectious antigenic polypeptides, antigenic polypeptide fragments and NRA comprising the prM/M and/or E proteins of flaviviruses, wherein the transmembrane signal sequence is derived from a first flavivirus and the M and/or E proteins are derived from a second flavivirus. Further, the prM/M protein can comprise amino acid sequences from both the first and the second flaviviruses. Further, the E protein can comprise amino acid sequences from both the first and second flaviviruses. "Chimeric" as used herein means any protein or nucleic acid comprising sequence from more than one flavivirus. As used herein, "non-virulent" means the antigen or vaccine of this invention is incapable of causing disease. More particularly, the recombinant protein antigens are free of contaminating genomic material from flaviviruses that is necessary for flavivirus infection, replication and pathogenesis.

The polypeptides of the present invention can comprise the amino acid sequences defined herein, or that are known in the art, of the prM, M and/or E proteins of selected flaviviruses. The nucleic acids of this invention can comprise nucleotide sequence that encodes the prM, M and/or E proteins of selected flaviviruses.

The antigens of the present invention can be unconjugated, or they can be conjugated to a carrier molecule that facilitates placement of the antigen on a solid phase. A carrier molecule is one to which antigens can be conjugated and which will not react with antibodies in human serum. An example of such a carrier is bovine serum albumin (BSA).

The antigens of the present invention can also be recombinant proteins obtained by expressing nucleic acids encoding the antigen in an expression system capable of producing the antigen.

The amino acid sequences of the present antigens can contain an immunoreactive portion of the prM, M and/or E antigen. These antigens may further be attached to sequences designed to provide for some additional property, such as to remove/add amino acids capable of disulfide bonding to increase the reactivity of an epitope by providing a more rigid secondary structure, to increase its bio-longevity or to alter its cytotoxicity or to prevent infection. In any case, the antigen must possess immunoreactivity and/or immunogenicity.

Brief Description of the Drawings

Figure 1 is a schematic representation of flaviviral polyprotein processing. The central horizontal region provides a schematic representation of the viral genome. The lines denote the 5' and 3' non-translated regions and the boxed regions represent the open reading frame for structural (left and top) and non-structural (right and bottom) proteins. Cleavage by host cell signalase occurs simultaneously with translation at the E protein C-terminus, separating structural and non-structural regions. A subtilase-like cellular enzyme, furin, may be responsible for prM cleavage. Potential transmembrane domains of viral polyprotein are indicated by shaded areas.

Figure 2 is a map of the JEV genome (top) and the DNA sequence of oligonucleotides used in a reverse transcriptase-polymerase chain reaction (RT-PCR) (center) to construct the transcription unit for the expression of prM-E protein coding regions (bottom). Potential transmembrane domains of viral polyprotein are indicated by shaded areas.

Figure 3 shows a schematic representation of the plasmid vectors, pCDNA3, pCBamp, and pCIBamp, and the relationship between them. These plasmids include the CMV (cytomegalovirus) promoter/enhancer element, BGHp(A) (bovine growth hormone polyadenylation signal and transcription termination sequence), ampicillin resistance gene and ColE1 origin of replication for selection and maintenance in *E. coli*. The f1 origin of replication for single-stranded rescue in *E. coli* cells, SV40 origin of replication (SV40 ORI), neomycin resistance coding region and SV40p(A) sequences

were deleted from pCDNA3 to generate pCBamp. An intron sequence was inserted in the NcoI-KpnI site of pCBamp to generate plasmid pCIBamp.

Figure 4 shows SDS-PAGE-immunoblot analyses of the sucrose gradient purified subviral particles from JE-4B COS-1 culture fluid (4B, right lane of each pair).

- 5 The density gradient purified JE virion from JEV infected C6/36 cell culture was used as a positive control (JEV, left lane of each pair). JE HIAF (hyperimmune ascitic fluid); 4G2, anti-E monoclonal antibody; JM01, anti-M monoclonal antibody; NMAF (normal mouse ascitic fluid).

- Figure 5 shows a profile of the E antigen in a rate zonal sucrose gradient analysis prepared from the PEG precipitate of JE-4B cell culture medium with or without Triton X-100 treatment.
- 10

- Figure 6 shows signal peptide probability of the pCBE1-14 (pCBE) predicted by the SignalP-HMM program (A). The signal peptide probability is improved by altering the c-region sequence at -4 and -2 positions (C-4G and G-2S) (panel B, JE-LSS-M), by shortening the n-region (panel C, JE-SS-ORI), or by a combination of both modifications (panel D, JE-SS-M).
- 15

- Figure 7 shows schematic representations of plasmid vectors pCBD2-14-16 (100% DEN-2 E), pCBD2-1J-4-3 (90% DEN-2 E : 10% JEV E), and pCB8D2-2J-2-9-1 (80% DEN-2 E : 20% JEV E). These plasmids include the human cytomegalovirus (CMV) early gene promoter; JE virus signal sequence; DEN-2 virus prM and E gene region (amino terminal 100%, 90%, or 80%, respectively); JE virus E gene region (none, 10% or 20%, respectively); and bovine growth hormone poly A signal (BGH).
- 20

- Figure 8 shows a comparison of secreted and membrane-bound recombinant protein by western blot. (A) Analysis of secreted recombinant antigen following PEG-precipitation and ethanol extraction of culture fluid for DEN-2 plasmids pCB8D2-2J-2-9-1, pCB9D2-1J-4-3, pCBD2-14-16, and control plasmid pEGFP. Lane 1(V), purified DEN-2 virus stained by Gold Blot (Owl Separation Systems, Portsmouth, NH).
- 25

- Reactivity of secreted, recombinant antigen from each plasmid with **a**, anti-envelope (E) specific Mab 1A6A-8; **b**, a mixture of MAB 1A6A-8, anti-capsid (C) specific Mab 1A2A-1, anti-serum specific for DEN-2 virus premembrane (prM) protein; and **c**, normal mouse ascites. (B) Analysis of recombinant plasmid-transformed cell
- 5 hydrophobic membrane proteins. Lane 1 (V), purified DEN-2 virus stained by Gold Blot; lane 2 (V), reactivity of purified DEN-2 virus with a mixture of Mab 1A6A-8, Mab 1A2A-1, anti-serum specific for DEN-2 virus M protein, and anti-serum for DEN-2 virus prM protein. Reactivity of isolated hydrophobic membrane proteins from each plasmid-transformed cell line with **a**, Mab 1A6A-8; **b**, a mixture of Mab 1A6A-8, Mab
- 10 1A2A-1, anti-serum specific for DEN-2 virus M protein, and anti-serum for DEN-2 virus prM protein; and **c**, normal mouse ascites.

Detailed Description of the Invention

- The invention encompasses nucleic acid transcriptional units which encode flaviviral antigenic proteins, such as the prM/M and E protein antigens. The nucleic
- 15 acids function to express the prM/M and E protein antigens when the nucleic acid is taken up by an appropriate cell, especially when the cell is the cell of a subject. The invention also encompasses a vaccine whose active agent is the nucleic acid transcriptional unit (TU). The invention further encompasses cells containing a TU. The invention in addition encompasses a method of immunizing a subject against
- 20 flaviviral infection by administering to the subject an effective amount of a vaccine containing the nucleic acid TU molecules.

- The invention provides an isolated nucleic acid comprising a transcriptional unit encoding a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus, wherein the transcriptional unit
- 25 directs the synthesis of the antigen. The invention further encompasses the use of the nucleic acid transcriptional unit (TU) to generate flaviviral antigens and the flaviviral antigens produced by the nucleic acid TU. The flaviviral antigens encompassed by the present invention include chimeric flavivirus antigens incorporating amino acid

sequence from a first flavivirus and at least one further flavivirus. The invention still further encompasses the use of the flaviviral antigens encoded by the TU of the invention to produce flavivirus-specific antibodies and to detect the presence of flavivirus-specific antibodies.

- 5 In one embodiment, the isolated nucleic acid of this invention can comprise a transcriptional unit encoding a Japanese encephalitis virus signal sequence.

 In another embodiment, the transcriptional unit of this invention can encode an immunogenic flavivirus antigen which can be from one or more of the following flaviviruses: yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus,
10 dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

 In another embodiment, the transcriptional unit of this invention can encode an immunogenic chimeric flavivirus antigen which can include sequence from more than one of the following flaviviruses: yellow fever virus, dengue serotype 1 virus, dengue
15 serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

 In a particular embodiment, the nucleic acid of this invention can encode a signal sequence of Japanese encephalitis virus and an M protein and an E protein of West Nile virus, SLEV, YFV and/or Powassan virus. The nucleic acid can also encode
20 an immunogenic antigen which can be an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and/or both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus. In a preferred embodiment, the isolated nucleic acid encodes both the M protein and the E protein of
25 the flavivirus. Further, the nucleic acid of the invention can be DNA and can comprise nucleotide sequence SEQ ID NO:15, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23 or SEQ ID NO: 42.

In another particular embodiment, the nucleic acid of this invention can encode a signal sequence of Japanese encephalitis virus, an M protein of a second virus and a chimeric E protein formed by from substitution of a portion of the nucleic acid encoding the second virus' E protein with nucleic acid encoding the corresponding
5 portion of the JEV E protein. Alternatively, the portion of sequence corresponding to the deleted portion of the second virus' E protein can be substituted by other sequence selected from a third virus or it can be a non-viral sequence. The second protein can be West Nile virus, SLEV, YFV, Powassan and/or a serotype of Dengue virus . Chimeric E proteins can include those where the carboxy terminal portion can be from one
10 flavivirus and the remainder of the chimeric E protein is from another flavivirus. The carboxy terminal portion can be, for example, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 60, or 75% of the chimeric E protein. The nucleic acid of the invention can be DNA and can comprise the protein-encoding sequence from nucleotide sequence SEQ ID NO:44 or SEQ ID NO:46. The nucleic acid of the invention can comprise nucleotide sequence
15 SEQ ID NO:44 or SEQ ID NO:46.

The transcriptional unit of this invention can also comprise a control sequence disposed appropriately so that it operably controls the synthesis of the antigen. The control sequence can be, for example, the cytomegalovirus immediate early promoter. The nucleic acid of this invention can also comprise a Kozak consensus sequence
20 located at a translational start site for a polypeptide comprising the antigen encoded by the transcriptional unit. The transcriptional unit of this invention can also comprise a poly-A terminator.

The present invention further provides a cell comprising the nucleic acid of this invention.

25 Also provided is a composition comprising a pharmaceutically acceptable carrier and nucleic acid or cell or antigen of this invention. The present invention additionally provides a method of immunizing a subject against infection by a flavivirus, comprising administering to the subject an effective amount of a composition of this invention. In a particular embodiment, the composition used to

immunize a subject directs the synthesis of both the M protein and the E protein of a flavivirus and a cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles comprising the M protein and the E protein. Alternatively, the composition can comprise an M protein and/or E protein of a
5 flavivirus or subviral particles comprising the M protein and E protein. In the methods of this invention, the immunizing composition can be administered to the subject in a single dose and can be administered via a parenteral route.

This invention further provides the antigens produced from the isolated nucleic acids of this invention. As an example, the antigen from the second flavivirus encoded
10 by the nucleotide sequence of TU can be the M protein which can be, for example, from West Nile virus. The antigen can also be protein from dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus. In a further embodiment, the antigen comprises a prM/M protein comprising the transmembrane signal sequence from a first flavivirus and further amino acid sequence
15 comprising the remainder of the prM/M protein from a second flavivirus, which can be from SLEV, JEV, YFV, WNV and/or Powassan virus. The transmembrane signal sequence from a first flavivirus can be an improved or modified signal sequence wherein the signal sequence imparts desired characteristics such as a high signal sequence probability. Accomplishing these goals by design or selection can be with the
20 use of machine-learning computer programs including, but not limited to, those using a hidden Markov model.

The antigen encoded by the nucleotide sequence of the TU can be West Nile virus antigen, dengue virus antigen, St. Louis encephalitis virus antigen, Japanese encephalitis virus antigen, Powassan virus antigen and/or yellow fever virus antigen.

25 The antigen encoded by the nucleotide sequence of the TU can also be the E protein, which can be the E protein from West Nile virus, dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus. The antigen encoded can also be a chimeric E protein comprising amino acid sequence selected from more than one flavivirus.

Additionally, the antigen encoded by the nucleotide sequence of the TU can be the M protein and the E protein, which can be from West Nile virus, dengue virus, St. Louis encephalitis virus, Japanese encephalitis virus, Powassan virus and/or yellow fever virus.

5 As used herein, "M protein" or "prM protein" or "prM/M protein" means a flavivirus M protein or flavivirus prM protein. Examples include, but are not limited to, prM proteins comprising amino acid sequence from one or more flavivirus prM proteins, M proteins comprising no additional amino acid sequence and proteins comprising additional amino acid sequences which are processed *in vitro* or *in vivo* to
10 generate the mature M protein.

 As used herein, "nucleic acid transcriptional unit" or "nucleic acid transcriptional unit molecule" means a nucleic acid encoding one or more specified proteins. The TU has biological activity such that, after having been introduced into a suitable cell, the nucleic acid induces the synthesis of one or more specified gene
15 products encoded by the nucleic acid. The gene product(s) is(are) other biological macromolecules, such as proteins, not chemically related to the TU. The nucleic acid TU induces the cell to employ its cellular components to produce the specific gene product or products encoded by the nucleic acid of the TU. Although any nucleic acid may serve as a TU, in a preferred embodiment, the TU is the DNA of a plasmid or
20 similar vector, wherein the plasmid or vector comprises coding sequences of marker genes or other sequence constructions that facilitate use of the TU for experimentation and biosynthesis.

 As used herein, a "control sequence" is a regulatory nucleotide sequence
25 incorporated within a TU which interacts with appropriate cellular components of the cell and leads to enhanced or activated biosynthesis of the gene products encoded by the TU. Thus a suitable control sequence is one with which the components of the cell have the capability to interact, resulting in synthesis of the gene product. When operably disposed in a nucleic acid with respect to a specified coding sequence, a

control sequence effectively controls expression of the specified nucleic acid to produce the gene product.

As used herein, a "promoter" is a nucleotide sequence in a TU which serves as a control sequence.

5 As used herein, a "Kozak sequence" or "Kozak consensus sequence" is a nucleotide sequence at the translational start site which optimizes translation of eukaryotic mRNAs (Kozak, *Mol. Cell. Biology* 9: 5134-5142 (1989)).

As used herein, a "terminator" is an extended nucleotide sequence which acts to induce polyadenylation at the 3' end of a mature mRNA. A terminator sequence is
10 found after, or downstream from, a particular coding sequence.

As used herein, a "cell" is a prokaryotic or eukaryotic cell comprising a TU coding for one or more gene products, or into which such a TU has been introduced. Thus, a cell harbors a foreign or heterologous substance, the TU, which is not naturally or endogenously found in the cell as a component. A suitable cell is one which has the
15 capability for the biosynthesis of the gene products as a consequence of the introduction of the TU. In particular, a suitable cell is one which responds to a control sequence and to a terminator sequence, if any, that may be included within the TU. In important embodiments of the present invention, the cell is a mammalian cell. In particularly important embodiments of this invention, the cell is a naturally occurring cell in the
20 body of a human or nonhuman subject to whom (which) the TU has been administered as a component of a vaccine. Alternatively, in analytical, or diagnostic applications, including preparation of antigen for use as a vaccine or in immunodiagnostic assays, or for demonstrative purposes, the cell may be a human or nonhuman cell cultured *in vitro*.

25 As used herein, a "vaccine" or a "composition for vaccinating a subject" specific for a particular pathogen means a preparation, which, when administered to a subject, leads to an immunogenic response in a subject. As used herein, an "immunogenic"

response is one that confers upon the subject protective immunity against the pathogen. Without wishing to be bound by theory, it is believed that an immunogenic response may arise from the generation of neutralizing antibodies (i.e., a humoral immune response) or from cytotoxic cells of the immune system (i.e., a cellular immune response) or both. As used herein, an "immunogenic antigen" is an antigen which induces an immunogenic response when it is introduced into a subject, or when it is synthesized within the cells of a host or a subject. As used herein, an "effective amount" of a vaccine or vaccinating composition is an amount which, when administered to a subject, is sufficient to confer protective immunity upon the subject.

Historically, a vaccine has been understood to contain as an active principle one or more specific molecular components or structures which comprise the pathogen, especially its surface. Such structures may include surface components such as proteins, complex carbohydrates, and/or complex lipids which commonly are found in pathogenic organisms.

As used herein, however, it is to be stressed that the terms "vaccine" or "composition for vaccinating a subject" extend the conventional meaning summarized in the preceding paragraph. As used herein, these terms also relate to the TU of the instant invention or to compositions containing the TU. The TU induces the biosynthesis of one or more specified gene products encoded by the TU within the cells of the subject, wherein the gene products are specified antigens of a pathogen. The biosynthetic antigens then serve as an immunogen. As already noted, the TU, and hence the vaccine, may be any nucleic acid that encodes the specified immunogenic antigens. In a preferred embodiment of this invention, the TU of the vaccine is DNA. The TU can include a plasmid or vector incorporating additional genes or particular sequences for the convenience of the skilled worker in the fields of molecular biology, cell biology and viral immunology (See Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly), which are incorporated herein by reference).

The TU molecules of the instant invention comprise nucleic acids, or derivatives of nucleic acids, having nucleotide sequences that encode specific gene products related to antigens of flaviviruses such as, but not limited to, WNV, JEV, dengue virus, yellow fever virus and SLEV. Although any nucleic acid may serve as a
5 TU, in an important embodiment, the TU is DNA. Alternatively, the nucleic acids may be RNA molecules. They may also be any one of several derivatives of DNA or RNA having a backbone of phosphodiester bonds that have been chemically modified to increase the stability of the TU as a pharmaceutical agent. Modifications so envisioned include, but are not limited to, phosphorothioate derivatives or phosphonate derivatives.
10 These and other examples of derivatives are well known to persons skilled in the field of nucleic acid chemistry.

The genome of JEV has been characterized and sequenced (Figures 1 and 2). The M structural protein is expressed as a portion of the polyprotein which includes a pre-M sequence (pr). This pr sequence, immediately amino terminal to the M protein
15 sequence, prevents conformational problems in the processing of the polyprotein. In particular, the presence of the pr sequence is important in preventing misfolding of the E protein. Thus, the presence of prM allows for assembly of JEV particles. Once the virion or particle is formed, the pr sequence can be cleaved from the prM protein to yield mature virus particles containing M proteins, although cleavage of the prM
20 protein to yield M protein is not necessary to produce infectious particles. The prM sequences from many different, related flaviviruses are cleaved to but a low extent, but the flaviviruses themselves are nonetheless, infectious. Examples of such related flaviviruses with similar genomic structures and functions include, but are not limited to WNV, YFV, dengue virus and SLEV.

25 In one embodiment, the TU encoding flaviviral M and E proteins in the instant invention is DNA. In accord with the discussion in the preceding paragraph, this DNA comprises a nucleotide sequence which encodes the M protein, comprising the pre-M sequence, and a nucleotide sequence encoding the E protein. In this way, the intended gene products are enabled to form subviral particles within the cell. The pre-M

sequence can then be cleaved in a fashion analogous to that which occurs with respect to replete virions.

In order to function effectively *in vivo* as a vaccine, it is advantageous to include within the TU a control sequence that has the effect of enhancing or promoting the transcription of the nucleotide sequences encoding the antigens. Use of such promoters is well known to those of skill in the fields of molecular biology, cell biology and viral immunology (See Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly)). When the TU is used as a vaccine in a mammalian host, the promoter to be employed is preferably one which operates effectively in mammalian cells. Such a promoter is disposed with respect to the coding sequences from which transcription is to be promoted, at a position at which it may operably promote such transcription. In a significant embodiment of the instant invention, this promoter is the cytomegalovirus early promoter. In addition, in a further preferred embodiment of the invention, the coding sequences are followed, in the TU nucleic acid, by a terminator sequence (Sambrook et al.). Particular embodiments of the invention relate to both prokaryotic and eukaryotic cells. Many promoter sequences are known that are useful in either prokaryotic or eukaryotic cells. (See Sambrook et al.)

The nucleic acids of the invention may further include DNA sequences known to those of skill in the art to act as immunostimulatory elements. Examples of such elements include, but are not limited to, certain CpG motifs in bacterial DNA (Sato et al., *Science* 273: 352-354 (1996); Klinman et al., *Vaccine* 17: 19-25 (1998)).

Preparation of the TU of the invention is readily accomplished by methods well known to workers of skill in the field of molecular biology. Procedures involved are set forth, for example, in Molecular Cloning: A Laboratory Manual, 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989 and Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly). The flaviviral RNA molecule may be

isolated from a sample of live virus by methods widely known among virologists familiar with flaviviruses, for example, and with other groups of viruses as well. Methods used with JEV are summarized in Kuno et al. (*J. Virol.* 72: 73-83 (1998)). The RNA is used as a template for the synthesis of cDNA using reverse transcriptase.

5 From the cDNA, a fragment containing the pre-M through E coding region (Figure 2) is obtained by digestion with restriction nucleases known to cleave the cDNA appropriately to provide such fragments. Examples of restriction digestion of JEV are provided in Nitayaphan et al. (1990) and Konishi et al. (1991). Incorporation of promoters, such as the cytomegalovirus promoter, sequences to promote efficient

10 translation, such as the Kozak sequence, and of the polyadenylation signal, is likewise well known to skilled practitioners in molecular biology and recombinant DNA engineering (Kozak, *Mol. Cell. Biology* 9: 5134-5142 (1989); Azevedo et al., *Braz. J. Med. Biol. Res.* 32: 147-153 (1999)). When a nucleic acid comprising a TU containing the desired coding sequences and control sequences is prepared, it may be obtained in

15 larger quantities by methods that amplify nucleic acids. Such methods are widely known to workers skilled in molecular biology and recombinant DNA engineering. Examples of these methods include incorporation of the nucleic acid into a plasmid for replication by culturing in a cell such as a prokaryotic cell and harvesting the plasmid after completing the culture, as well as amplification of the nucleic acid by methods

20 such as PCR and other amplification protocols, as are well known in the art. These examples are not intended to limit the ways in which the nucleic acid containing the TU may be obtained.

The TU-containing nucleic acid molecules of the instant invention may be introduced into appropriate cells in many ways well known to skilled workers in the

25 fields of molecular biology and viral immunology. By way of example, these include, but are not limited to, incorporation into a plasmid or similar nucleic acid vector which is taken up by the cells, or encapsulation within vesicular lipid structures such as liposomes, especially liposomes comprising cationic lipids, or adsorption to particles that are incorporated into the cell by endocytosis.

In general, a cell of this invention is a prokaryotic or eukaryotic cell comprising a TU, or into which a TU has been introduced. The TU of the present invention induces the intracellular biosynthesis of the encoded prM/M and E antigens. A suitable cell is one which has the capability for the biosynthesis of the gene products as a consequence of the introduction of the nucleic acid. In particular embodiments of the invention, a suitable cell is one which responds to a control sequence and to a terminator sequence, if any, which may be included within the TU. In order to respond in this fashion, such a cell contains within it components which interact with a control sequence and with a terminator and act to carry out the respective promoting and terminating functions. When the cell is cultured *in vitro*, it may be a prokaryote, a single-cell eukaryote or a multicellular eukaryote cell. In particular embodiments of the present invention, the cell is a mammalian cell. In these cases, the synthesized prM/M and E protein gene products are available for use in analytical, or diagnostic applications, including preparation of antigen for use as a vaccine or in immunodiagnostic assays, or for demonstrative purposes.

In some circumstances, such as when the cell is a cultured mammalian cell, the prM/M and E antigens are secreted in the form of subviral particles. These are aggregates of prM/M and E proteins resembling live virus in surface ultrastructural morphology and immunogenic properties. Since the TU of the invention does not include the remainder of the flaviviral genome, however, there is no capsid incorporated, and most importantly, no infectious viral RNA.

In another important embodiment of this invention, the cell is a natural cellular component of the subject to whom the TU has been administered as a vaccine. The TU, when administered to the subject, is taken up by the cells of the subject. The subject's cells have the capability of responding to any promoter sequences, and terminator, if present. In any case, the TU induces the subject's cells to synthesize flaviviral prM/M and E gene products. Without wishing to be constrained by theoretical considerations, it is believed that the subject's cells produce subviral particles *in vivo* consisting of the prM/M and E antigens, just as has been found to occur with cultured mammalian cells *in vitro*. Such subviral particles, it is believed,

then serve as the *in vivo* immunogen, stimulating the immune system of the subject to generate immunological responses which confer protective immunity on the subject. Again without wishing to be limited by theory, the resulting protective immunity may arise via either humoral or cellular immunity, i.e., via either an MHC class II- or class I-
5 restricted mechanism, respectively, or by both mechanisms.

According to the invention, subjects are immunized against infection by flaviviruses, such as JEV, YFV, dengue virus, SLEV, WNV or other flaviviruses by administering to them an effective amount of a TU comprising nucleic acid which encodes the prM and/or E antigens. The nucleic acid, after being incorporated into the
10 cells of the subject, leads to the synthesis of the flaviviral prM/M and/or E antigens.

In order to administer the TU to the subject, it is incorporated into a composition which comprises a pharmaceutically acceptable carrier. The term "pharmaceutically acceptable" means a material that is not biologically or otherwise undesirable, i.e., the material may be administered to an subject along with the
15 immunogenic material (i.e., recombinant flavivirus protein antigens or portions thereof) without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the vaccine in which it is contained. Examples of pharmaceutically acceptable carriers, or components thereof, include water, physiological saline and common physiological buffers (for further examples, see
20 Arnon, R. (Ed.) Synthetic Vaccines I: pp. 83-92, CRC Press, Inc., Boca Raton, Florida, 1987).

It is understood by those skilled in the art that the critical value in describing a vaccination dose is the total amount of immunogen needed to elicit a protective
25 response in a host which is subject to infectious disease caused by virulent or wild-type flavivirus infection. The number and volume of doses used can be varied and are determined by the practitioner based on such parameters as, age, weight, gender, species, type of vaccine to be administered, mode of administration, overall condition of the subject, et cetera, as well as other important factors recognized by those of skill
30 in the art.

The TU may be administered to a subject orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally, intranasally, topically or the like. Delivery can also be directly to any area of the respiratory system (e.g., lungs) via intubation. The exact amount of the TU required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the immunogenicity of the vaccine used, the strain or species of flavivirus against which the subject is being immunized, the mode of administration and the like. Thus, it is not possible to specify an exact amount for every embodiment of the present invention. However, an appropriate amount can be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein and what is available in the art.

Parenteral administration of the vaccine of the present invention, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

For solid compositions, conventional nontoxic solid carriers include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talc, cellulose, glucose, sucrose, magnesium carbonate, and the like. Liquid pharmaceutically administrable compositions can, for example, be prepared by dissolving, dispersing, etc. an active compound as described herein and optional pharmaceutical adjuvants in an excipient, such as, for example, water, saline, aqueous dextrose, glycerol, ethanol, and the like, to thereby form a solution or suspension. If desired, the pharmaceutical composition to be administered may also contain minor amounts of nontoxic auxiliary substances such as wetting or emulsifying agents, pH buffering agents and the like, for example, sodium acetate, sorbitan monolaurate, triethanolamine sodium acetate, triethanolamine oleate, etc. Actual methods of preparing such dosage forms are known, or will be apparent, to those skilled in this art;

for example, see Remington's Pharmaceutical Sciences (Martin, E.W. (ed.), latest edition, Mack Publishing Co., Easton, PA).

In one embodiment, the TU of this invention can be administered to the subject by the use of electrotransfer mediated *in vivo* gene delivery, wherein immediately
5 following administration of the TU to the subject, transcutaneous electric pulses are applied to the subject, providing greater efficiency and reproducibility of *in vivo* nucleic acid transfer to tissue in the subject (Mir et al., *Proc. Nat. Acad. Sci USA* 96: 4262-4267 (1999)).

In the methods of the present invention which describe the immunization of a
10 subject by administering a vaccine of this invention to a subject, the efficacy of the immunization can be monitored according the clinical protocols well known in the art for monitoring the immune status of a subject.

An effective amount of a vaccinating composition is readily determined by those of skill in the art to be an amount which, when administered to a subject, confers
15 protective immunity upon the subject. In order to undertake such a determination, the skilled artisan can assess the ability to induce flaviviral prM/M- and E-specific antibodies and/or flaviviral prM/M- and E-specific cytotoxic T lymphocytes present in the blood of a subject to whom the vaccine has been administered. One can also determine the level of protective immunity conferred upon an experimental subject by
20 challenge with live flavivirus corresponding to the antigenic composition used to immunize the experimental subject. Such challenge experiments are well known to those of skill in the art.

In general, in order to immunize a subject against infection by WNV, JEV, YFV, dengue virus, SLEV, or other flaviviruses according to the present invention, and
25 recognizing that the TUs employed in such methods may have differing overall sizes, doses ranging from about 0.1 $\mu\text{g/kg}$ body weight to about 50 $\mu\text{g/kg}$ body weight can be used.

It has unexpectedly been found that a TU of the present invention which is a DNA confers protective immunity at a level of effectiveness approximating 100% after administration of only a single effective dose of the TU by i.m. injection or by electrotransfer. This is in contrast to many immunization methods carried out using
5 conventional vaccines (as described above), which require one or more booster vaccinations and which may not confer protective immunity to an effectiveness near 100%.

It has further been found unexpectedly that protective immunity may be transmitted from a vaccinated female subject to the offspring of the subject. A
10 significant proportion of neonatal mice was shown to be protected against viral challenge after the mothers were vaccinated using the TU DNA of the invention. Without wishing to be limited by theory, it is known that passive immunity may be conferred on neonatal mammals due to the presence in maternal milk of neutralizing antibodies specific for various pathogens. It is possible that the protective immunity
15 against JEV found within the neonates was transmitted to them in this way.

In another embodiment of the invention, the TU encodes a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus. Thus, in one embodiment, for example, the signal sequence of structural protein of a first flavivirus is replaced by a signal sequence of structural
20 protein of a second flavivirus, which results in proper folding of the nascent polypeptide, proper processing in a host, and/or proper folding of the processed protein.

In another embodiment of the invention, the TU may encode an immunogenic flavivirus antigen wherein the antigen comprises sequence from one or more than one flavivirus. The signal sequence can be an improved signal peptide. Improvement of
25 signal sequences, or selection of more optimal signal sequences, can be accomplished by application of the principles and techniques taught in Example 18 and references cited therein, each of which are incorporated herein by reference for the express teachings in each related to the selection, identification and design of signal sequences

with desired properties and functions. Generally, these desired properties and functions will include a high signal sequence probability.

In another embodiment of the invention, more than one TU or a TU encoding an immunogenic flavivirus antigen from more than one flavivirus are included in a single composition. Thus, in one embodiment, for example, a TU can encode a nascent polypeptide or polypeptides that are processed into proteins from more than one flavivirus. Preferably, the processed proteins form subviral particles that elicit an immunological response against the proteins. The subviral particles can be formed from processed proteins derived from the sequence of the same flavivirus, a combination of flaviviruses, or chimeric flavivirus proteins. Combination vaccines, comprising more than one TU or a TU encoding an immunogenic flavivirus antigen from more than one flavivirus can be tailored for use in particular geographical regions by inclusion of proteins from flaviviruses endemic to the region or otherwise likely to be encountered. For example, a vaccine for tropical and subtropical Asia can include TU(s) that encode proteins from the four serotypes of DEN, WN and JE virus vaccines. Similarly useful vaccines for Africa and Latin America could include TU(s) that encode proteins from the four serotypes of DEN, WN and YF viruses and the four serotypes of DEN, Rocio and YF viruses, respectively.

In another embodiment, the TU encodes a signal sequence of a structural protein of a first flavivirus and an immunogenic chimeric flavivirus antigen that includes amino acid sequence from more than one flavivirus. The signal sequence can be a Japanese encephalitis virus signal sequence. The chimeric flavivirus antigen can include sequence from a Japanese encephalitis virus antigen. In certain embodiments, the chimeric antigen is an E protein. The carboxy terminal portion of the E protein can be E protein sequence from the Japanese encephalitis virus. The carboxy terminal portion can be, for example, 5, 10, 15, 20, 25, 30, 40, 50 or 75% of the chimeric E protein. In a preferred embodiment, the TU encodes a signal sequence of a structural protein of Japanese encephalitis virus, a prM protein of a Dengue virus and a chimeric E protein containing sequence from both the Japanese encephalitis virus and the Dengue virus. The chimeric protein can be an E protein wherein the carboxy terminal portion

comprises Japanese encephalitis virus sequence. Examples of TUs include nucleic acid sequences shown in SEQ ID NO:44 and SEQ ID NO:46 that can direct the synthesis of flavivirus antigens such as those shown in SEQ ID NO:45 and SEQ ID NO:47.

5 The present invention further provides immunogenic compositions comprising the polypeptides of this invention in a pharmaceutical acceptable carrier for use as a protein vaccine. Antigens produced from the transcriptional units of the present invention can be used to elicit effective immune responses in a subject. Antigens for this purpose can comprise flavivirus prM protein, flavivirus M protein, flavivirus E
10 protein or any combination thereof, including immunogenic fragments of the proteins. A particularly preferred embodiment is the use of the NRA described herein. A further preferred embodiment is a chimeric protein comprising the signal sequence of one flavivirus and the structural protein(s) of one or more different flaviviruses. In a particularly preferred embodiment, the signal sequence of the antigen is the Japanese
15 encephalitis virus signal sequence. In other preferred embodiments, the signal sequence is an improved signal peptide. Improvement of signal sequences, or selection of more optimal signal sequences, can be accomplished by application of the principles and techniques taught in Example 18 and references cited therein each of which are incorporated herein by reference for the express teachings in each related to
20 the selection, identification and design of signal sequences with desired properties and function. Generally, these desired properties and function will include a high signal sequence probability.

In other embodiments, the protein vaccine of this invention further comprises a suitable adjuvant. As used herein, an "adjuvant" is a potentiator or enhancer of the
25 immune response. The term "suitable" is meant to include any substance which can be used in combination with the vaccine immunogen (i.e., flavivirus prM protein, flavivirus M protein, flavivirus E protein, or any combination thereof) to augment the immune response, without producing adverse reactions in the vaccinated subject. Effective amounts of a specific adjuvant may be readily determined so as to optimize
30 the potentiation effect of the adjuvant on the immune response of a vaccinated subject. In a preferred embodiment, adjuvanting of the vaccines of this invention is a 2 – stage

process, utilizing first a 2% aluminum hydroxide solution and then a mineral oil. In specific embodiments, suitable adjuvants can be chosen from the following group: mineral, vegetable or fish oil with water emulsions, incomplete Freund's adjuvant, *E. coli* J5, dextran sulfate, iron oxide, sodium alginate, Bacto-Adjuvant, certain synthetic
5 polymers such as Carbopol (BF Goodrich Company, Cleveland, Ohio), poly-amino acids and co-polymers of amino acids, saponin, carrageenan, REGRESSIN (Vetrepharm, Athens, GA), AVRIDINE (N, N-dioctadecyl-N',N'-bis(2-hydroxyethyl)-propanediamine), long chain polydispersed β (1,4) linked mannan polymers interspersed with O-acetylated groups (e.g. ACEMANNAN), deproteinized highly
10 purified cell wall extracts derived from non-pathogenic strain of Mycobacterium species (e.g. EQUIMUNE, Vetrepharm Research Inc., Athens GA), Mannite monooleate, paraffin oil and muramyl dipeptide.

In another aspect, this invention provides a method for immunizing subjects
15 with immunogenic amounts of the protein vaccine of the invention to elicit an effective immune response in the subject. Immunization can be carried out orally, parenterally, intranasally, intratracheally, intramuscularly, intramammarily, subcutaneously, intravenously and/or intradermally. The vaccine containing the flavivirus prM protein, flavivirus M protein and/or the flavivirus E protein can be administered by injection, by
20 inhalation, by ingestion, or by infusion. A single dose can be given and/or repeated doses of the vaccine preparations, i.e. "boosters," can be administered at periodic time intervals to enhance the initial immune response or after a long period of time since the last dose. The time interval between vaccinations can vary, depending on the age and condition of the subject.

25 The term "immunogenic amount" means an amount of an immunogen, or a portion thereof, which is sufficient to induce an immune response in a vaccinated subject and which protects the subject against disease caused by wild-type or virulent flavivirus infections upon exposure thereto or which has a therapeutic or commercially beneficial effect that lessens the effect of flavivirus infection on the vaccinated subject.

The invention further provides an antibody produced in response to immunization by the antigen of this invention. The antibodies of the present invention can include polyclonal and monoclonal antibodies which can be intact immunoglobulin molecules, chimeric immunoglobulin molecules, "humanized antibodies," or Fab or F(ab')₂ fragments. Such antibodies and antibody fragments can be produced by techniques well known in the art which include those described in Harlow and Lane (Antibodies: A Laboratory Manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989) and Kohler et al. (*Nature* 256:495-97, 1975) and U.S. Patents 5,545,806, 5,569,825 and 5,625,126, incorporated herein by reference. The antibodies can be of any isotype IgG, IgA, IgD, IgE and IgM.

The present invention can also include single chain antibodies (ScFv), comprising linked V_H and V_L domains and which retain the conformation and specific binding activity of the native idiotype of the antibody. Such single chain antibodies are well known in the art and can be produced by standard methods. (see, e.g., Alvarez et al., *Hum. Gene Ther.* 8: 229-242 (1997)).

Antibodies can be produced against the antigens of this invention which are synthesized from nucleic acid sequences encoding immunogenic amino acid sequences of the prM, M and/or E antigens of one or more flaviviruses and the signal sequence of a different flavivirus (e.g., JEV). Immunogenic peptides synthesized from the use of these chimeric constructs can easily be identified by use of methods well known in the art for identifying immunogenic regions in an amino acid sequence and used to produce the antibodies of this invention.

Conditions whereby an antigen/antibody complex can form, as well as assays for the detection of the formation of an antigen/antibody complex and quantitation of the detected protein; are standard in the art. Such assays can include, but are not limited to, Western blotting, immunoprecipitation, immunofluorescence, immunocytochemistry, immunohistochemistry, fluorescence activated cell sorting (FACS), fluorescence *in situ* hybridization (FISH), immunomagnetic assays, ELISA, ELISPOT (Coligan et al., eds. 1995. Current Protocols in Immunology. Wiley, New

York.), agglutination assays, flocculation assays, cell panning, etc., as are well known to the artisan.

As used herein, the term "bind" means the well characterized binding of antibody to antigen as well as other nonrandom association with an antigen.

- 5 "Specifically bind" as used herein describes an antibody or other ligand that does not cross react substantially with any antigen other than the one specified, which in this case, is an antigen of this invention.

The antibody or ligand of this invention can be bound to a substrate (e.g., beads, tubes, slides, plates, nitrocellulose sheets, etc.) or conjugated with a detectable moiety or both bound and conjugated. The detectable moieties contemplated for the present invention can include, but are not limited to, an immunofluorescent moiety (e.g., fluorescein, rhodamine), a radioactive moiety (e.g., ^{32}P , ^{125}I , ^{35}S), an enzyme moiety (e.g., horseradish peroxidase, alkaline phosphatase), a colloidal gold moiety and a biotin moiety. Such conjugation techniques are standard in the art (for example, Harlow and Lane, Antibodies: A Laboratory Manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989); Yang et al., *Nature* 382: 319-324 (1996)).

The present invention further provides a method of detecting flavivirus antibody in a sample, comprising contacting the sample with the flavivirus antigen of the present invention, under conditions whereby an antigen/antibody complex can form; and
20 detecting formation of the complex, thereby detecting flavivirus antibody in the sample.

The present invention further provides a method of detecting flavivirus antigen in a sample, comprising contacting the sample with an antibody of this invention under conditions whereby an antigen/antibody complex can form; and detecting formation of the complex, thereby detecting flavivirus antigen in the sample.

25 The method of detecting flavivirus antigen in a sample can be performed, for example, by contacting a fluid or tissue sample from a subject with an antibody of this invention and detecting binding of the antibody to the antigen. It is contemplated that

the antigen will be on an intact flavivirus virion, will be a flavivirus-encoded protein displayed on the surface of a flavivirus-infected cell expressing the antigen, or will be a fragment of the antigen. A fluid sample of this method can comprise any biological fluid which could contain the antigen or a cell containing the antigen, such as
5 cerebrospinal fluid, blood, bile, plasma, serum, saliva and urine. Other possible examples of body fluids include sputum, mucus and the like.

The method of detecting flavivirus antibody in a sample can be performed, for example, by contacting a fluid or tissue sample from a subject with an antigen of this invention and detecting the binding of the antigen to the antibody. A fluid sample of
10 this method can comprise any biological fluid which could contain the antibody, such as cerebrospinal fluid, blood, bile, plasma, serum, saliva and urine. Other possible examples of body fluids include sputum, mucus and the like.

Enzyme immunoassays such as immunofluorescence assays (IFA), enzyme
15 linked immunosorbent assays (ELISA) and immunoblotting can be readily adapted to accomplish the detection of flavivirus antibodies according to the methods of this invention. An ELISA method effective for the detection of the antibodies can, for example, be as follows: (1) bind the antigen to a substrate; (2) contact the bound antigen with a fluid or tissue sample containing the antibody; (3) contact the above with
20 a secondary antibody bound to a detectable moiety which is reactive with the bound antibody (e.g., horseradish peroxidase enzyme or alkaline phosphatase enzyme); (4) contact the above with the substrate for the enzyme; (5) contact the above with a color reagent; and (6) observe/measure color change or development.

Another immunologic technique that can be useful in the detection of flavivirus
25 antibodies uses monoclonal antibodies (MAbs) for detection of antibodies specifically reactive with flavivirus antigens in a competitive inhibition assay. Briefly, sample is contacted with an antigen of this invention which is bound to a substrate (e.g., an ELISA 96-well plate). Excess sample is thoroughly washed away. A labeled (e.g., enzyme-linked, fluorescent, radioactive, etc.) monoclonal antibody is then contacted
30 with any previously formed antigen-antibody complexes and the amount of monoclonal

antibody binding is measured. The amount of inhibition of monoclonal antibody binding is measured relative to a control (no antibody), allowing for detection and measurement of antibody in the sample. The degree of monoclonal antibody inhibition can be a very specific assay for detecting a particular flavivirus variety or strain, when
5 based on monoclonal antibody binding specificity for a particular variety or strain of flavivirus. MAbs can also be used for direct detection of flavivirus antigens in cells by, for example, immunofluorescence assay (IFA) according to standard methods.

As a further example, a micro-agglutination test can be used to detect the presence of flavivirus antibodies in a sample. Briefly, latex beads, red blood cells or
10 other agglutinable particles are coated with the antigen of this invention and mixed with a sample, such that antibodies in the sample that are specifically reactive with the antigen crosslink with the antigen, causing agglutination. The agglutinated antigen-antibody complexes form a precipitate, visible with the naked eye or measurable by spectrophotometer. In a modification of the above test, antibodies of this invention can
15 be bound to the agglutinable particles and antigen in the sample thereby detected.

The present invention further provides a method of diagnosing a flavivirus infection in a subject, comprising contacting a sample from the subject with the antigen of this invention under conditions whereby an antigen/antibody complex can form; and detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection
20 in a subject.

The present invention further provides a method of diagnosing a flavivirus infection in a subject, comprising contacting a sample from the subject with the antibody of this invention under conditions whereby an antigen/antibody complex can form; and detecting antigen/antibody complex formation, thereby diagnosing a
25 flavivirus infection in a subject.

In the diagnostic methods taught herein, the antigen of this invention can be bound to a substrate and contacted with a fluid sample such as blood, serum, urine or saliva. This sample can be taken directly from the patient or in a partially purified

form. In this manner, antibodies specific for the antigen (the primary antibody) will specifically react with the bound antigen. Thereafter, a secondary antibody bound to, or labeled with, a detectable moiety can be added to enhance the detection of the primary antibody. Generally, the secondary antibody or other ligand, which is reactive, either
5 specifically with a different epitope of the antigen or nonspecifically with the ligand or reacted antibody, will be selected for its ability to react with multiple sites on the primary antibody. Thus, for example, several molecules of the secondary antibody can react with each primary antibody, making the primary antibody more detectable.

The detectable moiety allows for visual detection of a precipitate or a color
10 change, visual detection by microscopy, or automated detection by spectrometry, radiometric measurement or the like. Examples of detectable moieties include fluorescein and rhodamine (for fluorescence microscopy), horseradish peroxidase (for either light or electron microscopy and biochemical detection), biotin-streptavidin (for light or electron microscopy) and alkaline phosphatase (for biochemical detection by
15 color change).

Particular embodiments of the present invention are set forth in the examples which follow. These examples are not intended to limit the scope of the invention as disclosed in this specification.

Examples

20 General methods utilizing molecular biology and recombinant DNA techniques related to preparing and expressing the nucleic acid TU molecules of the invention are set forth in, for example, Current Protocols in Molecular Biology, Ausubel et al., John Wiley and Sons, New York 1987 (updated quarterly), and Molecular Cloning: A Laboratory Manual 2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor
25 Laboratory, Cold Spring Harbor, NY, 1989.

Example 1. Preparation of recombinant plasmids containing the transcriptional unit encoding JEV prM and E antigens. Genomic RNA was extracted from 150 μ L of JEV strain SA 14 virus seed grown from mouse brain using a QIAamp™ Viral RNA

Kit (Qiagen, Santa Clarita, CA). RNA, adsorbed on a silica membrane, was eluted in 80 μ L of nuclease-free water, and used as a template for the amplification of JEV prM and E gene coding sequences. Primer sequences were obtained from the work of Nitayaphan et al. (*Virology* 177: 541-552 (1990)). A single cDNA fragment containing the genomic nucleotide region 389-2478 was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction sites KpnI and XbaI, the consensus Kozak ribosomal binding sequence, and the translation initiation site were engineered at the 5' terminus of the cDNA by amplicon 14DV389 (nucleotide sequence, SEQ ID NO:1; amino acid sequence, SEQ ID NO:2). An in-frame translation termination codon, followed by a NotI restriction site, was introduced at the 3' terminus of the cDNA by amplicon c14DV2453 (SEQ ID NO:3) (Figure 2). One-tube RT-PCR was performed using a Titan RT-PCR Kit (Boehringer Mannheim, Indianapolis, IN). 10 μ L of viral RNA was mixed with 1 μ L each of 14DV389 (50 μ M) and c14DV2453 (50 μ M) and 18 μ L of nuclease-free water and the mixture was heated at 85°C for 5 min and then cooled to 4°C. 75 μ L of reaction mix [20 μ L 5x buffer, 2 μ L of dNTP mixture (10 mM each), 5 μ L of dithiothreitol (0.1 mM), 0.5 μ L of RNasin™ (40 U/ μ L, 15
Boehringer Mannheim), 2 μ L of polymerase mixture, and 45.5 μ L of nuclease-free water] was added and RT-PCR performed as follows: 1 cycle (50°C for 30 min, 94°C for 3 min, 50°C for 30 s, 68°C for 2.5 min), 9 cycles (94°C for 30 s, 50°C for 30 s, 68°C for 2.5 min), 20 cycles (94°C for 30 s, 50°C for 30 s, 68°C for 2.5 min in the first cycle, with an increment of 5 s per cycle thereafter), and a final extension at 68°C for 15 min. The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen) and eluted with 50 μ L of 1 mM Tris-HCl, pH 7.5.

All vector constructions and analyses were carried out by using standard techniques (Sambrook et al., 1989). RT-PCR amplified cDNA, digested with KpnI and NotI nucleases, was inserted into the KpnI-NotI site of eukaryotic expression plasmid vector (pCDNA3, Invitrogen, Carlsbad, CA). Electroporation-competent *Escherichia coli* XL1-Blue cells (Stratagene, La Jolla, CA) were transformed by electroporation (Gene Pulser™, Bio-Rad, Hercules, CA) and plated onto LB agar plates containing 100 μ g/mL carbenicillin (Sigma Chemical Co., St. Louis, MO). Clones were picked and inoculated into 3 mL LB broth containing 100 μ g/mL carbenicillin. Plasmid DNA was

extracted from a 14 h culture using a QIAprep™ Spin Miniprep Kit (Qiagen). Automated DNA sequencing was performed as recommended (Applied Biosystems/Perkin Elmer, Foster City, CA). Both strands of the cDNA were sequenced and shown to be identical to the sequence for the original SA14 strain (Nitayaphan et al., 1990).

The fragment of plasmid pCDNA3 (Invitrogen, Carlsbad, CA) from nucleotide (nt) 1289 to nt 3455, containing f1 ori, SV40 ori, the neomycin resistance gene, and SV40 poly(A) elements was deleted by PvuII digestion and then ligated to generate the pCBamp plasmid. The vector pCIBamp, containing a chimeric intron insertion at the NcoI/KpnI site of the pCBamp was constructed by excising the intron sequence from pCI (Promega, Madison, WI) by digestion with NcoI and KpnI. The resulting 566-bp fragment was cloned into pCBamp by digesting with NcoI-KpnI to replace its 289-bp fragment. Figure 3 presents the relationships between the plasmids pCDA3, pCBamp, and pCIBamp.

Plasmids containing the transcriptional unit encoding JEV prM and E proteins were prepared from these plasmids. The cDNA fragment containing the JEV prM and E coding regions in the recombinant plasmid pCDJE2-7 (nucleotide sequence, SEQ ID NO:10; amino acid sequence, SEQ ID NO:11), derived from the pCDNA3 vector, was excised by digestion with NotI and KpnI or XbaI and cloned into the KpnI-NotI site of pCBamp, pCIBamp, pCEP4 (Invitrogen, Carlsbad, CA), or pREP4 (Invitrogen, Carlsbad, CA), or into the SpeI-NotI site of pRc/RSV (Invitrogen, Carlsbad, CA) expression vector to create pCBJE1-14 (nucleotide sequence, SEQ ID NO:17; amino acid sequence, SEQ ID NO:18), pCIBJES14, pCEJE, pREFE, and pRCJE, respectively. Both strands of the cDNA from clones of each plasmid were sequenced and recombinant clones with the correct nucleotide sequence were identified. Plasmid DNA for use in the in vitro transformation of mammalian cells or mouse immunization experiments was purified by anion exchange chromatography using an EndoFree™ Plasmid Maxi Kit (Qiagen).

Example 2. Evaluation of JEV prM and E proteins expressed by various recombinant plasmids using an indirect immunofluorescent antibody assay. The expression of JEV specific gene products by the various recombinant expression plasmids was evaluated in transiently transfected cell lines of COS-1, COS-7 and SV-T2 (ATCC, Rockville MD; 1650-CRL, 1651-CRL, and 163.1-CCL, respectively) by indirect immunofluorescent antibody assay (IFA). The SV-T2 cell line was excluded from further testing since a preliminary result showed only 1-2% of transformed SV-T2 cells were JEV antigen positive. For transformation, cells were grown to 75% confluence in 150 cm² culture flasks, trypsinized, and resuspended at 4°C in phosphate buffered saline (PBS) to a final cell count 5×10^6 per mL. 10 µg of plasmid DNA was electroporated into 300 µL of cell suspension using a BioRad Gene Pulse™ (Bio-Rad) set at 150 V, 960 µF and 100 Ω resistance. Five minutes after electroporation, cells were diluted with 25 mL fresh medium and seeded into a 75 cm² flask. 48 h after transformation the medium was removed from the cells, and the cells were trypsinized and resuspended in 5 mL PBS with 3% normal goat serum. 10 µL aliquots were spotted onto slides, air dried and fixed with acetone at -20°C for 20 min. IFA was performed with acetone-fixed plasmid-transformed cells using fluorescein isothiocyanate-conjugated goat anti-mouse immunoglobulin G (Sigma Chemical Co.) and JEV HIAF.

To determine the influence of various promoter and poly(A) elements on the JEV prM and E protein expression, COS-1 and COS-7 cell lines were transiently transformed by an equal amount of pCDJE2-7 (SEQ ID NO:10), pCEJE, pREJE, or pRCJE plasmid DNA. JEV antigens were expressed in both cell lines transformed by all four recombinant plasmids, thus confirming that the CMV or RSV (rouv sarcoma virus) promoter and BGH or SV40 poly(A) elements were functionally active. However, the percentage of transformed cells and the level of JEV antigens expressed, as determined by the number of IFA positive cells and IFA intensity, respectively, differed greatly among the various plasmids (Table 1). A significantly high percentage of COS-1 cells transformed by pCDJE2-7 (SEQ ID NO:10), pCBE1-14 (SEQ ID NO:17) and pCIBJES14 expressed the JEV antigens, and the level of the expressed proteins was compatible with JEV-infected cells. Cells transfected with pCEJE,

pREJE, or pRCJE vectors, on the other hand, had a low percentage of antigen-expressing cells, as well as a low intensity of fluorescence, indicating weak expression of the antigens.

In order to ascertain whether the enhanced expression of JEV proteins by pCDJE2-7 (SEQ ID NO:10) was influenced by the SV40-encoded eukaryotic origin of replication, the plasmid pCBE1-14 (SEQ ID NO:17) was constructed so that a 2166-bp fragment, containing f1 ori, SV40 ori, the neomycin resistance gene and SV40 poly(a) elements from pCDJE2-7, was deleted. A chimeric intron was then inserted into pCBE1-14 to generate pCIBJES14. The pCIBJES14 plasmid was used to determine if the expression of JEV proteins could be enhanced by the intron sequence. Following transformation, cells harboring both pCBE1-14 and pCIBJES14 vectors expressed a level of JEV antigens similar to that observed with pCDJE2-7 (Table 1). This result indicates that expression of JEV prM and E antigens by recombinant vectors is influenced only by the transcriptional regulatory elements. Neither the eukaryotic origin of replication nor the intron sequence enhanced JEV antigen expression in the cells used. Vectors containing the CMV promoter and BGH poly(A) (Figure 3) were selected for further analysis.

Example 3. Selection of an in vitro transformed, stable cell line constitutively expressing JEV specific gene products. COS-1 cells were transformed with 10 μ g of pCDJE2-7 DNA by electroporation as described in the previous example. After a 24 hr incubation in non-selective culture medium, cells were treated with neomycin (0.5 mg/mL, Sigma Chemical Co.). Neomycin-resistant colonies, which became visible after 2-3 weeks, were cloned by limited dilution in neomycin-containing medium. Expression of vector-encoded JEV gene products was initially screened by IFA using JEV HIAF. One JEV-IFA positive clone (JE-4B) and one negative clone (JE-5A) were selected for further analysis and maintained in medium containing 200 μ g/mL neomycin.

Authenticity of the JEV E protein expressed by the JE-4B clone was demonstrated by epitope mapping by IFA using a panel of JEV E-specific murine

monoclonal antibodies (Mab) (Kimura-Kuroda et al., *J. Virol.* 45: 124-132 (1983); Kimura-Kuroda et al., *J. Gen. Virol.* 67: 2663-2672 (1986); Zhang et al., *J. Med. Virol.* 29: 133-138 (1989); and Roehrig et al., *Virol.* 128: 118-126 (1983)). JEV HIAF and normal mouse serum were used as positive and negative antibody controls, respectively.

- 5 Four JEV-specific, six flavivirus-subgroup specific, and two flavivirus-group reactive Mabs reacted similarly with the 4B clone or JEV-infected COS-1 cells (Table 2).

Example 4. Antigenic properties and immunological detection of subviral particles secreted by the JE-4B COS-1 cell line.

- a. Preparation of subviral particles. JE-4B COS-1 cells were grown and
- 10 maintained in medium containing 200 μ g/mL of neomycin. The cultured medium was routinely harvested and stored at 4°C, and replenished twice weekly, and the cells were split 1:5 every 7-10 days. Culture medium was clarified by centrifugation at 10,000 rpm for 30 min in a Sorvall F16/250 rotor at 4°C, and centrifuged further for 4 hr at 39,000 rpm in a Sorvall TH641 rotor at 4°C through a 5% sucrose cushion (w/w,
- 15 prepared with 10 mM Tris HCl, pH 7.5, 100 mM NaCl (TN buffer)). The pellet containing subviral particles was resuspended in TN buffer and stored at 4°C. Alternatively, 7% or 10% PEG-8000 (w/v) was added to the clarified culture medium. The mixture was stirred at 4°C for at least 2 hr, and the precipitated particles were collected by centrifugation at 10,000 rpm for 30 min. The precipitate was resuspended
- 20 in TN buffer and stored at 4°C. The subviral particles were purified from both pelleted and PEG-precipitated preparations by rate zonal centrifugation in a 5-25% continuous sucrose gradient in TN at 38,000 rpm at 4°C for 90 min. 1-mL fractions were collected from the top of the gradient, tested by antigen capture ELISA (see below), and the positive fractions loaded onto a 25-50% sucrose gradient in TN. This was centrifuged
- 25 overnight in an equilibrium density centrifugation at 35,000 rpm at 4°C. 0.9-mL fractions from the equilibrium gradients were collected from the bottom. They were tested by antigen-capture ELISA and assessed for hemagglutination (HA) activity at pH 6.6. An aliquot of 100 μ L of each fraction was weighed precisely to determine its density. The ELISA-positive fractions were pooled and pelleted at 39,000 rpm at 4°C
- 30 for 3-4 hr and the pellet resuspended in TN buffer. Antigen-capture ELISA and HA

titers were determined on the pelleted samples. JEV-infected COS-1 cell supernatant was also subjected to similar purification protocols as detailed above and used as a positive control for the gradient analysis. JE virions were also purified from infected C6/36 cells 5-6 days postinfection by sedimentation in a glycerol/tartrate equilibrium
5 gradient.

b. Western blots of subviral particles. Gradient-purified samples of the subviral particles were mixed with electrophoresis sample buffer and run on 10 or 12.5% sodium dodecyl sulfate-containing polyacrylamide gels (SDS-PAGE) as described by Laemmli (*Nature* 277: 680-685 (1970)). Proteins were transferred to a nitrocellulose
10 membrane and immunochemically detected with polyclonal JEV HIAF, flavivirus cross-reactive anti-E Mab 4G2 (Henchal et al., *Amer. J. Trop. Med. Hyg.* 31: 830-836 (1982)), or mouse anti-prM peptide hyperimmune serum (JM01). Figure 4 shows a comparison of the M and E proteins produced by JEV infected C6/36 and JE-4B COS-1 cells. Some nonspecific reactivity to E protein was observed in the normal mouse
15 ascitic fluid and Jm01 anti-peptide serum. Proteins identical in size to M and E were secreted in the subviral particles and could be detected by E-specific Mab 4G2 and prM-specific JM01 antiserum, respectively.

c. Density gradient detection of JEV subviral particles in culture medium. For ELISA, antigen-capture antibody (4G2) was diluted in 0.1 M sodium carbonate buffer, pH 9.6, and used to coat 96-well microtiter plates (Immulon II, Dynatech, Chantilly,
20 VA) by overnight incubation at 4° C. After blocking with 3% normal goat serum in PBS, two-fold serially-diluted samples were added to the 4G2-coated plate and incubated 1.5 hours at 37°C. Captured antigen was detected by horseradish peroxidase-conjugated 6B6C-1 Mag, and incubated for 1 hour at 37°C. The enzyme activity on the
25 solid phase was then detected with TMB (3,3',5,5'-tetramethylbenzidine)-ELISA (Life Technologies, Grand Island, NY).

Approximately 500 mL of cell culture medium from 15 X 150 cm² flasks of JE-4B cells was collected four days after cells were seeded. PEG-precipitated subviral particles were resuspended in 2 mL of TN buffer, pH 7.5; a 0.7 mL aliquot of this

resuspended pellet was loaded onto a 5-25% sucrose gradient. Triton X-100, which disrupts subviral particles, was added to another 0.7 mL aliquot to a final concentration of 0.1% and this was loaded onto a 5-25% sucrose gradient prepared in TN buffer containing 0.1% Triton X-100. A definite opaque band was observed approximately 2.5 cm from the top of the gradient containing Triton X-100, but not in the gradient without detergent. Fractions (1 mL) were collected from top to bottom for each gradient (Figure 5). Each collected fraction was analyzed by antigen capture ELISA. Antigen was detected in fractions 4-6, indicating relatively rapid sedimentation characteristic of subviral particles. Treatment of the PEG precipitate from JE-4B culture medium with Triton X-100 shifted the position of ELISA-reactive material to the top of the gradient. Thus treatment with Triton X-100 produces only slow-sedimenting molecules. A similar finding was reported by Konishi et al. (*Virology* 188: 714-720 (1992)). These results show that rapidly sedimenting subviral particles containing prM/M and E could be disrupted by detergent treatment.

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Hemagglutination (HA) activity was determined in the pH range from 6.1 to 7.0 by the method of Clarke and Casals (*Amer. J. Trop. Med. Hyg.* 7: 561-573 (1958)). The subviral particle secreted by JE-4B cells and the virion particle produced by JEV infected COS-1 cells had a similar HA profile with the optimum pH determined to be 6.6.

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Example 5. Comparison of the immune response in mice vaccinated with pCDJE2-7 nucleic acid vaccine of the invention and commercial JEV vaccine. Groups of five 3-week-old female, ICR outbred mice were injected intramuscularly in the left and right quadriceps with 100 μ g of pCDJE2-7 plasmid in 100 μ L of dH₂O or were given doses of JE-VAX (manufactured by the Research Foundation for Microbial Disease of Osaka University and distributed by Connaught Laboratories, Swiftwater, PA.) subcutaneously that are one-fifth the dose given to humans. The plasmid pCDNA3/CAT (Invitrogen), which encodes and expresses an unrelated protein, was used as the negative vaccination control. Except for one group of pCDJE2-7-vaccinated mice, all animals were boosted 3 weeks later with an additional dose of plasmid or JE-VAX. Mice were bled from the retroorbital sinus at 3, 6, 9, 23, 40 and

25

30

60 weeks after inoculation. JEV antibody titers were determined by enzyme-linked immunosorbent assay (ELISA) against purified JEV or by plaque reduction neutralization tests (PRNT) (Roehrig et al., *Virology* 171: 49-60 (1989); and Hunt and Calisher, *Amer. J. Trop. Med. Hyg.* 28: 740-749 (1979)).

5 The pCDJE2-7 nucleic acid vaccine and JE-VAX provided 100% seroconversion within three weeks after the first vaccination in all three groups of mice (Table 3). The JEV ELISA and PRNT antibody titers reached the highest level at week 6 and week 9, respectively, after immunization. Mice receiving 1 dose of DNA vaccine had similar antibody responses as those receiving 2 doses. Comparable ELISA
10 antibody titers were maintained in DNA-vaccinated groups up to 60 weeks, after which the experiment was terminated. However, only one of four mice in the JE-VAX group was JEV antibody positive at 60 weeks post-inoculation. The pCDNA3/CAT control group did not have any measurable JEV antibody. These results demonstrate that a single dose of JEV-specific nucleic acid vaccine is more effective in maintaining JEV
15 antibody in mice than the commercial, FDA-approved JE-VAX vaccine.

Example 6. Comparison of various nucleic acid vaccine constructs of the invention and commercial JEV vaccine for effectiveness of vaccination at different ages. A similar level of JEV protein was expressed by COS-1 cells transformed by either pCDJE2-7, pCBE1-14, or pCIBJES14. JEV antibody induction by these nucleic
20 acid constructs was compared to JE-VAX commercial vaccine at two different ages at vaccination. Three-day (mixed sex) or 3-week-old (female) ICR outbred mice, 10 per group, were vaccinated intramuscularly with 50 or 100 μ g of plasmid DNA, or subcutaneously with doses of JE-VAX that are one-tenth or one-fifth the dose given to humans. Serum specimens were collected at 3 and 7 weeks after immunization and
25 tested at a 1:1600 dilution by ELISA using purified JEV as an antigen. Results are shown in Table 4.

Plasmid pCBE1-14 provided the highest extent of seroconversion, i.e., antibody titer greater than 1:1600, achieving 80-100% at both ages of vaccination. Administration of pCDJE2-7 or pCIBJES14 provided moderate seroconversion by 7

weeks when 3-day old mice were vaccinated (60% for each), but weaker seroconversion (40% and 10%, respectively) when measured 3 weeks after vaccination. When these plasmids were administered at the age of 3 weeks, however, seroconversions of 90% or 100% were attained at both 3 weeks and 7 weeks after
5 vaccination. In contrast, the commercial vaccine, JE-VAX, conferred no seroconversion when administered at 3 days of age, and 100% when given at 3 weeks of age. Thus the nucleic acid TU's for JEV prM and E provided an extent of seroconversion better than a very high dose of the commercial vaccine, and unexpectedly high seroconversion in both young and more mature animals.

10 Example 7. Protective immunity conferred by the nucleic acid vaccine of the invention. Three-day old vaccinated groups from Example 6 were challenged 7 weeks after vaccination by intraperitoneal injection of 50,000 pfu/100 μ L of the mouse-adapted JEV strain SA14 and observed for 3 weeks. 100% protection was achieved in groups that received various nucleic acid TU-containing vaccine constructs for up to 21
15 days (Table 5). In contrast, 60% of the JE-VAX-vaccinated mice, as well as 70% of the pCDNA3/CAT-vaccinated negative controls, did not survive virus challenge by 21 days. These results indicate that the nucleic acid TU's of the invention confer unexpectedly effective protection on vaccinated mice. This suggests the possibility of employing the nucleic acid vaccine of the invention as an early childhood vaccine for
20 humans. In contrast, JE-VAX, the inactivated human vaccine currently used, does not appear to be effective in young animals.

Example 8. Passive protection of neonatal mice correlated with the maternal antibody titer. Female ICR mice at the age of 3 weeks were vaccinated with either one dose or two doses spaced two days apart of pCDJE2-7 plasmid DNA, at 100 μ g/100
25 μ L, or with two doses of JE-VAX that were one-fifth the dose given to humans. The negative control group received two doses of 100 μ g/100 μ L of pCDNA-3/CAT plasmid. Passive protection by maternal antibody was evaluated in pups resulting from matings of experimental females with non-immunized male mice that occurred nine weeks following the first vaccination or 6 weeks following the second vaccination.
30 Pups were challenged between 3-15 days after birth by intraperitoneal administration of

5,000 pfu/100 μ L of mouse-adapted SA14 virus and observed daily for 3 weeks (Table 6). The survival rates correlated with the maternal neutralizing antibody titers. 100% of pups nursed by mothers with a PRNT of 1:80 survived viral infection, whereas none of the pups from the control mother survived (Table 6). Partial protection of 45% and 75% was observed in older pups that were nursed by mothers with a PRNT titer of 1:20 and 1:40, respectively. The survival rates also correlated with the length of time that pups were nursed by the immune mother. As just indicated, 13-15 day old pups had high survival rates. None of the 3-4 day old pups, however, survived virus challenge when the mother had a PRNT titer of 1:20 or 1:40. Thus maternal antibody provides partial to complete protective immunity to the offspring. In addition, JEV antibody was detected by ELISA in the sera of 97% (29/30) of the post-challenge pups.

Mice were inoculated intramuscularly with 1 or 2, 100 μ g doses of plasmid DNA, or subcutaneously with two, 1/5 human doses of JE-VAX vaccine. Sera were collected 9 weeks post-vaccination for PRNT testing prior to mating with non-immune male.

Example 9. Preparation of recombinant plasmids containing the transcriptional unit encoding WNV prM and E antigens. Genomic RNA was extracted from 150 μ L of Vero cell culture medium infected with NY 99-6480 strain, an strain isolated from the outbreak in New York 1999, using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 μ l of nuclease-free water, and used as a template for the amplification of WNV prM and E gene coding sequences. Primer sequences were obtained from the work of Lanciotti et al. (*Science* 286: 2333-2337 (1999)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction sites BsmBI and KasI were engineered at the 5' terminus of the cDNA by using ampimer WN466 (nucleotide sequence, SEQ ID NO:12). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using ampimer cWN2444 (SEQ ID NO:13). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon produced by use of the two amplimers above (SEQ ID NO:12 and SEQ ID NO:13) was digested with KasI and NotI enzymes to generate a 998 bp (nt-1470 to 2468) fragment of DNA was inserted into the KasI and NotI sites of a pCBJESS vector to form an intermediate plasmid, pCBINT. The
5 pCBJESS was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal sequence element (Chang et al., *J. Virol.* 74: 4244-4252 (2000)). The JE signal sequence element comprises the JE signal sequence (SEQ ID NO:14).

The cDNA amplicon was subsequently digested with BsmBI and Kas I enzymes
10 and the remaining 1003 bp fragment (nt-466 to 1470) was inserted in to the KasI site of pCBINT to form pCBWN (nucleic acid sequence, SEQ ID NO:15; amino acid sequence, SEQ ID NO:16). Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Lanciotti
15 et al. (*Science* 286: 2333-2337 (1999)).

Plasmid DNA for use in the in vitro transformation of mammalian cells or mouse immunization experiments was purified by anion exchange chromatography as described in Example 1.

Example 10. Immunochemical characterization and evaluation of WNV prM
20 and E proteins expressed by pCBWN. WNV specific gene products encoded by the pCBWN plasmid were expressed in COS-1 cells. Cells were electroporated and transformed with pCBWN plasmid according to Chang et al. (*J. Virol.* 74: 4244-4252 (2000)). Electroporated cells were seeded onto 75 cm² culture flasks or a 12-well tissue culture dish containing one sterile coverslip/well. All flasks and 12-well plates were
25 kept at 37°C, 5% CO₂ incubator. Forty hours following electroporation, coverslips containing adherent cells were removed from the wells, washed briefly with PBS, fixed with acetone for 2 minutes at room temperature, and allowed to air dry.

Protein expression was detected using indirect immunofluorescence antibody

assay (IFA), as described in Example 2. Flavivirus E-protein specific monoclonal antibody (Mab) 4G2, WNV mouse hyperimmune ascitic fluid (HIAF) and normal mouse serum (NMS) at 1:200 dilution in PBS were used as the primary antibody to detect protein expression (Henchal et al., *Am. J. Trop. Med. Hyg.* 31: 830-836 (1982)).

5 Tissue culture medium was harvested 40 and 80 hours following electroporation. Antigen-capture (Ag-capture) ELISA was used to detect secreted WN virus antigen in the culture medium of transiently transformed COS-1 cells. The Mab 4G2 and horseradish peroxidase-conjugated Mab 6B6C-1 were used to capture the WN virus antigens and detect captured antigen, respectively (Chang et al., *J. Virol.* 74: 4244-4452 (2000); Henchal et al., *Am. J. Trop. Med. Hyg.* 31: 830-836 (1983); Roehrig et al., *Virology* 128: 118-126 (1983)).

WN virus antigen in the medium was concentrated by precipitation with 10 % polyethylene glycol (PEG)-8000. The precipitant was resuspended in TNE buffer (50 mM Tris, 100 mM NaCl, 10 mM EDTA, pH 7.5), clarified by centrifugation, and
15 stored at 4°C. Alternatively, the precipitant was resuspended in a lyophilization buffer (0.1 M TRIZMA and 0.4% bovine serum albumin in borate saline buffer, pH 9.0), lyophilized and stored at 4°C. Lyophilized preparations were used as antigen for the evaluation in MAC- and indirect IgG ELISAs.

WN virus-specific protein was detected by IFA on the transiently transformed
20 COS-1 cells. E, prM and M proteins expressed in these cells were secreted into the culture medium. WN virus antigen concentrated by PEG precipitation was extracted with 7.0 % ethanol to remove residual PEG (Aizawa et al., *Appl. Enviro. Micro.* 39: 54-57 (1980)). Ethanol extracted antigens and gradient-purified WN virions were analyzed on a NuPAGE, 4-12% gradient Bis-Tris Gel in a Excel Plus Electrophoresis Apparatus
25 (Invitrogen Corp., Carlsbad, CA) and followed by electroblotting onto nitrocellulose membranes using a Excel Plus Blot Unit (Invitrogen Corp.). WN virus-specific proteins produced by the transiently transformed COS-1 cells were detected by WN virus specific mouse HIAF or flavivirus E protein reactive Mab 4G2 in a Western blot analysis, using NMS as a negative serum control. The proteins displayed similar

reactivity and identical molecular weights to the corresponding gradient purified virion E, prM and M protein derived from WN virus infected suckling mouse brain (SMB).

In analysis of the NRA as an antigen for diagnostic ELISA, one vial of lyophilized NRA, representing antigen harvested from 40 ml of tissue culture fluid, was reconstituted in 1.0 ml of distilled water and compared with the reconstituted WN virus infected suckling mouse brain (SMB) antigen provided as lyophilized as β -propiolactone-inactivated sucrose-acetone extracts (Clarke et al., *Am. J. Trop. Med. Hyg.* 7: 561-573 (1958)). All recombinant proteins, prM, M and E, had a similar reactivity to that of the gradient-purified virion E, prM and M proteins.

10 Coded human specimens were tested concurrently with antigens in the same test at the developmental stage. The MAC- and IgG ELISA protocols employed were identical to the published methods (Johnson et al., *J. Clin. Microbiol.* 38: 1827-1831 (2000); Martin et al., *J. Clin. Microbiol.* 38: 1823-1826. (2000)). Human serum specimens were obtained from the serum bank in our facility, which consists of
15 specimens sent to the DVBID for WN virus confirmation testing during the 1999 outbreak. In these tests, a screening MAC- and IgG ELISA were performed on a 1:400 specimen dilution. Specimens yielding positive/negative (P/N) OD ratios between 2 and 3 were considered suspect positives. Suspect serum specimens were subject to confirmation as positives by both ELISA end-point titration and plaque-reduction
20 neutralization test (PRNT). All specimens yielding P/N OD ratios greater than 3.0 were considered positives without further confirmatory testing.

An Ag-capture ELISA employing flavivirus-group reactive, anti-E Mab, 4G2 and 6B6C-1, was used to detect NRA secreted into culture fluid of pCBWN transformed COS-1 cells. The antigen could be detected in the medium one day
25 following transformation; and the maximum ELISA titer (1:32-1:64) in the culture fluid without further concentration was observed between day two and day four. NRA was concentrated by PEG precipitation, resuspended in a lyophilization buffer, and lyophilized for preservation. For diagnostic test development, one vial of lyophilized NRA was reconstituted with 1.0 ml distilled water and titrated in the MAC- or indirect

IgG ELISA using WN virus positive and negative reference human sera (Johnson et al., *J. Clin. Microbiol.* 38: 1827-1831 (2000); Martin et al., *J. Clin. Microbiol.* 38: 1823-1826 (2000)). Dilutions 1:320 and 1:160 of the NRA were found to be the optimal concentrations for use in MAC- and IgG ELISA, respectively. These dilutions resulted in a P/N OD₄₅₀ ratio of 4.19 and 4.54, respectively, for MAC- and IgG test. The WN virus SMB antigens produced by NY-6480 and Eg101 strains were used at 1:320 and 1:640 dilution for MAC-ELISA, and 1:120 and 1:320 for IgG ELISA, respectively. The negative control antigens, PEG precipitates of the culture medium of normal COS-1 cells and normal SMB antigen, were used at the same dilutions as for the respective NRA and SMB antigen. Human serum specimens, diluted at 1:400, were tested concurrently in triplicate with virus-specific and negative control antigens. For the positive test result to be valid, the OD₄₅₀ for the test serum reacted with viral antigen (P) had to be at least two-fold greater than the corresponding optical density value of the same serum reacted with negative control antigen (N).

The reactivity of NRA and NY-06480, Eg101 and SLE virus SMBs were compared by the MAC- and IgG ELISAs using 21 coded human serum specimens. Of the 21 specimens, 19 had similar results on all three antigens (8 negatives and 11 suspect positives or positives). Eighteen specimens were also tested separately using SLE SMB antigen. Only three of 13 Eg-101-SMB positive specimens were positive in the SLE MAC-ELISA (Table 1). None of WN antigen negative specimens was positive by SLE MAC-ELISA. This result confirmed a previous observation that anti-WN virus IgM did not cross-react significantly with other flaviviruses (Tardei et al., *J. Clin. Microbiol.* 38: 2232-2239 (1940)) and was specific to diagnose acute WN virus infection regardless of whether NRA or SMB antigen was used in the test. All of the specimens were also tested concurrently by indirect IgG ELISA. Ten of 21 specimens were positive using any of the three antigens.

The two discrepant serum specimens (7 and 9) both from the same patient, collected on day-4 and 44 after onset of disease, respectively, were IgM-negative with NRA and SMB NY antigen and IgM-positive using Eg-101 SMB antigen in the initial test. To investigate these two discordant specimens further, six sequentially collected

specimens from this patient were retested by end-point MAC- and IgG ELISAs. A greater than 32-fold serial increase shown in the MAC-ELISA titer between day-3 and day-15 could be demonstrated with all antigens used. Cerebrospinal fluid collected on day-9 after onset of disease also confirmed that this patient indeed was infected by WN shortly prior to taking the sample. The cerebrospinal fluid had IgM P/N reading of 13.71 and 2.04 against Eg-101- and SLE-SMB antigens, respectively. Day-31 and day-44 specimens were negative (<1:400) by using NY-SMB antigen but positive by using NRA and Eg101-SMB. Compatible IgG titers were observed with all three antigens used in the test.

10 Example 11. Evaluation of the immune response in animals vaccinated with pCBWN. Groups of ten, three-wk-old female ICR mice were used in the study. Mice were injected intramuscularly (i.m.) with a single dose of pCBWN or a green fluorescent protein expressing plasmid (pEGFP) DNA (Clontech, San Francisco, CA.). The pCBWN plasmid DNA was purified from XL-1 blue cells with EndoFree Plasmid
15 Giga Kits (Qiagen) and resuspended in PBS, pH 7.5, at a concentration of 1.0 $\mu\text{g}/\mu\text{l}$. Mice that received 100 μg of pEGFP were used as unvaccinated controls. Mice were injected with the pCBWN plasmid at a dose of 100, 10, 1.0, or 0.1 μg in a volume of 100 μl . Groups that received 10, 1.0, or 0.1 μg of pCBWN were vaccinated by the electrotransfer mediated *in vivo* gene delivery protocol using the EMC-830 square wave
20 electroporator (Genetronics Inc. San Diego, CA.). The electrotransfer protocol was based on the method of Mir et al., (*Proc. Natl. Acad. Sci. USA* 96: 4262-4267.(1999)). Immediately following DNA injection, transcutaneous electric pulses were applied by two stainless steel plate electrodes, placed 4.5-5.5 mm apart, at each side of the leg. Electrical contact with the leg skin was ensured by completely wetting the leg with
25 PBS. Two sets of four pulses of 40 volts/mm of 25 msec duration with a 200 msec interval between pulses were applied. The polarity of the electrode was reversed between the set of pulses to enhance electrotransfer efficiency.

Mice were bled every 3 wks following injection. The WN virus specific antibody response was evaluated by Ag-capture ELISA and plaque reduction
30 neutralization test (PRNT). Individual sera were tested by IgG-ELISA, and pooled sera

from 10 mice of each group were assayed by PRNT. All the mice vaccinated with pCBWN had IgG ELISA titers ranging from 1:640 to 1:1280 three wks after vaccination. The pooled sera collected at three and six wks had a Nt antibody titer of 1:80. None of the serum specimens from pEGFP control mice displayed any ELISA or
5 Nt titer to WN virus.

To determine if the single i.m. vaccination of pCBWN could protect mice from WN virus infection, mice were challenged with NY-6480 virus either by intraperitoneal injection or by exposure to the bite of virus-infected *Culex* mosquitoes. Half of the mouse groups were challenged intraperitoneally (ip) at 6 wks post vaccination with
10 1,000 LD₅₀ (1,025 PFU/100 μ l) of NY99-6480 virus. The remaining mice were each exposed to the bites of three *Culex tritaeniorhynchus* mosquitoes that has been infected with NY99-6480 virus 7 days prior to the challenge experiment. Mosquitoes were allowed to feed on mice until they were fully engorged. Mice were observed twice daily for three wks after challenge.

15 It was evident that the presence of Nt antibodies correlated with protective immunity, since all mice immunized with WN virus DNA remained healthy after virus challenge while all control mice developed symptoms of CNS infection 4-6 days following virus challenge and died on an average of 6.9 and 7.4 days after intraperitoneal or infective mosquito challenge, respectively. In the vaccinated group,
20 the pooled sera collected three wks after virus challenge (9-wk post immunization) had Nt antibody titers of 1:640 or 1:320. Pooled vaccinated mouse sera reacted only with E protein in the Western blot analysis.

Groups of ten mice were immunized with 10.0 to 0.1 μ g of pCBWN per animal
25 by use of electrotransfer. All groups that received pCBWN were completely protected from virus challenge. At 6 wks after immunization all groups of electrotransfer mice had Nt titer less than four-fold different than animals receiving 100 μ g of pCBWN by conventional i.m. injection without electrotransfer. Both these results evidencing effective immunization suggest that the electrotransfer protocol enhances the
30 immunogenicity and protective efficacy of the DNA vaccine of the invention (when

carried out as described in (Mir et al., *Proc. Natl. Acad. Sci. USA.* 96: 4262-4267.(1999))).

Mixed-bred mares and geldings of various ages used in this study were shown to be WN virus and SLE virus antibody-negative by ELISA and PRNT. Four horses
5 were injected i.m. with a single dose (1,000 μ g/1,000 μ l in PBS, pH 7.5) of pCBWN plasmid. Serum specimens were collected every other day for 38 days prior to virus challenge, and the WN virus specific antibody response was evaluated by MAC- or IgG ELISA and PRNT.

Two days prior to virus challenge, 12 horses (4 vaccinated and 8 control) were
10 relocated into a bio-safety level (BSL)-3 containment building at the Colorado State University. The eight unvaccinated control horses were the subset of a study that was designed to investigate WN virus induced pathogenesis in horses and the potential of horses to serve as amplifying hosts. Horses were each challenged by the bite of 14 or
15 *Aedes albopictus* mosquitoes that had been infected by NY99-6425 or BC787 virus 12 days prior to horse challenge. Mosquitoes were allowed to feed on horses for a period of 10 min. Horses were examined for signs of disease twice daily. Body temperature was recorded, and serum specimens collected twice daily from days 0 (day of infection) to 10, then once daily through day 14. Pulse and respiration were recorded daily after challenge. The collected serum samples were tested by plaque titration for
20 detection of viremia, and by MAC- or IgG ELISA and PRNT for antibody response.

No systemic or local reaction was observed in any vaccinated horse. Individual horse sera were tested by PRNT. Vaccinated horses developed Nt antibody greater than or equal to 1:5 between days 14 and 31. End point titers for vaccinated horses, #5, #6, #7, and #8, on day-37 (two days prior to mosquito challenge) were 1:40, 1:5, 1:20, and
25 1:20, respectively. Horses vaccinated with the pCBWN plasmid remained healthy after virus challenge. None of them developed a detectable viremia or fever from days 1 to 14. All unvaccinated control horses became infected with WN virus after exposure to infected mosquito bites. Seven of the eight unvaccinated horses developed viremia that appeared during the first 6 days after virus challenge. Viremic horses developed Nt

antibody between day-7 and day-9 after virus challenge. The only horse from the entire study to display clinical signs of disease was horse #11, which became febrile and showed neurologic signs beginning 8 days after infection. This horse progressed to severe clinical disease within 24 hours and was euthanized on day 9. Four representing
 5 horses, #9, 10, 14 and 15, presenting viremia for 0, 2, 4, or 6 days, were selected and used as examples in this example. Virus titers ranged from $10^{1.0}$ PFU/ml of serum (in horse #10), the lowest level detectable in our assay, to $10^{2.4}$ /ml (in horse #9). Horse #14 did not develop a detectible viremia during the test period. However, this horse was infected by the virus, as evidenced by Nt antibody detected after day 12.

10 Anamnestic Nt antibody response was not observed in vaccinated horses as evidenced by the gradual increase in Nt titer during the experiment. Pre-existing Nt antibody in the vaccinated horse prior to mosquito challenge could suppress initial virus infection and replication. Without virus replication, the challenge virus antigen
 15 anamnestic immune response in the vaccinated horse. All vaccinated horses were euthanized at 14 days after virus challenge. Gross pathological and histopathological lesions indicative of WN viral infection were not observed.

Example 12. Preparation of recombinant plasmids containing coding sequences for yellow fever virus (YFV) or St. Louis encephalitis virus (SLEV) prM and E

20 proteins. A strategy similar to constructing the pCDJE2-7 recombinant plasmid was used to prepare YFV and SLEV recombinant plasmids. Genomic RNA was extracted from 150 μ L of YFV strain TRI-788379 or SLE strain 78V-6507 virus seeds using Q1Aamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA.). The viral RNA was used as a template for amplification of YFV or SLEV prM and E gene coding regions. Primers
 25 YFDV389 (nucleotide sequence, SEQ ID NO:4; amino acid sequence, SEQ ID NO:5), cYFDV2452 (SEQ ID NO:6), SLEDV410 (nucleotide sequence, SEQ ID NO:7; amino acid sequence, SEQ ID NO:8) and cSLEDV2449 (SEQ ID NO:9) were used to generate the corresponding recombinant nucleic acids as described above for the preparation of the JEV and WNV recombinant plasmids. RT-PCR amplified cDNA, digested with
 30 KpnI and NotI enzymes, was inserted into the KpnI-NotI site of a eukaryotic expression

plasmid vector, pCDNA3 (Invitrogen). Both strands of the cDNA were sequenced and verified for identity to sequences from YFV strain TRI-788379 or SLEV strain 78V-6507. Recombinant plasmids pCDYF2 and pCDSLE4-3, which contained the nucleotide sequences of the prM and E coding regions for YFV or SLEV, respectively,
5 were purified using an EndoFree™ Plasmid Maxi Kit (Qiagen), and used for *in vitro* transformation or mouse immunization.

YFV or SLEV specific antigens were expressed in COS-1 cells transformed by pCDYF2 or pCDSLE4-3, respectively. The level of expressed proteins was similar to a YFV- or SLEV-infected COS-1 cell control. As in the JEV model, COS-1 cell lines
10 transformed by vectors bearing genes for the viral antigens were obtained which constitutively express YFV or SLEV antigenic proteins. Epitope mapping by IFA using a panel of YFV or SLEV E-specific Mabs indicated that the authentic E protein was expressed by the pCDYF2- or pCDSLE4-3-transformed COS-1 cells. A preliminary study indicated that 100% of three week-old female, ICR mice seroconverted after
15 intramuscular inoculation with a single dose of 100 µg/100 µL of pCDSLE4-3 plasmid in deionized water.

Example 13. Preparation of recombinant plasmids containing coding sequences for St. Louis encephalitis virus prM and E antigens with JEV signal sequence.

Genomic RNA was extracted from 150 µL of Vero cell culture medium infected with
20 MSI-7 strain of St. Louis encephalitis virus using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 µl of nuclease-free water, and used as a template for the amplification of St. Louis encephalitis virus prM and E gene coding sequences. Primer sequences were obtained from the work of Trent et al. (*Virology* 156: 293-304 (1987)). A cDNA fragment
25 containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer SLE463 (SEQ ID NO:30). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cSLE2447 (SEQ ID NO:31). The RT-PCR
30 product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above (SEQ ID NO:30 and SEQ ID NO:31), was digested with AfeI and NotI enzymes to generate a 2004 fragment of DNA (463 to 2466nt), and inserted into the AfeI and NotI sites of a pCBJESS-M vector to form pCBSLE (nucleotide sequence, SEQ ID NO:21; amino acid sequence, SEQ ID NO:22). The pCBJESS-M was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered, modified JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at -4 (Cys to Gly) and -2 (Gly to Ser) position in the original pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Trent et al. (*Virology* 156: 293-304 (1987)).

Example 14. Preparation of recombinant plasmids containing coding sequences for yellow fever virus (YFV) prM and E proteins with JEV signal sequence. Genomic RNA was extracted from 150 μ L of Vero cell culture medium infected with 17D-213 strain of yellow fever virus using the QIAampTM Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 μ L of nuclease-free water, and used as a template for the amplification of yellow fever virus prM and E gene coding sequences. Primer sequences were obtained from the work of dos Santos et al. (*Virus Research* 35: 35-41 (1995)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer YF482 (SEQ ID NO:28). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cYF2433 (SEQ ID NO:29). The RT-PCR product was purified by a QIAquickTM PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above

(SEQ ID NO:28 and SEQ ID NO:29), was digested with AfeI and NotI enzymes to generate a 1971 fragment of DNA (482 to 2452nt), and inserted into the AfeI and NotI sites of a pCBJESS-M vector to form pCBYF (nucleotide sequence, SEQ ID NO:23; amino acid sequence, SEQ ID NO:24). The pCBJESS-M was derived from the

5 pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at -4 (Cys to Gly) and -2 (Gly to Ser) position of JESS in the pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied

10 Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by dos Santos et al. (*Virus Research* 35: 35-41 (1995)).

Example 15. Preparation of recombinant plasmids containing coding sequences for Powassan virus prM and E antigens with JEV signal sequence. Genomic RNA was

15 extracted from 150 μ L of Vero cell culture medium infected with LB strain of Powassan virus using the QIAamp™ Viral RNA Kit (Qiagen, Santa Clarita, CA). Extracted RNA was eluted and suspended in 80 μ L of nuclease-free water, and used as a template for the amplification of Powassan virus prM and E gene coding sequences. Primer sequences were obtained from the work of Mandl et al. (*Virology* 194: 173-184

20 (1993)). A cDNA fragment containing the genomic nucleotide region was amplified by the reverse transcriptase-polymerase chain reaction (RT-PCR). Restriction site AfeI was engineered at the 5' terminus of the cDNA by using amplimer POW454 (SEQ ID NO:25). An in-frame translation termination codon, followed by a NotI restriction site was introduced at the 3' terminus of the cDNA by using amplimer cPOW2417 (SEQ ID

25 NO:26). The RT-PCR product was purified by a QIAquick™ PCR Purification Kit (Qiagen).

The double-stranded amplicon, produced by use of the two amplimers above (SEQ ID NO:25 and SEQ ID NO:26), was digested with AfeI and NotI enzymes to generate a 1983 bp fragment of DNA (454 to 2436nt), and inserted into the AfeI and

NotI sites of a pCBJESS-M vector to form pCBPOW (nucleotide sequence, SEQ ID NO:19; amino acid sequence, SEQ ID NO:20). The pCBJESS-M was derived from the pCBamp plasmid, that contained the cytomegalovirus early gene promoter and translational control element and an engineered JE signal sequence element (SEQ ID NO:27). The JE signal sequence element comprises the modified JE signal sequence at -4 (Cys to Gly) and -2 (Gly to Ser) position of JESS in the pCBJESS plasmid.

Automated DNA sequencing using an ABI prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) was used to confirm that the recombinant plasmid had a correct prM and E sequence as defined by Mandl et al. (*Virology* 194:173-184, (1993)).

Example 16. Preparation of plasmids containing coding sequences for dengue serotype 2 structural proteins. Procedures such as those carried out for other flaviviruses (see Examples 1, 9 and 12-15) are to be followed to prepare vectors including nucleic acid TU's for dengue serotype 2 antigens. According to the examples, the amplimers used for construction of the vectors may be chosen to engineer the normal dengue virus signal sequence or they may be chosen so as to engineer a signal sequence from another flavivirus, such as a modified Japanese encephalitis virus signal sequence.

A plasmid containing the dengue serotype 2 gene region from prM to E is to be constructed. The dengue serotype 2 prM and E genes (Deubel et al., *Virology* 155:365-377 (1986); Gruenberg et al., *J. Gen. Virol.* 69: 1301-1398 (1988); Hahn et al., *Virology* 162:167-180 (1988)) are to be ligated into a plasmid such as pCDNA3, and then excised and cloned into vectors such as pCBamp, pCEP4, pREP4, or pRc/RSV (supplied by Invitrogen, Carlsbad, CA) to enable expression. If necessary, a dengue serotype 2 virus-specific sequence encoded in a cDNA sequence may be amplified using a procedure such as the polymerase chain reaction (PCR). Alternatively, if the viral RNA is the source of the gene region, a DNA sequence may be amplified by a RT-PCR procedure. A DNA fragment including an initiation codon at the 5' end, and a termination codon at the 3' end is to be cloned into an expression vector at an

appropriate restriction nuclease-specific site, in such a way that the cytomegalovirus (CMV) immediate early (IE) promoter, an initiation codon, and a terminator, are operably linked to the dengue serotype 2 virus sequence.

Example 17. Vaccination of mice using a dengue serotype 2 DNA vaccine. The
5 dengue serotype 2 nucleic TU vaccine encoding the gene region from prM to E
prepared in Example 16 is to be suspended in a suitable pharmaceutical carrier, such as
water for injection or buffered physiological saline, and injected intramuscularly into
groups of weanling mice. Control groups receive a comparable plasmid preparation
lacking the dengue serotype 2 specific genes. The generation of dengue serotype 2-
10 specific antibodies, and/or of dengue serotype 2-specific immune system cytotoxic
cells, is to be assessed at fixed intervals thereafter, for example at weekly intervals. At
about two to four months after administration of the nucleic acid TU vaccine, mice are
to be challenged with dengue serotype 2 virus. Levels of viremia are to be assessed at
appropriate intervals thereafter, such as every second day. Passive protection by
15 maternal antibody is to be assessed as indicated in Example 8.

Example 18. Design and construction of Improved Signal Peptides. Signal peptides can
determine the translocation and orientation of inserted protein, hence, the topology of
prM and E proteins. The most common feature of signal peptides of eukaryotes
consists of an 8 to 12 stretch of hydrophobic amino acids called the h-region (von
20 Heijne, "Signal sequences. The limits of variation" *J. Mol. Biol.* 184: 99-105 (1985)).
The region between the initiator Met and the h-region, which is known as the n-region,
usually has one to five amino acids, and normally carries positively charged amino
acids. Between the h-region and the cleavage site is the c-region, which consists of
three to seven polar but mostly uncharged amino acid residues. During viral
25 polyprotein synthesis, modulation of the signalase cleavage site from a cryptic to
cleavable conformation at the junction of C and prM proteins depends on prior removal
of the C protein by the viral protease complex, NS2B/NS3 (Lobigs, "Flavivirus
premembrane protein cleavage and spike heterodimer secretion require the function of
the viral proteinase NS3," *Proc. Natl. Acad. Sci. U S A.* 90: 6218-6222 (1993)). Thus,

it is critical to consider the effectiveness of the viral signal sequence when prM and E proteins are to be expressed alone by an expression plasmid.

Signal peptide differences in various plasmid constructs can account, at least in part, for the difference in protein translocation, cleavage site presentation and correct topology, thus, prM and E secretion and VLP formation. The modulation or optimization of these attributes can be improved by selection or use of signal sequences with properties that impart the desired characteristics. This can be accomplished by use of machine-learning computer programs using, for example, a hidden Markov model (HMM) trained on eukaryotes (see Henrik Nielsen et al., "Prediction of signal peptides and signal anchors by a hidden Markov model," In Proceedings of the Sixth International Conference on Intelligent Systems for Molecular Biology (ISMB 6), AAAI Press, Menlo Park, California, pp. 122--130 (1998); Nielsen et al., "Machine learning approaches to the prediction of signal peptides and other protein sorting signals," *Protein Engineering* 12: 3-9 (1999); Nielsen et al., "A neural network method for identification of prokaryotic and eukaryotic signal peptides and prediction of their cleavage sites," *Int. J. Neural Sys.* 8: 581-599 (1997); "From sequence to sorting: Prediction of signal peptides," Henrik Nielsen, Ph.D. thesis. Defended at Department of Biochemistry, Stockholm University, Sweden (May 25 1999); each of which is incorporated herein by reference, particularly for the teaching related to the optimization of signal sequences using computer-enabled algorithms).

An example of the type of program used is that found at <http://www.cbs.dtu.dk/services/SignalP-2.0/> as of April 3, 2002. The HMM described in the referred to and incorporated references was applied to calculate the signal peptide probability of the prM signal peptide sequences in different plasmid constructs (Table 7). SignalP-HMM searches correctly predicted the signal peptidase cleavage sites in all constructs. However, considerable differences in cleavage probability (ranging between 0.164 and 1.000) and in signal peptide probability (ranging between 0.165 to 1.00) were observed (Table 7). This is not surprising, as the cleavage site and signal peptide probability are known to also be influenced by the positively charged amino acids in the n-region, the length of the hydrophobic amino acid in the h-region and

amino acid composition in the c-region in the constructs (Chang et al., "Flavivirus DNA vaccines: current status and potential," *Annals of NY Acad. Sci.* 951: 272-285 (2001); Sakaguchi et al., "Functions of Signal and Signal-Anchor Sequences are Determined by the Balance Between the Hydrophobic Segment and the N-Terminal Charge," *Proc. Natl. Acad. Sci. USA* 89: 16-19 (1992)).

Three JE virus plasmid constructs, each derived from different strain of JE virus, showed different vaccine potentials (Lin et al., "DNA immunization with Japanese encephalitis virus nonstructural protein NS1 elicits protective immunity in mice," *J. Virol.* 72: 191-200 (1998); Konishi et al., "Induction of protective immunity against Japanese encephalitis in mice by immunization with a plasmid encoding Japanese encephalitis virus premembrane and envelope genes," *J. Virol.* 72: 4925-4930 (1998); Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)). The signal peptide sequences in these constructs are different in the length of n-region which may or may not contain charged amino acids (Table 7). The n-region containing positively charged amino acids forms a short loop in the cytoplasmic side that causes the h-region (transmembrane helix) to be inserted in a tail orientation, exposing the signalase cleavage site. In our study, secreted VLPs containing prM/M and E proteins could be purified from culture medium of the pCDJE2-7 transformed cell line, JE4B, or pCBJE1-14 transiently transformed COS-1 cells. The gradient-purified VLPs and virions have identical immunological and biochemical properties. Processing efficiency from prM to mature M protein, the hallmark of flavivirus morphogenesis, is also similar between VLPs and virion particles. Thus, prM and E proteins expressed by pCDJE2-7 and pCBJE1-14 can be expressed as type I transmembrane proteins in the orientation similar to that of virion prM and E (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)). In contrast, the prM protein of pcDNA3JEME could be expressed as a type II membrane protein with its transmembrane h-region inserted in a head orientation because of the absence of positively charged amino acids in its n-region (Konishi et al., "Induction of protective immunity against Japanese encephalitis

in mice by immunization with a plasmid encoding Japanese encephalitis virus premembrane and envelope genes," *J. Virol.* 72: 4925-4930 (1998)). Efficient protein synthesis in combination with the expressed protein having the correct topology, particularly of the expressed prM and E, can enhance VLP formation and secretion, and
5 thus promote the immunogenicity of the DNA vaccine (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)).

The use of computer-based computations, as described above, has been applied to optimize the design of expression plasmid. In particular, the predictive power of the
10 SignalP-HMM program was applied to design the WN virus expression plasmid (Table 2) (Davis et al., "West Nile virus recombinant DNA vaccine protects mouse and horse from virus challenge and expresses in vitro a noninfectious recombinant antigen that can be used in enzyme-linked immunosorbent assays," *J. Virol.* 75: 4040-4047 (2001)). The pCBWN plasmid consists of a short version of JE virus signal peptide followed by
15 WN virus prM-E gene sequence. Vaccine potential of this construct was amply demonstrated, as a single i.m. injection of pCBWN DNA not only induced a protective immunity but also prevented WN virus infection in mice and horses.

As discussed earlier, and as demonstrated in Examples 13-15, virus-encoded signal sequence from the same virus as the antigen-coding regions is not necessarily the
20 optimal signal peptide available. Further, non-modified signal sequence is not necessarily optimal. For example, the signal peptide encoded in the pCBJE1-14 plasmid can be improved, as measured by the signal sequence probability by shortening n-region, by altering c-region sequence, or by the combination of both modifications (Figure 6). By way of illustration, a shortened version of JE virus signal peptide has
25 been used for the expression of WN virus prM and E genes as described herein and in papers incorporated herein by reference for the teaching (Davis et al., "West Nile virus recombinant DNA vaccine protects mouse and horse from virus challenge and expresses in vitro a noninfectious recombinant antigen that can be used in enzyme-linked immunosorbent assays," *J. Virol.* 75: 4040-4047 (2001)). Dose titration studies

by single i.m. inoculation indicated that the pCBWN was at least 2-4 fold more immunogenic than pCBE1-14 in mice.

Example 19. Multivalent vaccines. Multivalent and/or combination vaccines designed to immunize against multiple flaviviruses can also be produced. In preparation of a multivalent vaccine, monovalent vaccine components are prepared that include elements related to pathogens of interest, such as YF, different serotypes of DEN, JE, WN, SLE and TBE (RSSE and CEE) viruses or any other combination of flaviviruses. Design and production of DNA constructs as described in the other examples and in the specification are carried out as described. Combinations of appropriate vaccines can be made to provide multivalent or combination vaccines protective against multiple pathogens. Preliminary data from our group has demonstrated that i.m. injection of the combined pCBE1-14 and pCBWN DNA vaccines induced JE virus- and WN virus-specific Nt antibodies in mice (Table 8). Each monovalent component, even if constructed using identical transcriptional and translational regulators, should preferably be tested in an analogous model system to ensure its vaccine potential. A combination vaccine cocktail can then be formulated. These vaccine cocktails can be tailored specifically for particular geographic regions. For example, a vaccine cocktail for tropical and subtropical Asia should include four serotypes of DEN, WN and JE virus vaccines. Similarly useful vaccine cocktails for Africa and Latin America should include four serotypes of DEN, WN and YF virus vaccines and four serotypes of DEN, Rocio and YF virus vaccines, respectively.

Example 20. Preparation and testing of recombinant Dengue virus type 2 vaccines.

a. Summary of example. A series of plasmids that encode the premembrane (prM) and envelope (E) proteins of the dengue virus type 2 (DEN-2) were constructed. These plasmids included an authentic DEN-2 prM-E construct (pCBD2-14-6)(SEQ ID NO:42) encoding the protein described by SEQ ID NO:43, a 90% DEN-2 E-10% Japanese encephalitis (JE) virus E chimeric construct (pCB9D2-1J-4-3)(SEQ ID NO:44) that encodes the protein described by SEQ ID NO:45 and an 80% DEN-2 E-20% JE E chimeric construct (pCB8D2-2J-2-9-1)(SEQ ID NO:46) that encodes the

protein described by SEQ ID NO:47. Monoclonal antibody (MAb) reactivity indicated that all three plasmids expressed E protein epitopes that reacted with a panel of domain 1, 2 and 3 antibodies. However, only the pCB8D2-2J-2-9-1 construct (SEQ ID NO:46) secreted high levels of prM, M (matured prM) and E into the media of the plasmid-transformed COS-1 cells. The major portion of the prM and E protein expressed by COS-1 cells transformed with pCBD2-14-6 plasmid (SEQ ID NO:42) and by COS-1 cells transformed with pCB9D2-4-3 plasmid (SEQ ID NO:44) remained membrane-bound. Replacement of 20% of the sequence encoding the E protein of DEN-2 E with sequence encoding the corresponding JE E protein sequence had no effect on MAb reactivity.

In testing, groups of mice received two intramuscular immunization of selected plasmids at 0 and 3-wks, and the immune response was evaluated by determining specific neutralizing and ELISA antibody. The plasmid expressing secreted prM and E, which can form subviral-particles (SVPs), was superior to other constructs in stimulating an antibody response. Ninety percent neutralization titers ranging from 1:40 to >1:1000 were observed from the 7 of 9 serum specimens of pCB8D2-2J-2-9-1 immunized mice.

b. Importance of DEN-2 virus and vaccines. Dengue (DEN) fever is an acute infection that occurs in subtropical and tropical areas. It is one of the most important flaviviral diseases of humans. As noted earlier, there are four distinct DEN serotypes (DEN-1, DEN-2, DEN-3 and DEN-4) of dengue virus. Infection with any of these is usually either asymptomatic or only causes a self-limited febrile illness known as dengue fever (DF). However, in a small percentage of cases, dengue virus infection results in a much more serious disease, life-threatening dengue hemorrhage fever or dengue shock syndrome (DHF/DSS). Thus, while there are approximately 100 million cases of the relatively mild DF world-wide annually which are of limited concern, there are also an estimated 500,000 hospitalized DHF/DSS cases reported annually. To protect against this disease, a safe and effective DEN vaccine effective against all four serotypes is required for administration to children and non-immune adults in the DEN endemic and epidemic regions.

Safe vaccines must minimize the potential risk of serious infection by virulent viruses. Such virulent viruses can arise by gene reversion or recombination of some types of vaccines derived from attenuated vaccine viruses. Such occurrences did arise in the poliovirus eradication campaign (Guillot et al., "Natural Genetic Exchanges between Vaccine and Wild Poliovirus Strains in Humans," *J. Virol.* 74: 8434-8443 (2000); Liu et al., "Molecular Evolution of a Type 1 Wild-Vaccine Poliovirus Recombinant during Widespread Circulation in China," *J. Virol.* 74: 11153-11161 (2000)). Further, genomic sequencing of an American strain of yellow fever virus, TRINID79A, demonstrates that there is extensive similarity between this strain and the attenuated yellow fever vaccine virus, FNV (Chang et al., "Nucleotide sequence variation of the envelope protein gene identifies two distinct genotypes of yellow fever virus," *J. Virol.* 69: 5773-5780 (1995); Pisano et al., "Complete nucleotide sequence and phylogeny of an American strain of yellow fever virus, TRINID79A," *Arch. Virol.* 144: 1837-1843 (1999)). While not conclusive in and of itself, the similarity strongly suggests that TRINID 79A is derived from the FNV vaccine virus.

The use of DNA-based vaccines is a novel and promising immunization approach for the development of flavivirus vaccines (as described herein, in Chang et al., "Flavivirus DNA vaccines: current status and potential," *Ann. NY Acad. Sci.* 951: 272-285 (2001), and in references cited therein). In this example, a number of DEN-2 vaccines were produced and the immune response in mice after i.m. immunization of DEN-2 constructs were correlated with the efficiency of prM/M and E secretion. One construct that led to the secretion of significant amounts of prM/M and E antigens was shown to be capable of stimulating high titers of neutralizing antibodies in plasmid-vaccinated mice.

c. Materials and methods.

i. Cell culture and virus strains. COS-1 cells (ATCC, Manassas, VA; 1650-CRL) were grown at 37°C with 5 % CO₂ in Dulbecco's modified Eagle minimal essential medium (DMEM, GIBCO, Grand Island, NY) supplemented with 10% heat-inactivated fetal bovine serum (FBS, Hyclone Laboratories, Inc., Logan, UT), 1mM sodium pyruvate, 1mM non-essential amino acids, 30 ml/liter 7.5% NaHCO₃, 100 units/ml of penicillin, and 100 µg/ml of streptomycin. Vero and C6/36 cells were grown under the same conditions used for COS-1 cells. DEN-2 virus, strain-16681,

was used for cDNA cloning , IgG ELISA and the plaque reduction neutralization test (PRNT). Virus was propagated in the C6/36 cell culture. Virus used for immunological or biochemical studies was purified by precipitation with 7% polyethylene glycol (PEG-8000; Fisher Scientific, Fair Lawn, NJ) followed by
 5 ultracentrifugation on 30% glycerol-45% potassium-tartrate gradients (Obijeski et al., "Segmented genome and nucleocapsid of La Crosse virus," *J. Virol.* 20:664-675 (1976)).

ii. Plasmid construction. Genomic RNA was extracted from 150 µl of C6/36 cell culture medium infected with DEN-2 16681 strain using the QIAamp™ Viral RNA
 10 Kit (Qiagen, Santa Clarita, CA). Extracted RNA was resuspended in 80 µl of diethyl pyrocarbonate-treated water (DEPC, Sigma, ST. Louis, MO) and then used as a template in reverse transcriptase-PCR (RT-PCR) amplification of DEN-2 virus prM and E genes. Primer sequences (Table 9) were designed based on published sequences (Gadkari et al., "Critical evaluation of Kyasanur Forest disease virus neutralizing
 15 antibodies found in bats (a preliminary report)," *Indian J. Med. Res.* 64: 64-67 (1976); Kinney et al., "Construction of infectious cDNA clones for dengue 2 virus: strain 16681 and its attenuated vaccine derivative, strain PDK-53," *Virology* 230: 300-308 (1997)). The recognition and cleavage site for restriction enzyme KasI was incorporated at the 5' terminus of the cDNA amplicon. An in-frame termination codon, followed by a
 20 NotI restriction site, was introduced at the 3' terminus of the cDNA amplicon. The DEN-2 virus cDNA amplicon was digested with KasI and NotI enzymes, and was then inserted into the KasI and NotI sites of a pCBJESS vector to form the 100% DEN-2 E plasmid, pCBD2-14-6 (SEQ ID NO:42).

To construct the 90% and 80% DEN-2 E plasmids, the 100% DEN-2 plasmid,
 25 pCBD2-14-6 (SEQ ID NO:42), and the JE plasmid, pCBJE1-14 (SEQ ID NO:17), were used as the PCR templates to amplify DEN-2 and JE DNA sequence, respectively. Sets of primers used in amplification reactions to obtain the DEN-2 and JE gene fragments are listed in Table 9. T7 and SP6 priming sites are found in the pCBamp plasmid, derived from original pCDNA-3 plasmid (Invitrogen, Carlsbad, CA), and can be
 30 utilized as desired or as required. PCR amplified DNA fragments for the 90% DEN-2-

10% JE E protein gene were digested with BstXI restriction endonuclease, ligated using T4 DNA ligase, digested with KsaI and NotI enzyme, and inserted into the KsaI and NotI sites of the pCBJESS vector to obtain the plasmid, pCB9D2-1J-4-3 (SEQ ID NO:44). PCR amplified DNA fragments for the 80% DEN-2-20% JE E gene were
5 digested with BsmBI, ligated with T4 DNA ligase, digested with KsaI and NotI enzyme, and inserted into the KsaI and NotI sites of the pCBJESS vector to obtain the plasmid, pCB8D2-2J-2-9-1 (SEQ ID NO:46). Schematic representations of the three plasmid constructs are shown in Figure 7. The 90% DEN-2-10% JE E and the 80% DEN-2-20% JE E protein junction regions, respectively, are shown in Table 9.

10 Automated DNA sequencing was performed on an ABI Prism 377 Sequencer (Applied Biosystems/Perkin Elmer, Foster City, CA) according to the manufacture's recommended procedures. Recombinant plasmids with the correct prM and E sequences were identified using sequence analysis.

iii. Transient expression of DEN-2 recombinant antigen in COS-1 cells by
15 electroporation. COS-1 cells were electroporated separately with each DEN-2 plasmid or green fluorescent protein expression plasmid control (pEGFP, Clontech, San Francisco, CA) using the protocol described elsewhere in the examples and in Chang et al., ("A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)).
20 Electroporated cells were seeded onto 75 cm² culture flasks, and kept at 37°C and 5% CO₂. Six hours following electroporation the growth media were replaced with a maintenance media containing 2% fetal bovine serum. Tissue culture medium and cells were harvested separately 48 hours following electroporation for antigen characterization.

25 iv. Epitope mapping using DEN-2 E-specific monoclonal antibodies. Forty-eight hours following electroporation, adherent cells were trypsinized, resuspended in PBS containing 5% goat serum, spotted on a 12 well spot-slide and air dried. Cells adhered to the spot-slide were fixed with acetone for 10 minutes at -20°C and allowed to air dry. E-protein specific monoclonal antibodies (MAb) were used to detect protein

expression by the indirect immunofluorescence antibody assay (IFA), as described previously (Table 10; Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)).

- 5 v. Characterization of the recombinant DEN-2 virus antigen. Tissue culture medium was harvested 48 hours following electroporation. Antigen-capture (Ag-capture) ELISA was used to detect secreted DEN-2 virus antigen in the culture medium of transiently transformed COS-1 cells. The MAb 4G2 and horseradish peroxidase-conjugated MAb 6B6C-1 were used to capture the DEN virus antigens and detect
10 captured antigen, respectively (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000); Hunt et al., "A recombinant particulate antigen of Japanese encephalitis virus produced in stably-transformed cells is an effective noninfectious antigen and subunit immunogen," *J. Virol. Methods.* 97:
15 133-149 (2001)).

- Forty-eight hours following electroporation, transformed cells for each plasmid were trypsinized and resuspended in PBS as aliquots containing 5×10^6 cells. These cell samples were processed for membrane protein extraction using the Mem-PER mammalian membrane protein extraction reagent kit (Pierce, Rockford, IL) following
20 the manufacturer's suggested protocol. Both hydrophobic and hydrophilic proteins are isolated. This procedure was developed for enrichment of integral membrane protein found in the hydrophobic phase. Both hydrophobic and hydrophilic fractions were analyzed by Ag-capture ELISA for DEN-2 recombinant antigen.

- Recombinant antigen in the medium was concentrated by precipitation with
25 10% polyethylene glycol (PEG)-8000. The precipitant was resuspended in TNE buffer (50 mM Tris, 100 mM NaCl, 10 mM EDTA, pH 7.5) to 1/100th of the original volume, clarified by centrifugation, and stored at 4° C. Recombinant antigen concentrated by PEG precipitation and resuspended in TNE buffer was extracted with 4.0 % ethanol to remove residual PEG (Hunt et al., "A recombinant particulate antigen of Japanese

encephalitis virus produced in stably-transformed cells is an effective noninfectious antigen and subunit immunogen," *J. Virol. Methods*. 97: 133-149 (2001)). Ethanol-extracted antigens, hydrophobic membrane proteins from the transformed cells and gradient-purified DEN-2 virions were analyzed on a NuPAGE, 4-12% Bis-Tris gradient gel in an Excel Plus Electrophoresis Apparatus™ (Invitrogen Corp., Carlsbad, CA), followed by electroblotting onto nitrocellulose membranes using a Excel Plus Blot Unit (Invitrogen Corp.). DEN-2 virus-specific protein was detected by western blot using DEN-2 virus specific MAbs 1A6A-8 (E specific) and 1A2A-1 (capsid specific), as well as rabbit antiserum specific for DEN-2 prM and mouse serum specific for a peptide composed of amino acid 1-34 of the DEN-2 M protein, and normal mouse ascitic fluid was used as negative control (Murray et al., "Processing of the dengue virus type 2 proteins prM and C-prM," *J. Gen. Virol.* 74 (Pt 2): 175-182 (1993); Roehrig et al., "Monoclonal antibody mapping of the envelope glycoprotein of the dengue 2 virus, Jamaica," *Virology* 246: 317-328 (1998)).

15 vi. Mouse vaccination. Groups of ten, three-wk-old female ICR out-bred mice were used in the study. Mice were i.m. injected with pCBD2-14-6, pCB9D2-1J-4-3, pCB8D2-2J-2-9-1 or pEGFP on week-0 and week-3 at a dose of 100 µg in a volume of 100 µl per mouse. The plasmid DNA was purified from XL-1 blue cells with EndoFree Plasmid Giga Kits™ (Qiagen) and resuspended in PBS, pH 7.5, at a concentration of 20 1.0 µg/µl. Mice that received 100 µg of pEGFP were used as plasmid-vaccinated controls. Mice were bled every 3 wks following injection, and the DEN-2 virus specific antibody response was evaluated by use of indirect ELISA and PRNT.

vii. Serological tests. Pre- and post-vaccination serum specimens were tested for antibody binding ability to purified DEN-2 virion by ELISA, for neutralizing (Nt) antibody by PRNT, and for antibodies that recognize purified DEN-2 virus proteins by Western blotting. PRNT was performed with Vero cells, as previously described (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)), using DEN-2 (strain-16681) and JE (strain-Nakayama) virus. Endpoints

were determined at a 90% plaque-reduction level (Hunt et al., "A recombinant particulate antigen of Japanese encephalitis virus produced in stably-transformed cells is an effective noninfectious antigen and subunit immunogen," *J. Virol. Methods*. 97: 133-149 (2001)).

5 d. Results.

i. Transient expression of DEN-2 virus recombinant antigen. Expression of the prM and E genes of DEN-2 virus or a chimeric E gene from a combination of DEN-2 and JE virus sequences (80% DEN-20% JE or 90% DEN-10% JE) was accomplished by separate transformations of each of the three recombinant DEN-2 DNA plasmids
10 into COS-1 cells. The basic plasmid design was based on results from previous studies with JE virus and WN virus recombinant plasmids in which plasmid-transformed cells expressed, and secreted authentic viral proteins into the cell culture fluid (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000);
15 Davis et al., "West Nile virus recombinant DNA vaccine protects mouse and horse from virus challenge and expresses in vitro a noninfectious recombinant antigen that can be used in enzyme-linked immunosorbent assays," *J. Virol.* 75: 4040-4047 (2001)). Transient expression of DEN-2 recombinant proteins was initially assessed by Ag-capture ELISA of cell culture supernatants and by IFA of acetone-fixed, transformed
20 COS-1 cells (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000)). The point of optimum antigen expression was determined to be 48 hours following electroporation.

ii. Epitope mapping of the E protein expressed by transiently-transformed
25 COS-1 cells. The DEN-2 protein expressed by each of the recombinant plasmids was evaluated by IFA using a panel of murine MAbs with known reactivity to DEN-2 virus (Table 10; Henchal et al., "Epitopic analysis of antigenic determinants on the surface of dengue-2 virions using monoclonal antibodies," *Am. J. Trop. Med. Hyg.* 34: 162-169 (1985); Roehrig et al., "Monoclonal antibody mapping of the envelope glycoprotein of
30 the dengue 2 virus, Jamaica," *Virology* 246: 317-328 (1998)). The MAb panel included

antibodies reactive with each of the three antigenic domains of the E protein of flaviviruses as well as prM and C proteins. (Mandl et al., "Antigenic structure of the flavivirus envelope protein E at the molecular level, using tick-borne encephalitis virus as a model," *J. Virol.* 63: 564-571 (1989); Rey et al., "The envelope glycoprotein from tick-borne encephalitis virus at 2 Å resolution," *Nature* 375: 291-298 (1995)). The MAb specific for flavivirus antigenic Domains 2 and 3 showed nearly identical qualitative reactivity with DEN-2 virus and each of the three plasmid-expressed proteins. One of the Domain 1-specific MAbs, 1B4C-2, also showed a similar reactivity pattern with all expressed proteins. However, two of the Domain 1-specific MAbs, 2B3A-1 and 9A4D-1, were much less reactive with E protein expressed by plasmids pCBD2-14-6 and pCB9D2-1J-4-3 as shown by endpoint titration (values in parentheses, Table 10). Comparison of the endpoint titers revealed the apparent poor expression of epitopes C3 and C4 in constructs containing 100% DEN-2 E and 90% DEN-2 E-10% JE E. MAb 2H2, specific for prM, had the same reactivity with antigen expressed by all three plasmids. Anti-C MAb 1A2A-1 reacted well with DEN-2 virus and had low level, non-specific reactivity with the plasmid-expressed viral proteins, which included prM and E, but not C.

iii. Comparison of secreted protein and membrane-bound protein produced by each of the three DEN-2 recombinant plasmids. Similar amounts of cell culture fluid were harvested from COS-1 cells 48 hours post-transformation for each recombinant DEN-2 plasmid. Secreted recombinant antigen found in the culture fluid was concentrated 100-fold by PEG precipitation, followed by ethanol extraction to remove PEG which interfered with subsequent analysis by polyacrylamide gel electrophoresis. The relative amount of secreted antigen expressed by each plasmid was determined by Ag-capture ELISA analysis of both PEG-precipitated and ethanol-extracted cell culture fluid preparations (Table 11). Secreted antigen was detected only from cells transfected with pCB8D2-2J-2-9-1 (SEQ ID NO:34), which contained 80% DEN-2 E and 20% JE E genes. The recombinant plasmids containing either 100% DEN-2 E or 90% DEN-2 E-10% JE E genes produced no ELISA-detectable antigen in the culture fluid, despite efforts to concentrate expressed protein.

Western blot analysis was also used to evaluate the production of secreted antigen by each of the DEN-2 recombinant plasmids. For comparison purposes, equivalent volumes of PEG-precipitated, ethanol-extracted cell culture supernatant were run on NuPAGE gradient gels, electroblotted to nitrocellulose, and analyzed using

5 MAbs or polyclonal antisera capable of reacting with all DEN-2 structural proteins (Figure 8A). Western blot analysis showed greater sensitivity in detecting recombinant antigen than Ag-capture ELISA since DEN-2-specific proteins were detected in the culture fluid from two of the plasmids, pCB8D2-2J-2-9-1 and pCB9D2-1J-4-3 (SEQ ID NOS:46 and 44, respectively). Plasmid pCB8D2-2J-2-9-1 (SEQ ID NO:46) expressed

10 the greatest amount of secreted antigen which was shown to be composed of E, prM, and M proteins. Relatively less secreted antigen was produced by pCB9D2-1J-4-3 (SEQ ID NO:44) and barely detectable levels were found for the pCBD2-14-6 (SEQ ID NO:42) preparation, which appeared to contain relatively less expressed E protein, especially if the non-specific reactivity of the E-specific MAb, 1A6A-8, on the control

15 pEGFP was taken into consideration (Figure 8A, lanes a, b for 14-6 and GFP).

Since E, prM, and M are membrane-associated proteins throughout their intracellular synthesis, any assessment of the expression of these proteins by the three recombinant DEN-2 plasmids should include an evaluation of cell membrane preparations from plasmid-transformed cells. The Mem-PER Mammalian Membrane

20 Protein Extraction Reagent kit (Pierce) was used to isolate the integral membrane proteins from equivalent numbers of cells transformed by each of the recombinant plasmids. Hydrophobic proteins were separated from hydrophilic proteins by phase partitioning. Preliminary analysis by Ag-capture ELISA indicated that the hydrophilic protein fraction was non-reactive; however, the hydrophobic protein fractions from

25 COS-1 cells transformed with each of the recombinant DEN-2 plasmids had similar titers in ELISA tests (Table 11). These results indicated that recombinant antigen encoded by all three plasmids was expressed following transformation, but that the recombinant antigens expressed were not all secreted at the same level.

Confirmation of the Ag-capture results for the hydrophobic protein fractions

30 was accomplished by western blot (Figure 8B). Equivalent volumes of hydrophobic

protein fractions from each of the plasmid-transformed cells were diluted according to the manufacturer's recommendations for SDS-polyacrylamide gel electrophoresis in order to reduce band and lane distortion. Immunoblotting with E-, prM-, C-, and M-specific MAbs or polyclonal antisera demonstrated that all three recombinant DEN-2
5 plasmids induced the production of similar amounts of recombinant antigen composed of E and prM. No M protein was detected, either because it was not processed from prM or because the levels were too low to be detected. Despite efforts to reduce band distortion, high levels of detergent in the hydrophobic protein samples apparently caused E and prM to run in a slightly aberrant manner (slower migration) compared to
10 samples without such high concentrations of detergents (compare E and prM migration in Figure 8A and 8B).

iv. Comparison of the immune response in mice vaccinated with three different DEN-2 recombinant DNA plasmids. Three-week old ICR mice were immunized by i.m. injection with 100 µg of pCB8D2-2J-2-9-1 (SEQ ID NO:46), pCB9D2-1J-4-3
15 (SEQ ID NO:44), pCBD2-14-6 (SEQ ID NO:42), or pEGFP on weeks 0 and 3. Mice were bled 3, 6, and 9 weeks after the primary immunization. Individual and pooled sera were tested by indirect ELISA, using screening dilutions of 1:100 and 1:400 at 3 and 6 weeks post-vaccination and endpoint titrations at 9 weeks post-vaccination. Nine-week sera were also tested by PRNT with both DEN-2 and JE viruses. The
20 ELISA results showed that after one immunization (3-week sera), all mice given pCB8D2-2J-2-9-1 had seroconverted, whereas only 50% of pCB9D2-1J-4-3 and 20% of pCBD2-14-6-vaccinated mice reacted with DEN-2 virus (Table 12). By 9 weeks post-vaccination, all mice vaccinated with either pCB8D2-2J-2-9-1 or pCBD2-1J-4-3 demonstrated anti-DEN-2 ELISA reactivity; however, the geometric mean titers
25 differed significantly (titers of 1:20,000 versus 1:708, respectively). Only 40% of pCBD2-14-6-immunized mice had anti-DEN-2 ELISA titers greater than 1:100. A western blot of pooled 9-week sera from pCB8D2-2J-2-9-1-immunized mice on purified DEN-2 virus showed that the immunodominant response was to the E glycoprotein. Slight reactivity reactivity to prM and M were also detected.
30 More significantly, in terms of evaluating the vaccine potential of the three DEN-2 plasmids, the induction of virus-neutralizing antibody in 7 of 9 mice immunized

with pCB8D2-2J-2-9-1 (SEQ ID NO:46) was observed based on a 90% plaque-reduction endpoint (Table 10). However, if a 50% neutralization endpoint is used, then all 9 of 9 sera have PRNT titers of $\geq 1:40$. Ninety percent neutralization titers ranged from 1:40 to $>1:1000$ for the 7 sera with neutralizing activity. None of the mice immunized with pCB9D2-1J-4-3 produced neutralizing antibody, and only 1 of 10 sera from pCBD2-14-6-vaccinated mice neutralized virus, but at a titer of only 1:8.

Since two of the recombinant plasmids, namely pCB9D2-1J-4-3 (SEQ ID NO:44) and pCB8D2-2J-2-9-1 (SEQ ID NO:46) contained JE virus E-gene sequences, all sera were also evaluated for the presence of JE virus neutralizing activity. However, no such activity was detected at the 90% neutralization endpoint for mice in any of the immunization groups. Not surprisingly, mice immunized with the control plasmid pEGFP exhibited no reactivity to either DEN-2 or JE viruses.

e. Discussion. The same steps used earlier for the JE and WN vaccines was initially used to construct a recombinant DEN-2 plasmid, pCBD2-14-6 (SEQ ID NO:42), consisting of the authentic DEN-2 prM and E gene region. Antigenic mapping of DEN-2 proteins expressed by COS-1 cells transformed by this plasmid, using a panel of MAb by IFA, indicated the prM and E protein had a compatible fluorescent intensity and a similar MAb reactivity as virus infected cells (Table 10). However, these COS-1 cells transformed by the plasmid encoding an authentic DEN-2 prM and E region failed to secrete detectable DEN-2 antigen into the culture fluid (as measured by antigen-capture ELISA). In addition, vaccination using the plasmid encoding an authentic DEN-2 prM and E region failed to stimulate anti-DEN-2 virus neutralizing antibody in i.m. immunized-mice (Table 13). Interestingly, transformation of cells by pCBD2-14-6 resulted in a punctuated-globular fluorescent staining that suggested that the C-terminus of the E protein of DEN-2 may contribute to that protein's membrane retention signal. This IFA staining pattern was not observed in either the JE or WN construct-transformed cells (Chang et al., "A single intramuscular injection of recombinant plasmid DNA induces protective immunity and prevents Japanese encephalitis in mice," *J. Virol.* 74: 4244-4252 (2000); Davis et al., "West Nile virus recombinant DNA vaccine protects mouse and horse from virus challenge and

expresses in vitro a noninfectious recombinant antigen that can be used in enzyme-linked immunosorbent assays," *J. Virol.* 75: 4040-4047 (2001)). Therefore, in light of the observation made in accordance with the teachings of the present application, two additional plasmids, pCB9D2-1J-4-3 (SEQ ID NO:44) and pCB8D2-2J-2-9-1 (SEQ ID
5 NO:46), in which appropriate manipulation of the DNA sequence was made for 10 % or 20 % of C-terminal E of DEN-2 to be replaced with the corresponding region of JE virus E protein, respectively. Relative effectiveness of the different constructs in stimulating detectable anti-DEN-2 ELISA antibody in vaccinated mice is shown in Table 13.

10 These results are consistent with the model that interactions between prM and E can influence the processes of particle assembly and secretion. Support for this model can be found in a study of tick-borne encephalitis virus that putatively suggests that interactions between prM and the ectodomain of E are involved in prM-mediated intracellular transport of prM-E, thus, secretion of virus-like particle (Allison et al.,
15 "Mapping of functional elements in the stem-anchor region of tick-borne encephalitis virus envelope protein E," *J. Virol.* 73: 5605-5612 (1999)).

In the present example, replacing a C-terminal portion of DEN-2 E protein with JE E protein, corresponding to TBE H1^{pred} to TM2, resulted in DEN-2 prM protein and chimeric E protein secretion. However, in contrast, replacement of TM1 and TM2, in
20 TBE, caused only a minor improvement in antigen secretion. The major portion of the prM and E protein expressed by either COS-1 transformed pCBD2-14-6 and pCB9D2-4-3 plasmid remained membrane-bound (Table 13). These results indicated that an unidentified membrane retention sequence is located in the C-terminal stem region of DEN-2 E protein. Substitution of this C-terminal stem region with sequence from the
25 JE virus removes or renders ineffective this retention sequence.

It has been asserted by others that the prM protein is essential for maintaining proper conformation and secretion of E protein during prM-E maturation (Aberle et al., "A DNA immunization model study with constructs expressing the tick-borne encephalitis virus envelope protein E in different physical forms," *J. Immunol.* 163:

6756-6761 (1999), Allison et al., "Synthesis and secretion of recombinant tick-borne encephalitis virus protein E in soluble and particulate form," *J. Virol.* 69:5816-5820 (1995)). Further, it has also been demonstrated that the ectodomain of the E protein interacts with prM. This interaction has been estimated to involve amino acid sequence
5 within amino acid residues 200-327 of E in the Murray Valley encephalitis virus (Guirakhoo et al., "The Murray Valley encephalitis virus prM protein confers acid resistance to virus particles and alters the expression of epitopes within the R2 domain of E glycoprotein," *Virology* 191: 921-931 (1992)).

Proper prM and E interactions and retained integrity of the E protein's structure
10 are likely maintained in the protein expressed by all three DEN-2 constructs, at least insofar as they are required for immunoreactivity. Furthermore, the replacement of the C-terminus 20 % E in the pCB8D2-2J-2-9-1 resulted in a protein that maintained 395 amino acids of authentic DEN-2 E. Any such modification is expected to have minimal influence on E and prM-E interactions and their influence on the antigenic nature of the
15 chimeric E protein. As replacing the C-terminus region of DEN-2 E with JE stem-anchor sequence had no effect on MAbs reactivity (Table 10), retention of the DEN-2 sequence so replaced can only be optional in attaining a DEN-2 specific immunological response.

Previously, it has been shown that a plasmid construct encoding a secreted
20 subviral particle of tick-borne encephalitis virus prM and E protein was superior, in terms of the extent and functionality of the antibody response and in terms of response to virus challenge, to other constructs that encoded C-terminally truncated soluble E-dimer that was secreted, full-length E that was not secreted, or a truncated E that was not efficiently secreted (Aberle et al., "A DNA immunization model study with
25 constructs expressing the tick-borne encephalitis virus envelope protein E in different physical forms," *J. Immunol.* 163: 6756-6761 (1999)). However, here we have demonstrated that the vaccine potency of DEN-2 DNA is correlated with the secretion of prM/M and E (Table 13). The morphology and physical character of secreted prM and E were not demonstrated in this study. However, the prM and E, secreted by the
30 pCB8D2-2J-2-9-1 construct, does likely form a viral-like particle. It is believed that

the presentation of multiple protective antigens on the particle surface improves vaccine potency of this construct.

Previous attempts for the development of DEN-2 virus DNA vaccine has resulted in varying degrees of success (Kochel et al., "Inoculation of plasmids expressing the dengue-2 envelope gene elicit neutralizing antibodies in mice," *Vaccine* 15: 547-552 (1997); Konishi et al., "A DNA vaccine expressing dengue type 2 virus premembrane and envelope genes induces neutralizing antibody and memory B cells in mice," *Vaccine* 18: 1133-1139 (2000)). To improve the level of effectiveness, differing strategies have been adopted. For example, coimmunization of the immuno-
stimulatory CpG motif containing pUC19 plasmid, plasmid expressing murine GM-CSF in the vaccine regimen, or replacing C-terminus 43 amino acid of E with lysosome-associated membrane retention sequence improved the antibody response to the DEN-2 vaccine has been used (Porter et al., "Protective efficacy of a dengue 2 DNA vaccine in mice and the effect of CpG immuno-stimulatory motifs on antibody responses," *Arch. Virol.* 143: 997-1003 (1998); Raviprakash et al., "Synergistic Neutralizing Antibody Response to a Dengue Virus Type 2 DNA Vaccine by Incorporation of Lysosome-Associated Membrane Protein Sequences and Use of Plasmid Expressing GM-CSF," *Virology* 290: 74-82 (2001)). Unmethylated CpG motifs directly activates macrophages, natural killer cells and lymphocytes to secrete cytokines and chemokines, and supports the development of immune responses mediated by Th1 cytokines (Manders et al., "Immunology of DNA vaccines: CpG motifs and antigen presentation," *Inflamm. Res.* 49: 199-205 (2000)). However, inclusion of the CpG might bias the host's cytokine profile and thereby contribute to the both the development of Th-1-mediated organ-specific autoimmune disorders and interfere with immune homeostasis (Smith et al., "The regulation of DNA vaccines," *Curr. Opin. Biotech.* 12: 299-303 (2001)). There is also evidence in mice that excess levels of cytokine, although increasing the response of certain T-helper cells, can decrease or shutdown the response of other actors in the immune response, leading to generalized immunosuppression or chronic inflammation (Robertson et al., "Assuring the quality, safety, and efficacy of DNA vaccines," *Mol. Biotechnol.* 17: 143-149 (2001)). Correspondingly, the safety and efficacy of flavivirus DNA immunization

could be benefited by manipulation of an expression plasmid to enhance transcription and translation and targeting prM and E proteins for secretion that promote correct polyprotein processing and assembly (Chang et al., "Flavivirus DNA vaccines: current status and potential," *Ann. NY Acad. Sci.* 951: 272-285 (2001)). Future improvements

5 could be focused on enhancing DNA uptake by antigen presenting cells or by muscle cells (Rodriguez et al., "Enhancing DNA immunization," *Virology* 268: 233-238 (2000)).

Table 1. Transient expression of JE prM and E proteins by various recombinant plasmids in two transferred cell lines.

	Vector				Recombinant Plasmid	IFA intensity/percentage of antigen-positive cells*	
	Promotor	Intron	Poly (A)	ORI		COS-1	COS-7
pCDNA3	CMV	No	BGH	SV40	pCDJE2-7	3+/40	3+/35
pCBamp	CMV	No	BGH	No	pCBJE1-14	3+/45	nd
pC1Bamp	CMV	Yes	BGH	No	pC1BJES14	3+/39	nd
pCEP4	CMV	No	SV40	OriP	pCEJE	2+/4	2+/3
pREP4	RSV	No	SV40	OriP	pREJE	1+/3	1+/2
pRe/RSV	RSV	No	BGH	SV40	pRCJE	1+/3	1+/3
pCDNA3	CMV	No	BGH	SV40	pCDNA3/CAT	-	-

*Various cell lines were transformed with pCDNA3/CAT (negative control), pCDJE2-7, pCBJE1-14, pC1BJES14, pCEJEm pREJE, or pRCJE. Cells were trypsinized 48 hours later and tested by an indirect immunofluorescent antibody assay (IFA) with JE virus-specific HIAF. Data are presented as the intensity (scale of 1+ to 4+) and the percentage of IFA positive cells. The pCDNA3/CAT transformed cells were used as the negative control.

Table 2. Characterization of proteins expressed by a pCDJE2-7 stably transformed clone (JE-4B) of COS-1 cells with JE virus-reactive antibodies.

Mab or antiserum	Biological Activity of Mab		Immunofluorescent intensity of cells	
	Specificity	Biological Function	JEV infected	4B
Mab:				
MC3	JEV Specific		2+	2+
2F2	JEV Specific	HI, N	4+	4+
112	JEV Specific		4+	4+
503	JEV Specific	N	4+	3+
109	Subgroup	HI	2+	1+
N.04	Subgroup	HI, N	3+	4+
201	Subgroup		1+	1+
203	Subgroup		4+	3+
204	Subgroup		2+	2+
301	Subgroup	HI	2+	2+
504	Flavivirus		4+	4+
6B6C-1	Flavivirus		2+	2+
3B4C-4	VEE		-	-
H1AF:				
Anti-JEV			4+	3+
Anti-WEE			-	-
PBS			-	-

Table 3. Persistence of the immune response in mice immunized with pCDJE2-7 or JE-VEX vaccine.

	ELISA Titer (\log_{10})						PRNT _{90%} Titer		
	3 wks	6 wks	9 wks	23 wks	40 wks	60 wks*	3 wks	6 wks	9 wks
1x pCDJE2-7	2.6-3.2	3.8-5.0	3.8-4.4	>3.2	>3.2	2.4, 2.4, 3.8, 4.4	<20	20	40-160
2x pCDJE2-7	2.6-3.8	4.4	3.8-4.4	>3.2	>3.2	2.6, 3.8, 3.8	<20	20-40	40-160
2x JE-VAX	2.6-3.8	4.4-5.0	3.8-5.6	>3.2	>3.2	<2, <2, 4.4	<20	20-40	20-160
2x pCDNA3/CAT	<2	<2	<2	ND	ND	<2	<20	<20	<20

Mice were inoculated with 1 or 2, 100 μ g/dose plasmid DNA, or 1/5 human dose of JE-VAX vaccine. Sera were collected for testing prior to the second immunization.

* Individual serum titers.

Table 4. The age-dependent percent seropositive rate in mice following vaccination with various JEV vaccines.

	3-day old		3-week old	
	3 weeks PV	7 weeks PV	3 weeks PV	7 weeks PV
JE-VAX	0	0	100	100
pCDNA3/CAT	0	0	0	0
pCDJE2-7	40	60	90	90
pC1BJES14	10	60	80	100
pCBE1-14	80	100	100	100

Table 5. Protection from JEV challenge in 8 week old mice following vaccination at 3 days old with various JEV vaccines.

Vaccine	Pre-challenge JEV seroconversion	Days post-challenge survival rate (%)				
		6	7	8	9	21
JE-VAX	0	100	100	60	40	40
pCDNA3/CAT	0	100	80	30	30	30
pCDJE2-7	60	100	100	100	100	100
pC1BJES14	60	100	100	100	100	100
pCBE1-14	100	100	100	100	100	100

Table 6. Evaluation of the ability of maternal antibody from JEV-nucleic acid-vaccinated female mice to protect their pups from fatal JEV encephalitis.

Vaccinated mother		JEV challenged pups		
Vaccine	PRNT _{90%}	Challenge age (days)	No. survival ¹	ELISA ²
1 x pCDJE2-7	40	4	0/11	
2 x pCDJE2-7	80	4	12/12	12/12
2 x JE-VAX	20	3	0/16	
2 x pCDNA-3/CAT	<10	5	0/14	
1 x pCDJE2-7	20	15	5/11	5/5
2 x pCDJE2-7	40	14	8/12	7/8
2 x JE-VAX	80	13	5/5	5/5
2 x pCDNA-3/CAT	<10	14	0/14	

Mice were inoculated intramuscularly with 1 or 2, 100 µg dose of plasmid DNA, or subcutaneously with two, 1/5 human dose of JE-VAX vaccine. Sera were collected 9 weeks post-vaccination for PRNT testing prior to mating with non-immune male.

¹: No Survivors/total for each litter.

²: Number of JEV ELISA-antibody-positive animals (titer ≥ 1:400)/No. of survivors; sera were collected for testing 12 weeks after challenge.

Table 7. Characteristic of the signal peptides and their vaccine of potentials among flavivirus DNA vaccine constructs

Plasmid	Signal peptide sequence preceding prM protein	Signal peptide probability ^a			Immunization protocol/protection
		SP	AP	Csite	
pSLE1	? <u>LD</u> TINRRPS <u>KKRGGT</u> RSLLGLAALIGLASS/LQLLSTYQG (SEQ ID NO: 32)	0.702	0.292	0.352	im x 2/Partial
pJME	MWLASLAVVIACAGA/M <u>KLSNFQ</u> GK (SEQ ID NO: 33)	0.998	0.000	0.778	im x 2/Partial
pCJEME	MNEGSIMWLASLAVVIACAGA/M <u>KLSNFQ</u> GK (SEQ ID NO: 34)	0.985	0.012	0.785	im x 2/100%
pCBJE1-14	MGRKQNKRGGEGSIMWLASLAVVIACAGA/M <u>KLSNFQ</u> GK (SEQ ID NO: 35)	0.791	0.199	0.623	im x 1/100%
pcDNA3prM-E	MS <u>KKRGGSET</u> SVLMVIFMLIGFAAA/L <u>KLSNFQ</u> GK (SEQ ID NO: 36)	0.721	0.277	0.622	im x 4 / Partial
pCBWN	MG <u>KRS</u> AGSIMWLASLAVVIACAGA/VTL <u>SNFQ</u> GK (SEQ ID NO: 37)	0.976	0.024	0.526	gg x 2-4/100%
p1012D2ME	MNVLRG <u>FRKE</u> IGRMILNLR <u>RRRT</u> AGMIIMLIPVMA/FHLT <u>TRNGE</u> (SEQ ID NO: 38)	0.165	0.778	0.164	im x 1/100%
SV-PE	MVGLO <u>KRGKRR</u> SATDWM <u>SWLLVITLLG</u> MTLA/ATV <u>RKER</u> GD (SEQ ID NO: 39)	0.943	0.056	0.899	id x 2/None
pWRG7077-RSSE	MGWLLVVVLLGVTLA/ATV <u>RKER</u> GD (SEQ ID NO: 40)	1.000	0.000	0.912	im or gg x 2/100%
pWRG7077-CEE	MSWLLVITLLG <u>MTIA</u> /ATV <u>RKER</u> GD (SEQ ID NO: 41)	0.999	0.000	0.821	gg x 2/100%

^a The SignalP HMM program was applied to calculate signal peptide (SP), anchor peptide (AP) and signalase cleavage site (C site) probabilities.

Single amino acid codes were used, and charged amino acids were highlighted by underline-bold letters. The signalase cleavage site separating SP and prM is indicated by "r". DNA vaccines were inoculated by intramuscular (im), intradermal (id), or gene gun (gg) method.

Table 8. Neutralizing antibody (Nt) responses in mice immunized with different doses of the combined WN and JE virus DNA vaccines.

Dose per plasmid (μg)	pCBWN + pCBJE1-14				pCB Control	
	100 +100	40 + 40	20 +20	10 +10	100	100
Percentage of mice with Nt :						
WN virus/JE virus:	100 / 100	100 / 70	70 / 0	60 / 0	0 / 0	
Range of PRNT ₉₀ titer:						
WN virus:	1:320 - 1:80	1:80 - 1:20	1:80 - <1:10	1:20 - <1:10	<1:10	
JE virus:	1:40 - 1:10	1:10 - <1:10	<1:10	<1:10	<1:10	

Groups of ten, three-week-old female ICR out-bred mice were i.m. injected with a single dose of Combined plasmid DNAs as indicated. The serum specimens collected 12-week after immunization were assayed by the plaque-reduction neutralization test (PRNT). The end-point titers against JE and WN virus were calculated based on the 90% percent plaque reduction using JE virus (strain SA-14) and West Nile virus (strain NY-6480), respectively.

Table 9. Oligonucleotides used to construct DEN-2 virus prM-E expression plasmids, and the junction region of chimeric DEN-2 and JE E indicated.

100% DEN-2 prM-E:

D2KasI-438^a 5' TGTGCAGGCGCCTTCCATTTAACCACACGTAACG (SEQ ID NO:48)

CD2NotI-2402 5' TCGAGCGGCCGCTCAACTAATTAGGCTGCACCATGACTC (SEQ ID NO:49)

90% DEN-2 E & 10% JE E:

T7 5' CTTATCGAAATTAATACGACTCACTATAGG (SEQ ID NO:50)

CD2BstXI-2244 5' ATAGATTGCTCCAAACACTTGGTGG (SEQ ID NO:51)

JE-2281 5' ACTCCATAGGAAAAGCCGTTACC (SEQ ID NO:52)

CSP6 5' GCGAGCTCTAGCATTTAGGTGACACTATAG (SEQ ID NO:53)

DEN-2 ↔ JE

90-10 Junction: Leu His Gln Val Phe Gly Gly Ala Phe Arg Thr (SEQ ID NO:55)

CTC CAC CAA GTG TTT GGT GGT GCC TTC AGA ACA (SEQ ID NO:54)

80% DEN-2 E & 20% JE E:

T7 5' CTTATCGAAATTAATACGACTCACTATAGG (SEQ ID NO:56)

CD2BsmBI-2097 5' GAATTCTGCTCACTTCCTTTCTTAAACCAGTTGAGCTTC (SEQ ID NO:57)

JE BsmBI-2175 5' GGAATTCTGCTCGGAAGCACGCTGGGCAAGG (SEQ ID NO:58)

CSP6 5' GCGAGCTCTAGCATTTAGGTGACACTATAG 3' (SEQ ID NO:59)

DEN-2 ↔ JE

80-20 Junction: Asn Trp Lys Lys Gly Ser Thr Leu Gly Lys Ala (SEQ ID NO:61)

AAC TGG TTT AAG AAA GGA AGC ACG CTG GGC GCC (SEQ ID NO:60)

^a Restriction enzyme sites encoded in oligonucleotides were highlighted by the bold, italic and underline.

Table 10. Characterization of DEN-2 E glycoprotein epitopes expressed by the recombinant DEN-2 plasmids as determined by the indirect fluorescent antibody assay (IFA).

Antibody		Controls ^a		Plasmid construct ^a		
MAb (Epitope) ^b	Antigenic Domain ^c	PRNT ^d	DEN-2 infected cells	Normal cells	pCBD2-1J-4-3	pCB8D2-2J-2-9-1
4G2 (A1)	2	+/-	4+	-	4+	4+
4E5 (A2)	2	Yes	3+	-	3-4+	2-3+
1B7 (A5)	2	Yes	3-4+	-	4+	2-3+
1B4C-2(C1)	1	No	3-4+ (8000)	-	2-3+ (4000)	2-3+ (8000)
2B3A-1 (C3)	1	No	3-4+ (≥3200)	-	3+ (100)	2-3+ (≥3200)
9A4D-1 (C4)	1	No	3-4+	-	2-3+ (400)	3+ (≥12800)
3H5 (B2)	3	Yes	4+	-	4+	4+
10A4D-2 (B3)	3	Yes	2-3+	-	3-4+	2-3+
1A1D-2 (B4)	3	Yes	4+	-	3-4+	3-4+
9D12-6		Yes	2-4+		2-3+	3-4+
2H2	prM	No	4+	-	3-4+	3-4+
1A2A-1	Capsid	No	2-3+	-	1+	1-2+

^aTFA substrates were acetone-fixed COS-1 cells, either infected with DEN-2 16681, uninfected controls, or transformed with a DEN-2 recombinant plasmid.

^bMonoclonal antibodies were used at a predetermined optimum dilution based on reactivity with DEN-2 16681 virus. For some

MAbs, endpoint titers, shown in parentheses, are reported and for others, only qualitative values are reported based on a scale from 1+ to 4+, with 3-4+ considered positive, 2+ equivocal, and 1+ negative.

^cAntigenic domains based on the E-glycoprotein of TBE virus (Mandl et al., "Antigenic structure of the flavivirus envelope protein E at the molecular level, using tick-borne encephalitis virus as a model," *J. Virol.* 63: 564-571 (1989); Rey et al., "The envelope glycoprotein from tick-borne encephalitis virus at 2 Å resolution," *Nature* 375: 291-298 (1995)).

^dPlaque-reduction neutralization activity at a 1:100 dilution of ascitic fluid, using a 90% plaque-reduction endpoint, except for 4G2 and 9D12-6, for which a 50% neutralization endpoint is reported (Henchal et al., "Epitopic analysis of antigenic determinants on the surface of dengue-2 virions using monoclonal antibodies," *Am. J. Trop. Med. Hyg.* 34: 162-169 (1985); Roehrig et al., "Monoclonal antibody mapping of the envelope glycoprotein of the dengue 2 virus, Jamaica," *Virology* 246: 317-328 (1998)).

Table 11. Detection of secreted and membrane-bound DEN-2 recombinant protein by antigen-capture ELISA.

Plasmid	Sample type	Endpoint ELISA titer
pCBD2-14-6	PEG-precipitated culture fluid ^a	<1:10
pCBD2-14-6	PEG-precipitated, ethanol-extracted culture fluid ^b	<1:20
pCBD2-14-6	Hydrophobic membrane protein preparation ^c	1:160
pCB9D2-1J-4-3	PEG-precipitated culture fluid ^a	<1:10
pCB9D2-1J-4-3	PEG-precipitated, ethanol-extracted culture fluid ^b	<1:20
pCB9D2-1J-4-3	Hydrophobic membrane protein preparation ^c	1:80
pCB8D2-2J-2-9-1	PEG-precipitated culture fluid ^a	1:640
pCB8D2-2J-2-9-1	PEG-precipitated, ethanol-extracted culture fluid ^b	1:80
pCB8D2-2J-2-9-1	Hydrophobic membrane protein preparation ^c	1:80
pEGFP	PEG-precipitated culture fluid ^a	<1:10
pEGFP	PEG-precipitated, ethanol-extracted culture fluid ^b	<1:10
pEGFP	Hydrophobic membrane protein preparation ^c	<1:10

^aCulture supernatant from plasmid-transformed cells was precipitated with 10% polyethylene glycol (PEG) and resuspended in 1/100th of original volume.

^bPEG-precipitated culture supernatant was extracted with 4% ethanol to remove PEG and the pellet was resuspended in 1/5 of the volume extracted.

^cHydrophobic membrane fractions were prepared as described in Materials and Methods.

Table 12. Immunogenicity of three DEN-2 recombinant plasmids in ICR mice.

Plasmid DNA ^b	Mouse #	ELISA on DEN-2 virus				Endpoint titer 9 wks, p.v.	PRNT on DEN-2 virus ^a	PRNT on JE virus ^a
		Screen 3 wks, p.v. ^c		Screen 6 wks, p.v. ^c			Endpoint titer 9 wks, p.v.	Endpoint titer 9 wks, p.v.
		1:100	1:400	1:100	1:400			
pCB8D2-2J-2-9-1	Pool, 1,2,4-10	ND ^d	ND	+	+	64,000	ND	ND
	1	+	+	+	+	64,000	>1000	<2
	2	+	+	+	+	32,000	>1000	<2
	4	+	+	+	+	16,000	200	<2
	5	+	+	+	+	4,000	<10	<2
	6	+	+	+	+	16,000	200	<2
	7	+	-	+	+	64,000	100	<2
	8	+	-	+	+	8,000	40	<2
	9	+	+	+	+	6,400	<2	<4
	10	+	+	+	+	64,000	>1000	<2
pCB9D2-1J-4-3	Pool, 1-10	ND	ND	+	+	1,000	ND	<2 ^e
	1	-	-	+	-	400	<10	ND
	2	+	-	+	+	200	<10	ND
	3	+	+	+	+	4,000	<2	≤4
	4	+	-	+	-	200	<10	ND
	5	-	-	+	+	400	<10	ND
	6	+	+	+	+	4,000	<2	2
	7	-	+/-	-	-	100	<10	ND
	8	-	-	-	-	200	<10	ND
	9	+	-	+	-	4,000	<2	<2
	10	-	-	+	+	4,000	<2	<2
pCBD2-14-6	Pool, 1-10	ND	ND	+	-	200	<2 ^f	<2 ^g
	1	-	-	-	-	400	<10	ND
	2,3,6-9	-	-	-	-	<100	ND	ND
	4	+	+	+	+	1,000	<2	<2
	5	-	-	+	-	2,000	8	<2
	10	+	-	-	-	<100	ND	ND
pEGFP	Pool, 1-10	-	ND	-	ND	<100	<2	<2

^aPRNT, plaque-reduction neutralization test, 90% neutralization endpoint.

^bMice were immunized intramuscularly with 100 μ g plasmid DNA on weeks 0 and 3.

^cELISA screens used sera diluted 1:100 and 1:400.

^dND, not done.

^ePool, 1,2,4,5,7,8.

^fPool, 2,3,6-10.

^gPool, 1-3,6-10.

Table 13. Summary of the characteristics of three DEN-2 recombinant plasmids.

Plasmid	IFA ^a		Ag-capture ELISA titer		ELISA titer on DEN-2 ^b		DEN-2 PRNT ^c
	+/-	Globular/Diffuse	Secreted antigen	Hydrophobic membrane protein prep	No. sera ≥ 1:100	Pooled sera titer	No. sera ≥ 1:10
pCB8D2-2J-2-9-1	+	Diffuse	1:640	1:80	9/9	1:64000	7/9 ^d
pCB9D2-1J-4-3	+	Globular	<1:10	1:80	10/10	1:1000	0/10
pCBD2-14-6	+	Globular	<1:10	1:160	3/10	1:200	0/10

^aIndirect fluorescent antibody assay (IFA) staining characteristics, + or –, and diffuse or globular pattern.

^bAnti-DEN-2 ELISA titer of sera from mice immunized with the recombinant plasmids.

Sera were collected 9 weeks post-vaccination (weeks 0 and 3). Number of mice with titer of ≥ 1:100/ total number of mice is shown, including the endpoint ELISA titer of the pooled serum sample.

^cNumber of mice with plaque-reduction neutralization titers (PRNT, 90% reduction) ≥ 1:10/ total number of mice. Sera were collected 9 weeks post-vaccination.

^dOf the 7 mice with neutralizing antibody, 3 mice had PRNT titers of >1:1000, 3 had titers of ≥1:100<1:1000, and one had a titer of 1:40.

What is claimed is:

1. An isolated nucleic acid comprising a transcriptional unit encoding a signal sequence of a structural protein of a first flavivirus and an immunogenic flavivirus antigen of a second flavivirus, wherein the transcriptional unit directs the synthesis of the antigen.
2. The nucleic acid of claim 1, wherein the signal sequence is a Japanese encephalitis virus signal sequence.
3. The nucleic acid of claim 1, wherein the immunogenic flavivirus antigen is of a flavivirus selected from the group consisting of yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.
4. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of West Nile virus.
5. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of yellow fever virus.
6. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of St. Louis encephalitis virus.
7. The nucleic acid of claim 1, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus and an M protein and an E protein of Powassan virus.
8. The nucleic acid of claim 1, wherein the antigen is selected from the group

consisting of an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus or any combination thereof.

9. The nucleic acid of claim 8, wherein the antigen is both the M protein and the E protein of a flavivirus.

10. The nucleic acid of claim 1, wherein the nucleic acid is DNA.

11. The nucleic acid of claim 10, comprising a nucleotide sequence selected from the group consisting of SEQ ID NO:15, SEQ ID NO:19, SEQ ID NO:21 and SEQ ID NO:23.

12. The nucleic acid of claim 1, wherein the transcriptional unit comprises a control sequence disposed appropriately such that it operably controls the synthesis of the antigen.

13. The nucleic acid of claim 12, wherein the control sequence is the cytomegalovirus immediate early promoter.

14. The nucleic acid of claim 1, comprising a Kozak consensus sequence located at a translational start site for a polypeptide comprising the antigen encoded by the TU.

15. The nucleic acid of claim 1 wherein the transcriptional unit comprises a poly-A terminator.

16. A cell comprising the nucleic acid of claim 1.

17. A composition comprising the nucleic acid of claim 1 and a pharmaceutically acceptable carrier.

18. A method of immunizing a subject against infection by a flavivirus, comprising administering to the subject an effective amount of the composition of claim 17.

19. The method of claim 18, wherein the flavivirus antigen is of a flavivirus selected from the group consisting of yellow fever virus, dengue serotype 1 virus, dengue serotype 2 virus, dengue serotype 3 virus, dengue serotype 4 virus, Japanese encephalitis virus, Powassan virus and West Nile virus.

20. The method of claim 18, wherein the antigen is selected from the group consisting of an M protein of a flavivirus, an E protein of a flavivirus, both an M protein and an E protein of a flavivirus, a portion of an M protein of a flavivirus, a portion of an E protein of a flavivirus and both a portion of an M protein of a flavivirus and a portion of an E protein of a flavivirus or any combination thereof.

21. The method of claim 20, wherein the antigen is both the M protein and the E protein of a flavivirus, and wherein a cell within the body of the subject, after incorporating the nucleic acid within it, secretes subviral particles comprising the M protein and the E protein.

22. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of West Nile virus.

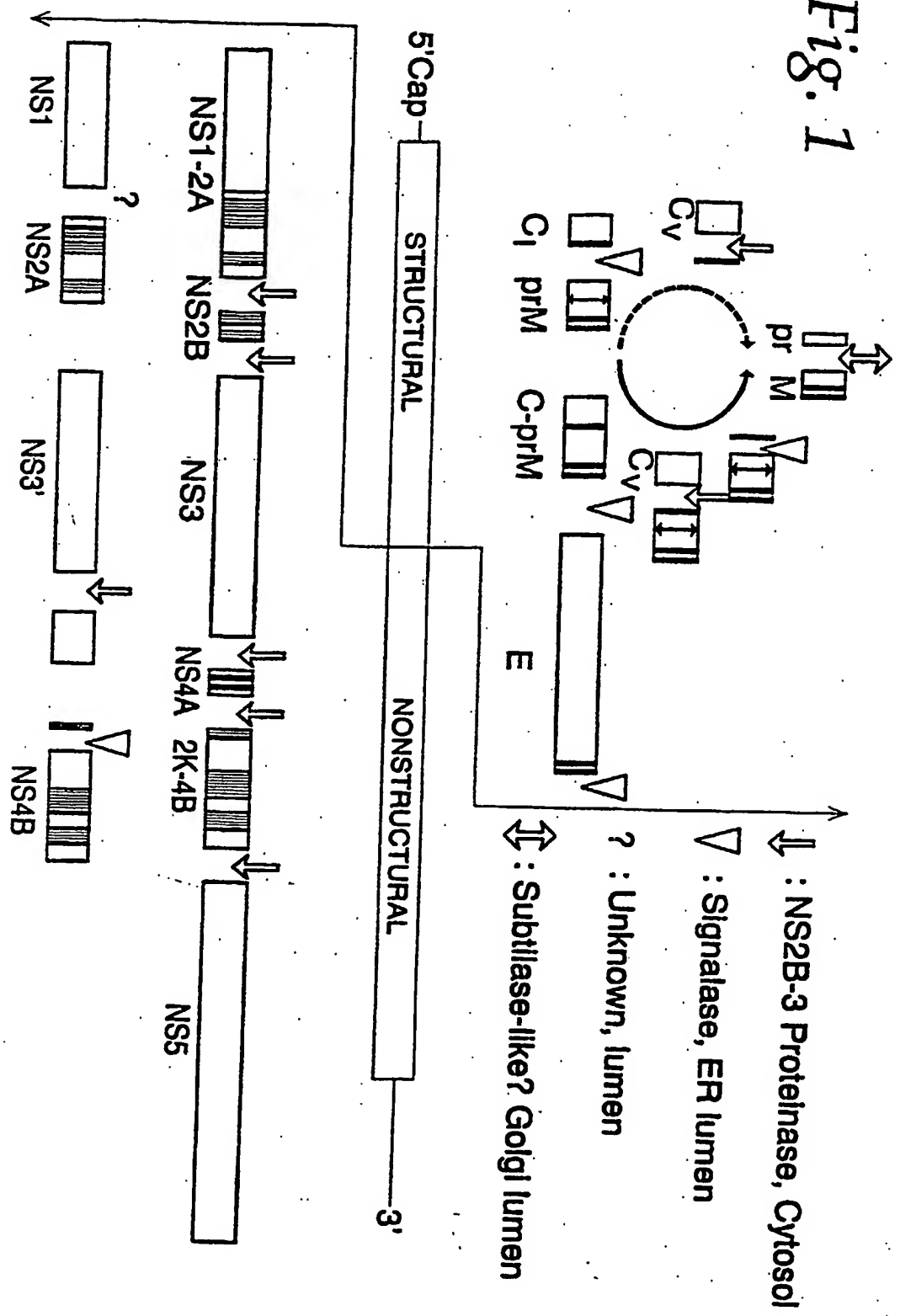
23. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of yellow fever virus.

24. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of St. Louis encephalitis virus.

25. The method of claim 18, wherein the transcriptional unit encodes a signal sequence of Japanese encephalitis virus, and an M protein and an E protein of Powassan virus.
26. The method of claim 18, comprising administering the composition to the subject in a single dose.
27. The method of claim 18, wherein the composition is administered via a parenteral route.
28. The nucleic acid of claim 1, wherein the antigen is a St. Louis encephalitis virus antigen.
29. The method of claim 18, wherein the antigen is a St. Louis encephalitis virus antigen.
30. The nucleic acid of claim 1, wherein the antigen is a Japanese encephalitis virus antigen.
31. The method of claim 18, wherein the antigen is a Japanese encephalitis virus antigen.
32. The nucleic acid of claim 1, wherein the antigen is a yellow fever virus antigen.
33. The method of claim 18, wherein the antigen is a yellow fever virus antigen.
34. The nucleic acid of claim 1, wherein the antigen is a dengue virus antigen.
35. The method of claim 18, wherein the antigen is a dengue virus antigen.
36. The nucleic acid of claim 1, wherein the antigen is a West Nile virus antigen.

37. The method of claim 18, wherein the antigen is a West Nile virus antigen.
38. An antigen produced from the nucleic acid of claim 1.
39. A method of detecting a flavivirus antibody in a sample, comprising:
(a) contacting the sample with the antigen of claim 38 under conditions whereby an antigen/antibody complex can form; and
(b) detecting antigen/antibody complex formation, thereby detecting a flavivirus antibody in the sample.
40. An antibody produced in response to immunization by the antigen of claim 38.
41. A method of detecting a flavivirus antigen in a sample, comprising:
(a) contacting the sample with the antibody of claim 40 under conditions whereby an antigen/antibody complex can form; and
(b) detecting antigen/antibody complex formation, thereby detecting a flavivirus antigen in a sample.
42. A method of diagnosing a flavivirus infection in a subject, comprising:
(a) contacting a sample from the subject with the antigen of claim 38 under conditions whereby an antigen/antibody complex can form; and
(b) detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.
43. A method of diagnosing a flavivirus infection in a subject, comprising:
(a) contacting a sample from the subject with the antibody of claim 40 under conditions whereby an antigen/antibody complex can form; and
(b) detecting antigen/antibody complex formation, thereby diagnosing a flavivirus infection in a subject.

Fig. 1



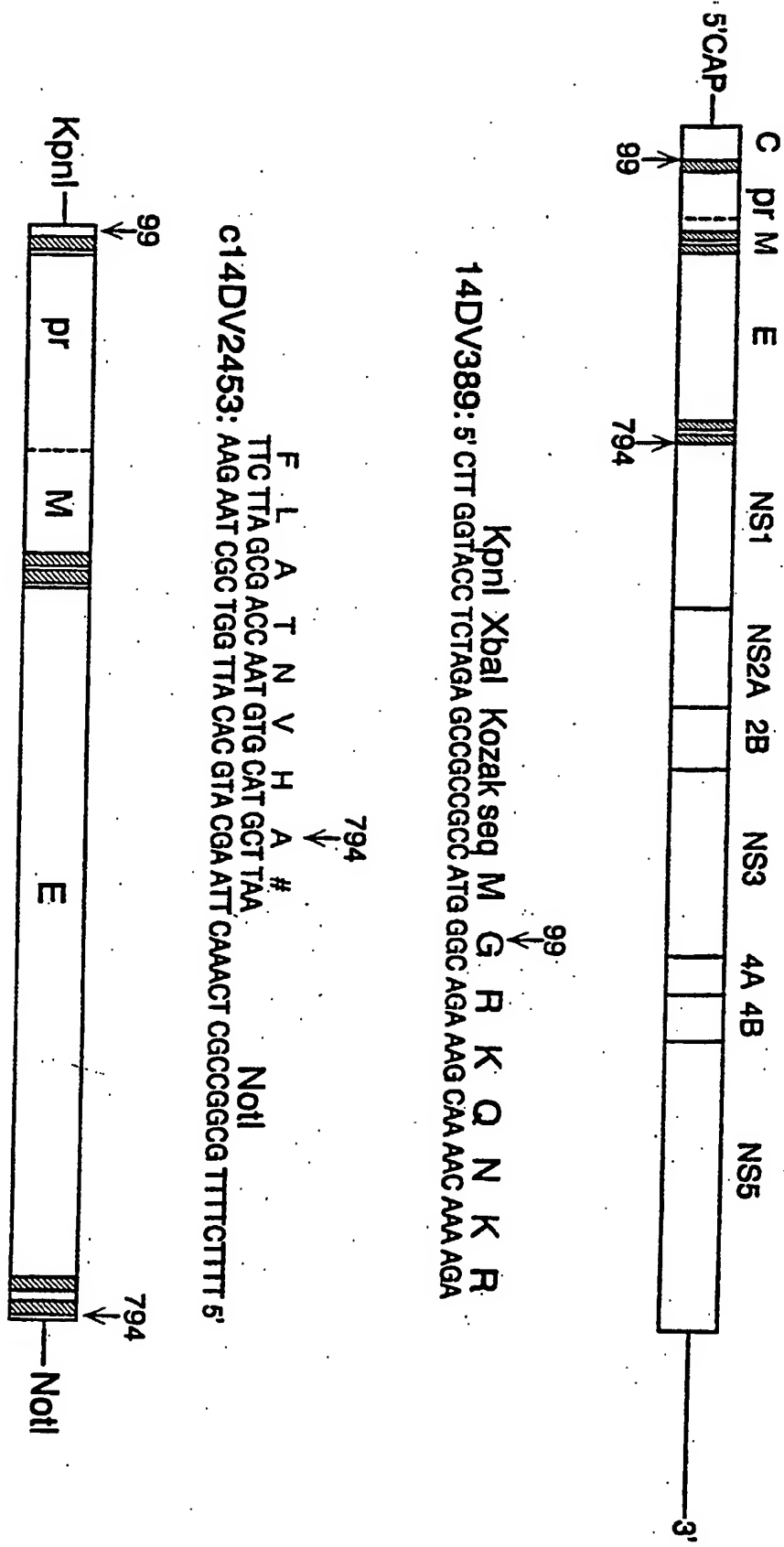


Fig. 2

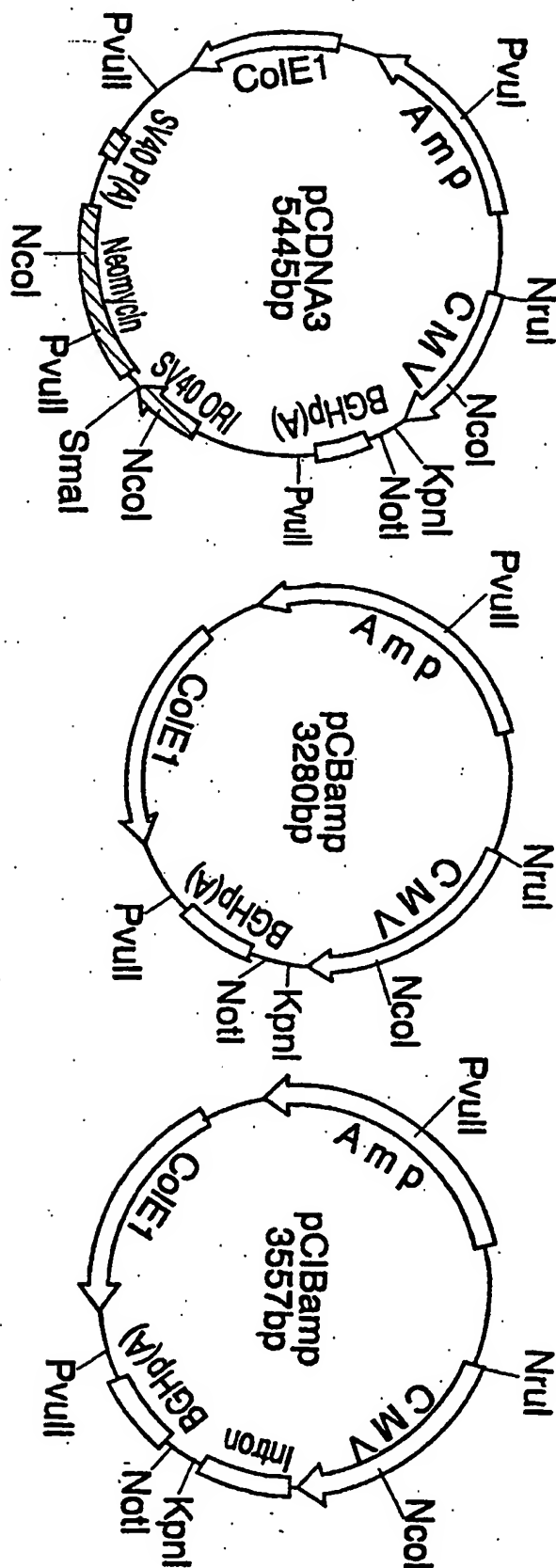
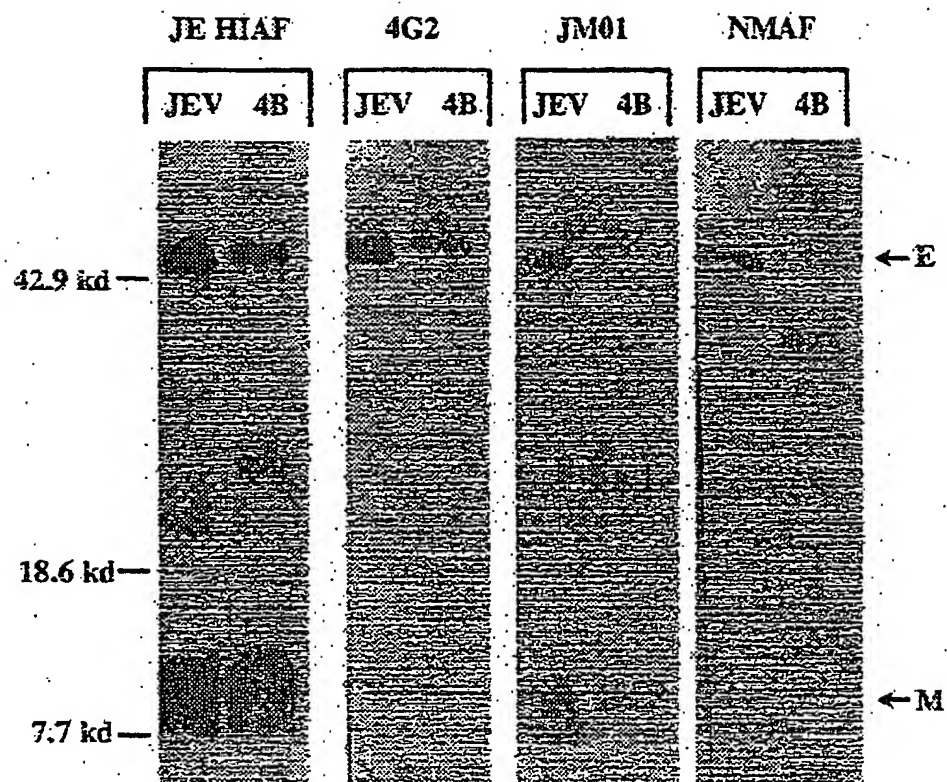


Fig. 3

**Fig. 4**

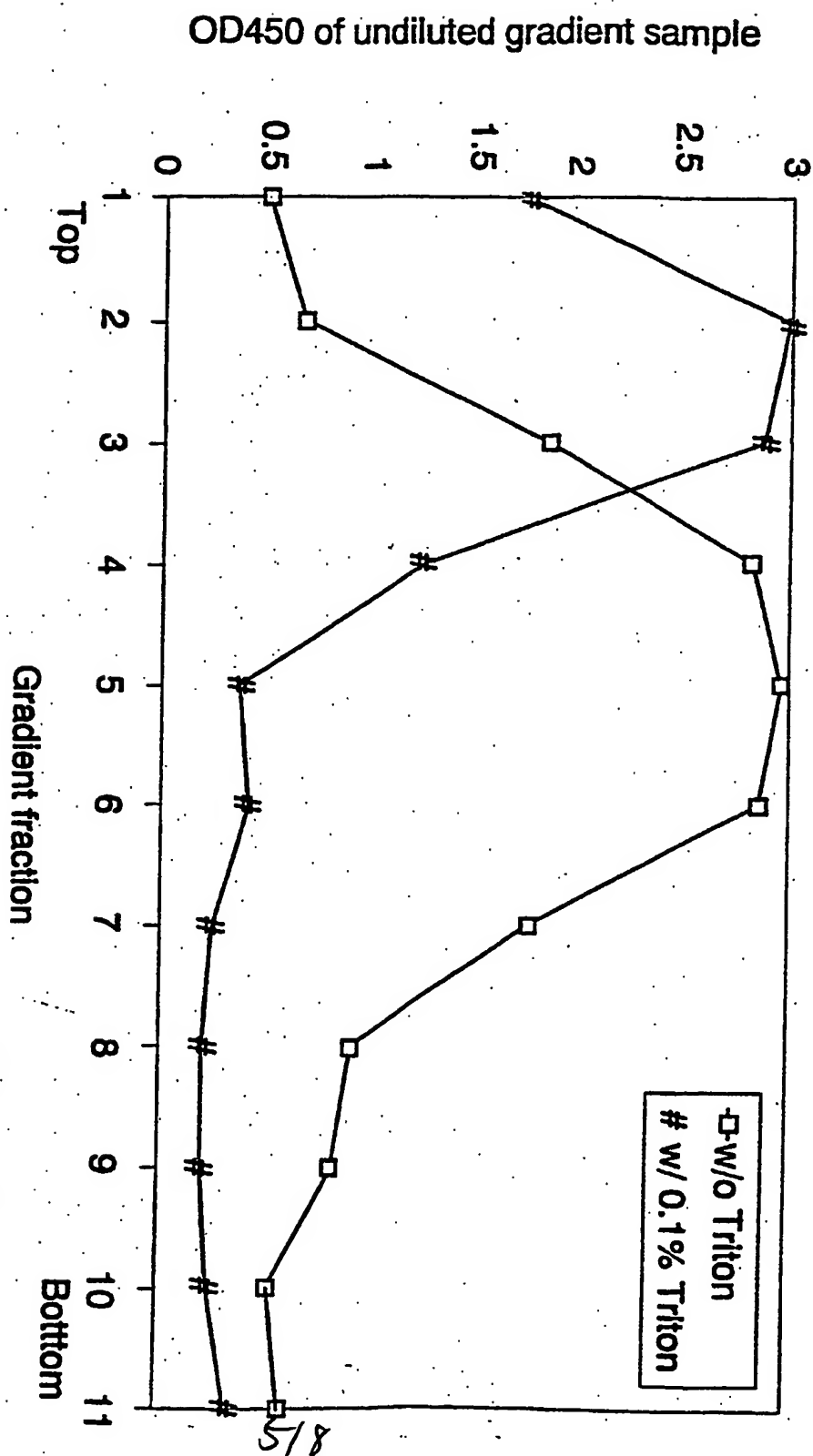
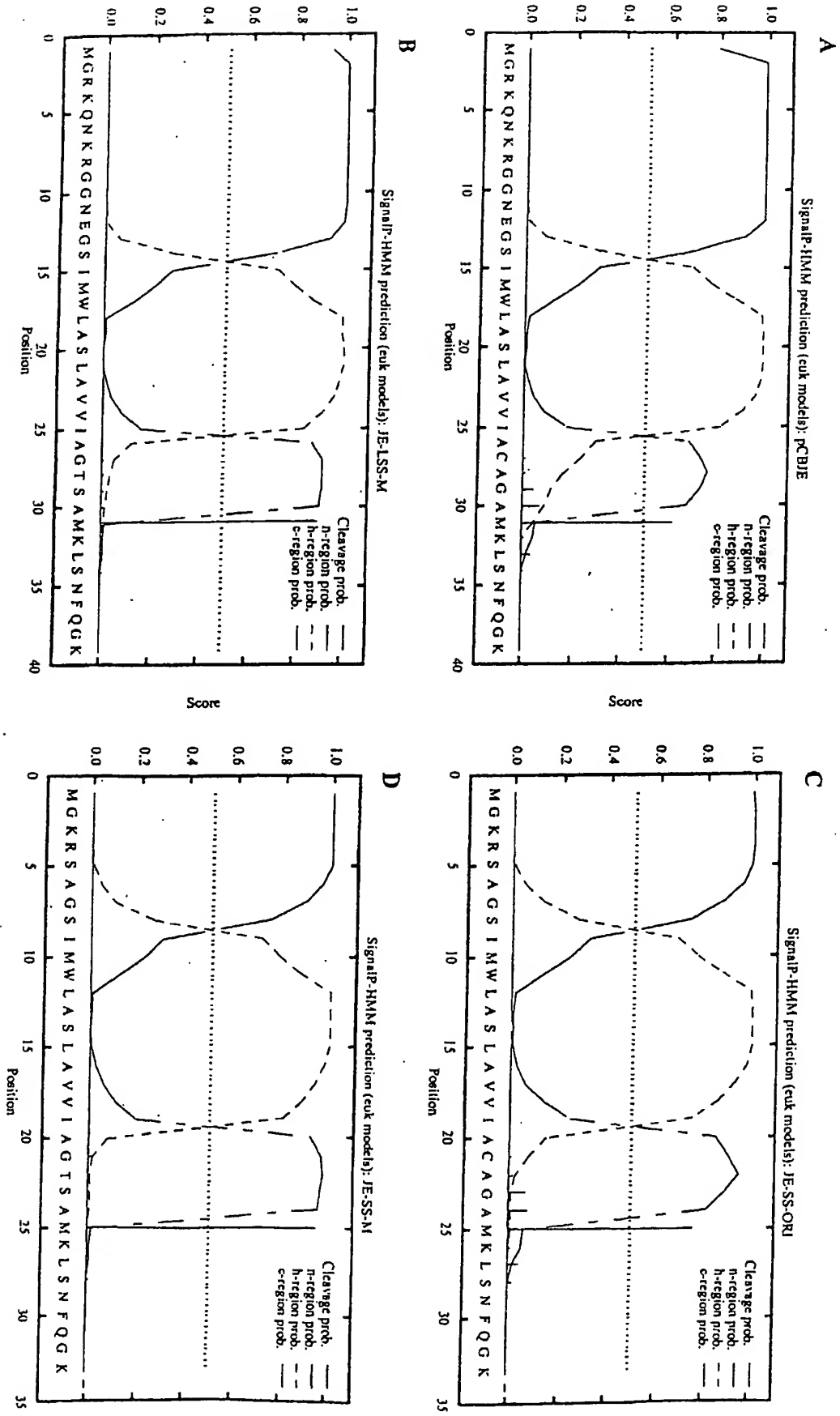


FIG. 6



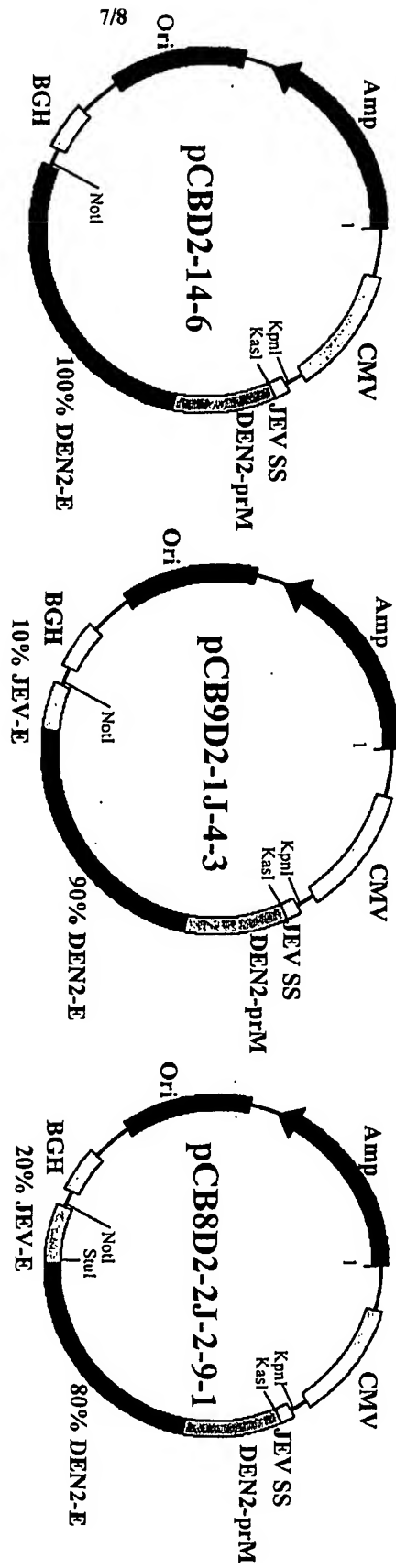


FIG. 7

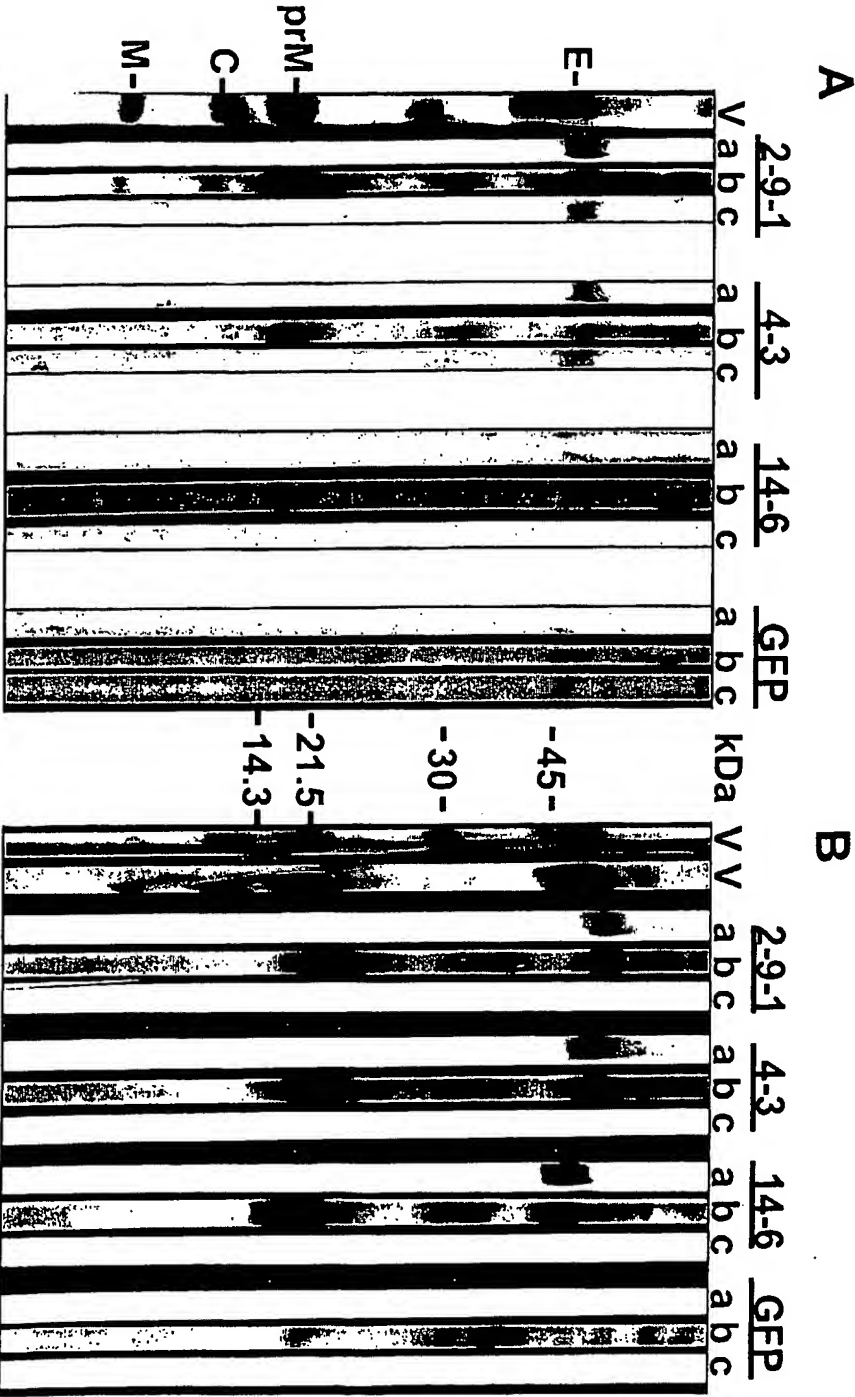


FIG. 8

SEQUENCE LISTING

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Chang, Gwong-Jen J

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<210> 11

<211> 697

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<223> pCDJE 2-7

<400> 11

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Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile Asn Asn Thr Asp
      35             40             45

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Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly Glu Asn Arg Cys
 50          55          60
Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu Asp Thr Ile Thr
65          70          75          80
Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro Glu Asp Val Asp
      85          90          95
Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr Gly Arg Cys Thr
      100          105          110
Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val Ser Val Gln Thr
      115          120          125
His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala Trp Leu Asp Ser
      130          135          140
Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn Trp Ile Ile Arg
145          150          155          160
Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly Trp Met Leu Gly
      165          170          175
Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu Leu Leu Leu Val
      180          185          190
Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly Asn Arg Asp Phe
      195          200          205
Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu Val Leu Glu Gly
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Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro Thr Leu Asp Val
225          230          235          240
Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu Val Arg Ser Tyr
      245          250          255
Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val Ala Arg Cys Pro
      260          265          270
Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp Ser Ser Tyr Val
      275          280          285
Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu
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Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe Ser Cys Thr Ser
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Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile Lys Tyr Glu Val
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Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn His Gly Asn Tyr
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Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe Thr Val Thr Pro
      355          360          365
Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr Gly Glu Val Thr
      370          375          380
Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu Ala Phe Tyr Val
385          390          395          400
Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg Glu Trp Phe His
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Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr Ala Trp Arg Asn
      420          425          430
Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala Thr Lys Gln Ser
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Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His Gln Ala Leu Ala
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Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys Leu Thr Ser Gly
465          470          475          480
His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala Leu Lys Gly Thr
      485          490          495
Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala Lys Asn Pro Ala
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Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser Tyr Ser Gly Ser
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Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala Ser Leu Asn Asp
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 Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu Pro Pro Phe Gly
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 Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln Ile Asn His His
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 Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe Ser Thr Thr Leu
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 Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe
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 Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys Ala Val His Gln
 625 630 635 640
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 Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met Gly Val Asn Ala
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 <212> DNA
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<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<221> misc_feature
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<210> 13
 <211> 43
 <212> DNA
 <213> Artificial Sequence

<220>
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<221> misc_feature
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 <223> CWN2444

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<223> Description of artificial sequence; note =
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<223> JE Signal

<400> 14

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			20												

<210> 15

<211> 5308

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

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<222> (911)...(2987)

<221> misc_feature

<222> (1)...(5308)

<223> pCBWN

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cccgccatt	gacgtcaata	atgacgtatg	ttcccatagt	aacgccaata	gggactttcc	420
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cgccgccgcc	atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg					949
	Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala					
	1 5 10					

agc	ttg	gca	gtt	gtc	ata	gct	tgt	gca	ggc	gcc	gtg	acc	ctc	tcg	aac	997
Ser	Leu	Ala	Val	Val	Ile	Ala	Cys	Ala	Gly	Ala	Val	Thr	Leu	Ser	Asn	
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ttc	cag	ggc	aag	gtg	atg	atg	acg	gta	aat	gct	act	gac	gtc	aca	gat	1045
Phe	Gln	Gly	Lys	Val	Met	Met	Thr	Val	Asn	Ala	Thr	Asp	Val	Thr	Asp	
	30				35				40					45		

gtc	atc	acg	att	cca	aca	gct	gct	gga	aag	aac	cta	tgc	att	gtc	aga	1093
Val	Ile	Thr	Ile	Pro	Thr	Ala	Ala	Gly	Lys	Asn	Leu	Cys	Ile	Val	Arg	
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Pro Val Leu Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys	
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Ser Thr Leu Ala Asn Lys Lys Gly Ala Trp Met Asp Ser Thr Lys Ala	
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Tyr Ala Leu Val Ala Ala Val Ile Gly Trp Met Leu Gly Ser Asn Thr	
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Asn Met Glu Ala Ala Asn Leu Ala Glu Val Arg Ser Tyr Cys Tyr Leu	
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290 295 300	

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ctc cct tgg agc agt gct gga agt act gtg tgg agg aac aga gag acg	2197
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Pro Val Glu Phe Ser Ser Asn Thr Val Lys Leu Thr Ser Gly His Leu	
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aag tgt aga gtg aag atg gaa aaa ttg cag ttg aag gga aca acc tat	2389
Lys Cys Arg Val Lys Met Glu Lys Leu Gln Leu Lys Gly Thr Thr Tyr	
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Gly His Gly Thr Val Val Leu Glu Leu Gln Tyr Thr Gly Thr Asp Gly	
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<210> 16

<211> 692

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<223> pCBWN

<400> 16

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Lys Val Met Met Thr Val Asn Ala Thr Asp Val Thr Asp Val Ile Thr
      35              40              45
Ile Pro Thr Ala Ala Gly Lys Asn Leu Cys Ile Val Arg Ala Met Asp
      50              55              60
Val Gly Tyr Met Cys Asp Asp Thr Ile Thr Tyr Glu Cys Pro Val Leu
      65              70              75              80
Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Thr Lys Ser
      85              90              95
Ala Val Tyr Val Arg Tyr Gly Arg Cys Thr Lys Thr Arg His Ser Arg
      100             105             110
Arg Ser Arg Arg Ser Leu Thr Val Gln Thr His Gly Glu Ser Thr Leu
      115             120             125
Ala Asn Lys Lys Gly Ala Trp Met Asp Ser Thr Lys Ala Thr Arg Tyr
      130             135             140
Leu Val Lys Thr Glu Ser Trp Ile Leu Arg Asn Pro Gly Tyr Ala Leu
      145             150             155             160
Val Ala Ala Val Ile Gly Trp Met Leu Gly Ser Asn Thr Met Gln Arg
      165             170             175
Val Val Phe Val Val Leu Leu Leu Leu Val Ala Pro Ala Tyr Ser Phe
      180             185             190
Asn Cys Leu Gly Met Ser Asn Arg Asp Phe Leu Glu Gly Val Ser Gly
      195             200             205
Ala Thr Trp Val Asp Leu Val Leu Glu Gly Asp Ser Cys Val Thr Ile
      210             215             220
Met Ser Lys Asp Lys Pro Thr Ile Asp Val Lys Met Met Asn Met Glu
      225             230             235             240

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 Ser Asp Leu Ser Thr Lys Ala Ala Cys Pro Thr Met Gly Glu Ala His
 260 265 270
 Asn Asp Lys Arg Ala Asp Pro Ala Phe Val Cys Arg Gln Gly Val Val
 275 280 285
 Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile
 290 295 300
 Asp Thr Cys Ala Lys Phe Ala Cys Ser Thr Lys Ala Ile Gly Arg Thr
 305 310 315 320
 Ile Leu Lys Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val His Gly
 325 330 335
 Pro Thr Thr Val Glu Ser His Gly Asn Tyr Ser Thr Gln Val Gly Ala
 340 345 350
 Thr Gln Ala Gly Arg Phe Ser Ile Thr Pro Ala Ala Pro Ser Tyr Thr
 355 360 365
 Leu Lys Leu Gly Glu Tyr Gly Glu Val Thr Val Asp Cys Glu Pro Arg
 370 375 380
 Ser Gly Ile Asp Thr Asn Ala Tyr Tyr Val Met Thr Val Gly Thr Lys
 385 390 395 400
 Thr Phe Leu Val His Arg Glu Trp Phe Met Asp Leu Asn Leu Pro Trp
 405 410 415
 Ser Ser Ala Gly Ser Thr Val Trp Arg Asn Arg Glu Thr Leu Met Glu
 420 425 430
 Phe Glu Glu Pro His Ala Thr Lys Gln Ser Val Ile Ala Leu Gly Ser
 435 440 445
 Gln Glu Gly Ala Leu His Gln Ala Leu Ala Gly Ala Ile Pro Val Glu
 450 455 460
 Phe Ser Ser Asn Thr Val Lys Leu Thr Ser Gly His Leu Lys Cys Arg
 465 470 475 480
 Val Lys Met Glu Lys Leu Gln Leu Lys Gly Thr Thr Tyr Gly Val Cys
 485 490 495
 Ser Lys Ala Phe Lys Phe Leu Gly Thr Pro Ala Asp Thr Gly His Gly
 500 505 510
 Thr Val Val Leu Glu Leu Gln Tyr Thr Gly Thr Asp Gly Pro Cys Lys
 515 520 525
 Val Pro Ile Ser Ser Val Ala Ser Leu Asn Asp Leu Thr Pro Val Gly
 530 535 540
 Arg Leu Val Thr Val Asn Pro Phe Val Ser Val Ala Thr Ala Asn Ala
 545 550 555 560
 Lys Val Leu Ile Glu Leu Glu Pro Pro Phe Gly Asp Ser Tyr Ile Val
 565 570 575
 Val Gly Arg Gly Glu Gln Gln Ile Asn His His Trp His Lys Ser Gly
 580 585 590
 Ser Ser Ile Gly Lys Ala Phe Thr Thr Thr Leu Lys Gly Ala Gln Arg
 595 600 605
 Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Val Gly Gly
 610 615 620
 Val Phe Thr Ser Val Gly Lys Ala Val His Gln Val Phe Gly Gly Ala
 625 630 635 640
 Phe Arg Ser Leu Phe Gly Gly Met Ser Trp Ile Thr Gln Gly Leu Leu
 645 650 655
 Gly Ala Leu Leu Trp Met Gly Ile Asn Ala Arg Asp Arg Ser Ile
 660 665 670
 Ala Leu Thr Phe Leu Ala Val Gly Gly Val Leu Leu Phe Leu Ser Val
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 Asn Val His Ala
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<210> 17

<211> 5334

<212> DNA
 <213> Artificial Sequence
 <220>
 <223> Description of artificial sequence; note =
 synthetic construct
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 <222> (1)...(5334)
 <223> pCBE 1-14

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 cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc 180
 ttaggggttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt 240
 gattattgac tagttattaa tagtaatcaa ttacggggtc attagttcat agcccatata 300
 tggagttccg cgttacataa cttacggtaa atggcccgcc tggctgaccg cccaacgacc 360
 cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccaata gggactttcc 420
 attgacgtca atgggtggag tatttacggt aaactgccc cttggcagta catcaagtgt 480
 atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt 540
 atgccagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca 600
 tcgctattac catggtgatg cgggttttggc agtacatcaa tgggcgtgga tagcggtttg 660
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 aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg 780
 gtaggcgtgt acggtgggag gtctatatata gcagagctct ctggctaact agagaaccca 840
 ctgcttactg gcttatcgaa attaatacga ctactatag ggagacccaa gcttgggtacc 900
 tctagagccg ccgcc atg ggc aga aag caa aac aaa aga gga gga aat gaa 951
 Met Gly Arg Lys Gln Asn Lys Arg Gly Gly Asn Glu
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 ggc tca atc atg tgg ctc gcg agc ttg gca gtt gtc ata gct tgt gcg 999
 Gly Ser Ile Met Trp Leu Ala Ser Leu Ala Val Val Ile Ala Cys Ala
 15 20 25
 gga gcc atg aag ttg tgc aat ttc cag ggg aag ctt ttg atg acc atc 1047
 Gly Ala Met Lys Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile
 30 35 40
 aac aac acg gac att gca gac gtt atc gtg att ccc acc tca aaa gga 1095
 Asn Asn Thr Asp Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly
 45 50 55 60
 gag aac aga tgc tgg gtc cgg gca atc gac gtc ggc tac atg tgt gag 1143
 Glu Asn Arg Cys Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu
 65 70 75
 gac act atc acg tac gaa tgt cct aag ctt acc atg ggc aat gat cca 1191
 Asp Thr Ile Thr Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro
 80 85 90
 gag gat gtg gat tgc tgg tgt gac aac caa gaa gtc tac gtc caa tat 1239
 Glu Asp Val Asp Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr
 95 100 105

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Gly Arg Cys Thr Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val	
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tcg gtc caa aca cat ggg gag agt tca cta gtg aat aaa aaa gag gct	1335
Ser Val Gln Thr His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala	
125 130 135 140	
tgg ctg gat tca acg aaa gcc aca cga tat ctc atg aaa act gag aac	1383
Trp Leu Asp Ser Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn	
145 150 155	
tgg atc ata agg aat cct ggc tat gct ttc ctg gcg gcg gta ctt ggc	1431
Trp Ile Ile Arg Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly	
160 165 170	
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Trp Met Leu Gly Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu	
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Leu Leu Leu Val Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly	
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Asn Arg Asp Phe Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu	
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Val Leu Glu Gly Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro	
225 230 235	
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Thr Leu Asp Val Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu	
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Val Arg Ser Tyr Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val	
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Ala Arg Cys Pro Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp	
270 275 280	
agt agc tat gtg tgc aaa caa ggc ttc act gac cgt ggg tgg ggc aac	1815
Ser Ser Tyr Val Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn	
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Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe	
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tcc tgc acc agt aaa gcg att ggg aga aca atc cag cca gaa aac atc	1911
Ser Cys Thr Ser Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile	
320 325 330	
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Lys Tyr Glu Val Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn	
335 340 345	

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gcg ttt tac gtc atg acc gtg ggg tca aag tca ttt ctg gtc cat agg Ala Phe Tyr Val Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg 400 405 410	2151
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aca aaa cag tcc gtt gtt gct ctt ggg tca cag gaa gga ggc ctc cat Thr Lys Gln Ser Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His 445 450 455 460	2295
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ctg aaa ggc aca acc tat ggc atg tgt aca gaa aaa ttc tcg ttc gcg Leu Lys Gly Thr Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala 495 500 505	2439
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ccc ccc ttc gga gac tcc tac atc gta gtt gga agg gga gac aag cag Pro Pro Phe Gly Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln 575 580 585	2679

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Ile Asn His His Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe	
590 595 600	
tca aca act ttg aag gga gct caa aga ctg gca gcg ttg ggc gac aca	2775
Ser Thr Thr Leu Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr	
605 610 615 620	
gcc tgg gac ttt ggc tct att gga ggg gtc ttc aac tcc ata gga aaa	2823
Ala Trp Asp Phe Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys	
625 630 635	
gcc gtt cac caa gtg ttt ggt ggt gcc ttc aga aca ctc ttt ggg gga	2871
Ala Val His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly	
640 645 650	
atg tct tgg atc aca caa ggg cta atg ggt gcc cta ctg ctc tgg atg	2919
Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met	
655 660 665	
ggc gtc aac gca cga gac cga tca att gct ttg gcc ttc tta gcc aca	2967
Gly Val Asn Ala Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr	
670 675 680	
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Gly Gly Val Leu Val Phe Leu Ala Thr Asn Val His Ala	
685 690 695	
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<210> 18

<211> 697

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<223> pCBJE 1-14

<400> 18

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Leu Ser Asn Phe Gln Gly Lys Leu Leu Met Thr Ile Asn Asn Thr Asp
35     40     45
Ile Ala Asp Val Ile Val Ile Pro Thr Ser Lys Gly Glu Asn Arg Cys
50     55     60
Trp Val Arg Ala Ile Asp Val Gly Tyr Met Cys Glu Asp Thr Ile Thr
65     70     75     80
Tyr Glu Cys Pro Lys Leu Thr Met Gly Asn Asp Pro Glu Asp Val Asp
85     90     95
Cys Trp Cys Asp Asn Gln Glu Val Tyr Val Gln Tyr Gly Arg Cys Thr
100    105    110
Arg Thr Arg His Ser Lys Arg Ser Arg Arg Ser Val Ser Val Gln Thr
115    120    125
His Gly Glu Ser Ser Leu Val Asn Lys Lys Glu Ala Trp Leu Asp Ser
130    135    140
Thr Lys Ala Thr Arg Tyr Leu Met Lys Thr Glu Asn Trp Ile Ile Arg
145    150    155    160
Asn Pro Gly Tyr Ala Phe Leu Ala Ala Val Leu Gly Trp Met Leu Gly
165    170    175
Ser Asn Asn Gly Gln Arg Val Val Phe Thr Ile Leu Leu Leu Val
180    185    190
Ala Pro Ala Tyr Ser Phe Asn Cys Leu Gly Met Gly Asn Arg Asp Phe
195    200    205
Ile Glu Gly Ala Ser Gly Ala Thr Trp Val Asp Leu Val Leu Glu Gly
210    215    220
Asp Ser Cys Leu Thr Ile Met Ala Asn Asp Lys Pro Thr Leu Asp Val
225    230    235    240
Arg Met Ile Asn Ile Glu Ala Ser Gln Leu Ala Glu Val Arg Ser Tyr
245    250    255
Cys Tyr His Ala Ser Val Thr Asp Ile Ser Thr Val Ala Arg Cys Pro
260    265    270
Thr Thr Gly Glu Ala His Asn Glu Lys Arg Ala Asp Ser Ser Tyr Val
275    280    285
Cys Lys Gln Gly Phe Thr Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu
290    295    300
Phe Gly Lys Gly Ser Ile Asp Thr Cys Ala Lys Phe Ser Cys Thr Ser
305    310    315    320
Lys Ala Ile Gly Arg Thr Ile Gln Pro Glu Asn Ile Lys Tyr Glu Val
325    330    335

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Gly Ile Phe Val His Gly Thr Thr Thr Ser Glu Asn His Gly Asn Tyr
 340 345 350
 Ser Ala Gln Val Gly Ala Ser Gln Ala Ala Lys Phe Thr Val Thr Pro
 355 360 365
 Asn Ala Pro Ser Ile Thr Leu Lys Leu Gly Asp Tyr Gly Glu Val Thr
 370 375 380
 Leu Asp Cys Glu Pro Arg Ser Gly Leu Asn Thr Glu Ala Phe Tyr Val
 385 390 395 400
 Met Thr Val Gly Ser Lys Ser Phe Leu Val His Arg Glu Trp Phe His
 405 410 415
 Asp Leu Ala Leu Pro Trp Thr Ser Pro Ser Ser Thr Ala Trp Arg Asn
 420 425 430
 Arg Glu Leu Leu Met Glu Phe Glu Glu Ala His Ala Thr Lys Gln Ser
 435 440 445
 Val Val Ala Leu Gly Ser Gln Glu Gly Gly Leu His Gln Ala Leu Ala
 450 455 460
 Gly Ala Ile Val Val Glu Tyr Ser Ser Ser Val Lys Leu Thr Ser Gly
 465 470 475 480
 His Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Ala Leu Lys Gly Thr
 485 490 495
 Thr Tyr Gly Met Cys Thr Glu Lys Phe Ser Phe Ala Lys Asn Pro Ala
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 Asp Thr Gly His Gly Thr Val Val Ile Glu Leu Ser Tyr Ser Gly Ser
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 Asp Gly Pro Cys Lys Ile Pro Ile Ala Ser Val Ala Ser Leu Asn Asp
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 Met Thr Pro Val Gly Arg Leu Val Thr Val Asn Pro Phe Val Ala Thr
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 Ser Ser Ala Ser Ser Lys Val Leu Val Glu Met Glu Pro Pro Phe Gly
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 Asp Ser Tyr Ile Val Val Gly Arg Gly Asp Lys Gln Ile Asn His His
 580 585 590
 Trp His Lys Ala Gly Ser Thr Leu Gly Lys Ala Phe Ser Thr Thr Leu
 595 600 605
 Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr Ala Trp Asp Phe
 610 615 620
 Gly Ser Ile Gly Gly Val Phe Asn Ser Ile Gly Lys Ala Val His Gln
 625 630 635 640
 Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile
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 Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met Gly Val Asn Ala
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<210> 19

<211> 5283

<212> DNA

<213> Artificial Sequence

<220>

 <223> Description of artificial sequence; note =
 synthetic construct

<221> CDS

<222> (910)...(2965)

<400> 19

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ttagggttag gcgtttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt	240
gattattgac tagttattaa tagtaataca ttacggggtc attagttcat agcccatata	300
tggagttccg cgttacataa cttacggtaa atggcccgcg tggctgaccg cccaacgacc	360
cccgccatt gacgtcaata atgacgtatg ttcccatagt aacgcccaata gggactttcc	420
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Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser	
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Leu Ala Val Val Ile Ala Gly Thr Ser Ala Thr Thr Ile His Arg Asp	
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Arg Glu Gly Tyr Met Val Met Arg Ala Ser Gly Arg Asp Ala Ala Ser	
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Gln Val Arg Val Gln Asn Gly Thr Cys Val Ile Leu Ala Thr Asp Met	
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Gly Lys Arg Ser Val Val Ile Pro Thr His Ala Gln Lys Asp Met Val	
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Thr Arg Val Glu Gly Trp Met Trp Lys Asn Lys Leu Leu Thr Ala Ala	
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Ile Val Ala Leu Ala Trp Leu Met Val Asp Ser Trp Met Ala Arg Val	
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Cys Thr His Leu Glu Asn Arg Asp Phe Val Thr Gly Thr Gln Gly Thr	
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Thr Arg Val Ser Leu Val Leu Glu Leu Gly Gly Cys Val Thr Ile Thr	
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225 230 235	
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Glu Ser Pro Ala Glu Thr Arg Glu Tyr Cys Leu His Ala Lys Leu Thr	
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Asn Thr Lys Val Glu Ala Arg Cys Pro Thr Thr Gly Pro Ala Thr Leu	
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Pro Glu Glu His Gln Ala Asn Met Val Cys Lys Arg Asp Gln Ser Asp	
275 280 285	
cgt gga tgg gga aac cac tgc ggg ttt ttt ggg aag ggc agt ata gtg	1815
Arg Gly Trp Gly Asn His Cys Gly Phe Phe Gly Lys Gly Ser Ile Val	
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gct tgt gca aag ttt gaa tgc gag gaa gca aaa aaa gct gtg ggc cac	1863
Ala Cys Ala Lys Phe Glu Cys Glu Glu Ala Lys Lys Ala Val Gly His	
305 310 315	
gtc tat gac tcc aca aag atc acg tat gtt gtc aag gtt gag ccc cac	1911
Val Tyr Asp Ser Thr Lys Ile Thr Tyr Val Val Lys Val Glu Pro His	
320 325 330	
aca ggg gat tac ttg gct gca aat gag acc aat tca aac agg aaa tca	1959
Thr Gly Asp Tyr Leu Ala Ala Asn Glu Thr Asn Ser Asn Arg Lys Ser	
335 340 345 350	
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Ala Gln Phe Thr Val Ala Ser Glu Lys Val Ile Leu Arg Leu Gly Asp	
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Tyr Gly Asp Val Ser Leu Thr Cys Lys Val Ala Ser Gly Ile Asp Val	
370 375 380	
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Ala Gln Thr Val Val Met Ser Leu Asp Ser Ser Lys Asp His Leu Pro	
385 390 395	
tct gca tgg caa gtg cac cgt gac tgg ttt gag gac ttg gcg ctg ccc	2151
Ser Ala Trp Gln Val His Arg Asp Trp Phe Glu Asp Leu Ala Leu Pro	
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Trp Lys His Lys Asp Asn Gln Asp Trp Asn Ser Val Glu Lys Leu Val	
415 420 425 430	

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Ser Val Glu Gly Gln Lys Tyr His Leu Lys Ser Gly His Val Thr Cys	
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Asp Val Gly Leu Glu Lys Leu Lys Leu Lys Gly Thr Thr Tyr Ser Met	
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Cys Asp Lys Ala Lys Phe Lys Trp Lys Arg Val Pro Val Asp Ser Gly	
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His Asp Thr Val Val Met Glu Val Ser Tyr Thr Gly Ser Asp Lys Pro	
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Cys Arg Ile Pro Val Arg Ala Val Ala His Gly Val Pro Ala Val Asn	
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 675 680 685

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<211> 681

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
 synthetic construct

<400> 20

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 Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr Phe Ser
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Cys	Pro	Asp	Ser	Met	Glu	Tyr	Asn	Cys	Pro	Asn	Leu	Ser	Pro	Arg	Glu
65					70					75					80
Glu	Pro	Asp	Asp	Ile	Asp	Cys	Trp	Cys	Tyr	Gly	Val	Glu	Asn	Val	Arg
				85					90					95	
Val	Ala	Tyr	Gly	Lys	Cys	Asp	Ser	Ala	Gly	Arg	Ser	Arg	Arg	Ser	Arg
			100					105					110		
Arg	Ala	Ile	Asp	Leu	Pro	Thr	His	Glu	Asn	His	Gly	Leu	Lys	Thr	Arg
		115					120					125			
Gln	Glu	Lys	Trp	Met	Thr	Gly	Arg	Met	Gly	Glu	Arg	Gln	Leu	Gln	Lys
		130				135					140				
Ile	Glu	Arg	Trp	Phe	Val	Arg	Asn	Pro	Phe	Phe	Ala	Val	Thr	Ala	Leu
145				150					155						160
Thr	Ile	Ala	Tyr	Leu	Val	Gly	Ser	Asn	Met	Thr	Gln	Arg	Val	Val	Ile
				165					170					175	
Ala	Leu	Leu	Val	Leu	Ala	Val	Gly	Pro	Ala	Tyr	Ser	Ala	His	Cys	Ile
			180					185					190		
Gly	Ile	Thr	Asp	Arg	Asp	Phe	Ile	Glu	Gly	Val	His	Gly	Gly	Thr	Trp
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Val	Ser	Ala	Thr	Leu	Glu	Gln	Asp	Lys	Cys	Val	Thr	Val	Met	Ala	Pro
		210				215					220				
Asp	Lys	Pro	Ser	Leu	Asp	Ile	Ser	Leu	Glu	Thr	Val	Ala	Ile	Asp	Arg
225				230					235					240	
Pro	Ala	Glu	Val	Arg	Lys	Val	Cys	Tyr	Asn	Ala	Val	Leu	Thr	His	Val
				245					250					255	
Lys	Ile	Asn	Asp	Lys	Cys	Pro	Ser	Thr	Gly	Glu	Ala	His	Leu	Ala	Glu
		260						265					270		
Glu	Asn	Glu	Gly	Asp	Asn	Ala	Cys	Lys	Arg	Thr	Tyr	Ser	Asp	Arg	Gly
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Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys	Gly	Ser	Ile	Val	Ala	Cys
		290				295					300				
Ala	Lys	Phe	Thr	Cys	Ala	Lys	Ser	Met	Ser	Leu	Phe	Glu	Val	Asp	Gln
305				310					315					320	
Thr	Lys	Ile	Gln	Tyr	Val	Ile	Arg	Ala	Gln	Leu	His	Val	Gly	Ala	Lys
				325					330					335	
Gln	Glu	Asn	Trp	Thr	Thr	Asp	Ile	Lys	Thr	Leu	Lys	Phe	Asp	Ala	Leu
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Ser	Gly	Ser	Gln	Glu	Val	Glu	Phe	Ile	Gly	Tyr	Gly	Lys	Ala	Thr	Leu
		355					360					365			
Glu	Cys	Gln	Val	Gln	Thr	Ala	Val	Asp	Phe	Gly	Asn	Ser	Tyr	Ile	Ala
		370				375					380				
Glu	Met	Glu	Thr	Glu	Ser	Trp	Ile	Val	Asp	Arg	Gln	Trp	Ala	Gln	Asp
385				390					395					400	
Leu	Thr	Leu	Pro	Trp	Gln	Ser	Gly	Ser	Gly	Gly	Val	Trp	Arg	Glu	Met
				405					410					415	
His	His	Leu	Val	Glu	Phe	Glu	Pro	Pro	His	Ala	Ala	Thr	Ile	Arg	Val
		420							425				430		
Leu	Ala	Leu	Gly	Asn	Gln	Glu	Gly	Ser	Leu	Lys	Thr	Ala	Leu	Thr	Gly
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		450				455					460				
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Lys	Gly	Thr	Ser	Tyr	Lys	Ile	Cys	Thr	Asp	Lys	Met	Phe	Phe	Val	Lys
				485					490					495	
Asn	Pro	Thr	Asp	Thr	Gly	His	Gly	Thr	Val	Val	Met	Gln	Val	Lys	Val
		500						505					510		
Ser	Lys	Gly	Ala	Pro	Cys	Arg	Ile	Pro	Val	Ile	Val	Ala	Asp	Asp	Leu
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Ser Thr Asn Asp Asp Glu Val Leu Ile Glu Val Asn Pro Pro Phe Gly
545                      550                      555                      560
Asp Ser Tyr Ile Ile Val Gly Arg Gly Asp Ser Arg Leu Thr Tyr Gln
565                      570                      575
Trp His Lys Glu Gly Ser Ser Ile Gly Lys Leu Phe Thr Gln Thr Met
580                      585                      590
Lys Gly Val Glu Arg Leu Ala Val Met Gly Asp Thr Ala Trp Asp Phe
595                      600                      605
Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile His Thr
610                      615                      620
Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn Trp Ile
625                      630                      635                      640
Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile Asn Thr
645                      650                      655
Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val Ile Met
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<210> 21

<211> 5304

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> CDS

<222> (910)...(2986)

<400> 21

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cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc      180
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ctgcttactg gcttatcgaa attaatacga ctactatag ggagacccaa gcttgggtacc      900
gcgcgcgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc      951
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser
1                      5                      10

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Leu Ala Val Val Ile Ala Gly Thr Ser Ala Leu Gln Leu Ser Thr Tyr
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gtg ctt tca gcg gga aat gat ccc gag gac att gac tgt tgg tgt gac Val Leu Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Asp 80 85 90	1191
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tcg agg cgt agc cga cgg tca atc tct gtg cag cat cat gga gat tcc Ser Arg Arg Ser Arg Arg Ser Ile Ser Val Gln His His Gly Asp Ser 115 120 125	1287
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gcc cta gtt gcg ctg gcg att gga tgg atg ctc ggt agc aac aac aca Ala Leu Val Ala Leu Ala Ile Gly Trp Met Leu Gly Ser Asn Asn Thr 160 165 170	1431
cag aga gtg gtt ttt gtg atc atg ctg atg ctg att gct ccg gca tac Gln Arg Val Val Phe Val Ile Met Leu Met Leu Ile Ala Pro Ala Tyr 175 180 185 190	1479
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Ser Ile Asp Thr Cys Ala Lys Phe Thr Cys Lys Asn Lys Ala Thr Gly	
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Lys Thr Ile Leu Arg Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val	
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His Gly Ser Thr Asp Ser Thr Ser His Gly Asn Tyr Ser Glu Gln Ile	
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Pro Trp Thr Ser Pro Ala Thr Thr Asp Trp Arg Asn Arg Glu Thr Leu	
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Gly Ser Gln Glu Gly Ala Leu His Thr Ala Leu Ala Gly Ala Ile Pro	
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Ala Thr Val Ser Ser Ser Thr Leu Thr Leu Gln Ser Gly His Leu Lys	
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Cys Arg Ala Lys Leu Asp Lys Val Lys Ile Lys Gly Thr Thr Tyr Gly	
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Asn Asn Lys Val Met Ile Glu Val Glu Pro Pro Phe Gly Asp Ser Tyr	
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Ile Val Val Gly Arg Gly Thr Thr Gln Ile Asn Tyr His Trp His Lys	
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Glu Gly Ser Ser Ile Gly Lys Ala Leu Ala Thr Thr Trp Lys Gly Ala	
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<211> 692

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
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<400> 22

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Lys Val Leu Met Ser Ile Asn Lys Thr Asp Ala Gln Ser Ala Ile Asn
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Ile Pro Ser Ala Asn Gly Ala Asn Thr Cys Ile Val Arg Ala Leu Asp
 50             55             60
Val Gly Val Met Cys Lys Asp Asp Ile Thr Tyr Leu Cys Pro Val Leu
 65             70             75             80
Ser Ala Gly Asn Asp Pro Glu Asp Ile Asp Cys Trp Cys Asp Val Glu
 85             90             95
Glu Val Trp Val His Tyr Gly Arg Cys Thr Arg Met Gly His Ser Arg
100            105            110
Arg Ser Arg Arg Ser Ile Ser Val Gln His His Gly Asp Ser Thr Leu
115            120            125
Ala Thr Lys Asn Thr Pro Trp Leu Asp Thr Val Lys Thr Thr Lys Tyr
130            135            140
Leu Thr Lys Val Glu Asn Trp Val Leu Arg Asn Pro Gly Tyr Ala Leu
145            150            155            160
Val Ala Leu Ala Ile Gly Trp Met Leu Gly Ser Asn Asn Thr Gln Arg
165            170            175
Val Val Phe Val Ile Met Leu Met Leu Ile Ala Pro Ala Tyr Ser Phe
180            185            190
Asn Cys Leu Gly Thr Ser Asn Arg Asp Phe Val Glu Gly Ala Ser Gly
195            200            205
Ala Thr Trp Ile Asp Leu Val Leu Glu Gly Gly Ser Cys Val Thr Val
210            215            220

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 Asp Thr Leu Ser Thr Val Ala Arg Cys Pro Thr Thr Gly Glu Ala His
 260 265 270
 Asn Thr Lys Arg Ser Asp Pro Thr Phe Val Cys Lys Arg Asp Val Val
 275 280 285
 Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile
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 Asp Thr Cys Ala Lys Phe Thr Cys Lys Asn Lys Ala Thr Gly Lys Thr
 305 310 315 320
 Ile Leu Arg Glu Asn Ile Lys Tyr Glu Val Ala Ile Phe Val His Gly
 325 330 335
 Ser Thr Asp Ser Thr Ser His Gly Asn Tyr Ser Glu Gln Ile Gly Lys
 340 345 350
 Asn Gln Ala Ala Arg Phe Thr Ile Ser Pro Gln Ala Pro Ser Phe Thr
 355 360 365
 Ala Asn Met Gly Glu Tyr Gly Thr Val Thr Ile Asp Cys Glu Ala Arg
 370 375 380
 Ser Gly Ile Asn Thr Glu Asp Tyr Tyr Val Phe Thr Val Lys Glu Lys
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 Ser Trp Leu Val Asn Arg Asp Trp Phe His Asp Leu Asn Leu Pro Trp
 405 410 415
 Thr Ser Pro Ala Thr Thr Asp Trp Arg Asn Arg Glu Thr Leu Val Glu
 420 425 430
 Phe Glu Glu Pro His Ala Thr Lys Gln Thr Val Val Ala Leu Gly Ser
 435 440 445
 Gln Glu Gly Ala Leu His Thr Ala Leu Ala Gly Ala Ile Pro Ala Thr
 450 455 460
 Val Ser Ser Ser Thr Leu Thr Leu Gln Ser Gly His Leu Lys Cys Arg
 465 470 475 480
 Ala Lys Leu Asp Lys Val Lys Ile Lys Gly Thr Thr Tyr Gly Met Cys
 485 490 495
 Asp Ser Ala Phe Thr Phe Ser Lys Asn Pro Thr Asp Thr Gly His Gly
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 Thr Val Ile Val Glu Leu Gln Tyr Thr Gly Ser Asn Gly Pro Cys Arg
 515 520 525
 Val Pro Ile Ser Val Thr Ala Asn Leu Met Asp Leu Thr Pro Val Gly
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 Arg Leu Val Thr Val Asn Pro Phe Ile Ser Thr Gly Gly Ala Asn Asn
 545 550 555 560
 Lys Val Met Ile Glu Val Glu Pro Pro Phe Gly Asp Ser Tyr Ile Val
 565 570 575
 Val Gly Arg Gly Thr Thr Gln Ile Asn Tyr His Trp His Lys Glu Gly
 580 585 590
 Ser Ser Ile Gly Lys Ala Leu Ala Thr Thr Trp Lys Gly Ala Gln Arg
 595 600 605
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 Val Phe Asn Ser Ile Gly Lys Ala Val His Gln Val Phe Gly Gly Ala
 625 630 635 640
 Phe Arg Thr Leu Phe Gly Gly Met Ser Trp Ile Thr Gln Gly Leu Leu
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 Gly Ala Leu Leu Leu Trp Met Gly Leu Gln Ala Arg Asp Arg Ser Ile
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 Ser Val Gln Ala
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 <211> 5271
 <212> DNA
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<220>
 <223> Description of artificial sequence; note =
 synthetic construct

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 Leu Ala Val Val Ile Ala Gly Thr Ser Ala Val Thr Leu Val Arg Lys
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 Asn Arg Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr
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ttc tct gtg ggc aca ggc aac tgc aca aca aac att ttg gaa gcc aag 1095
 Phe Ser Val Gly Thr Gly Asn Cys Thr Thr Asn Ile Leu Glu Ala Lys
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tac tgg tgc cca gac tca atg gaa tac aac tgt ccc aat ctc agt cca 1143
 Tyr Trp Cys Pro Asp Ser Met Glu Tyr Asn Cys Pro Asn Leu Ser Pro
 65 70 75

aga gag gag cca gat gac att gat tgc tgg tgc tat ggg gtg gaa aac 1191
 Arg Glu Glu Pro Asp Asp Ile Asp Cys Trp Cys Tyr Gly Val Glu Asn
 80 85 90

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 Val Arg Val Ala Tyr Gly Lys Cys Asp Ser Ala Gly Arg Ser Arg Arg
 95 100 105 110

tca aga agg gcc att gac ttg cct acg cat gaa aac cat ggt ttg aag 1287
 Ser Arg Arg Ala Ile Asp Leu Pro Thr His Glu Asn His Gly Leu Lys
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Asp Phe Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile	
610 615 620	
cat acg gtg ttt ggc tct gcc ttt cag ggg cta ttt ggc ggc ttg aac	2823
His Thr Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn	
625 630 635	
tgg ata aca aag gtc atc atg ggg gcg gta ctt ata tgg gtt ggc atc	2871
Trp Ile Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile	
640 645 650	
aac aca aga aac atg aca atg tcc atg agc atg atc ttg gta gga gtg	2919
Asn Thr Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val	
655 660 665 670	
atc atg atg ttt ttg tct cta gga gtt ggg gcg t gagcgggcgc	2963
Ile Met Met Phe Leu Ser Leu Gly Val Gly Ala	
675 680	
tcgagcatgc atctagaggg ccctatttcta tagtgtcacc taaatgctag agctcgctga	3023
tcagcctcga ctgtgccttc tagttgccag ccatctgttg tttgccctc ccccgtcct	3083
tccttgaccc tggaggtgc cactccact gtcttttct aataaaatga ggaaattgca	3143
tcgcattgtc tgagtaggtg tcattctatt ctggggggtg ggggtgggca ggacagcaag	3203
ggggaggatt gggaagacaa tagcaggcat gctggggatg cggtgggctc tatggcttct	3263
gaggcgga aacacgtgc attaatgaat cggccaacgc gcggggagag gcggtttgag	3323
tattgggcgc tcttcgcctt cctcgctcac tgactcgctg cgctcggtcg ttcggctgcg	3383
gcgagcggtg acgtggcact caaaggcggt aatacgggta tccacagaat caggggataa	3443
cgcaggaaa aacatgtgag caaaaggcca gcaaaaggcc aggaaccgta aaaaggccgc	3503
gttgctggcg tttttccata ggctccgccc ccctgacgag catcacaaaa atcgacgctc	3563
aagtacagag tggcgaaacc cgacaggact ataaagatac caggcggttc ccctggaag	3623
ctccctcgct cgctctctg ttccgacct gccgcttacc ggatacctgt ccgcctttct	3683
cccttcggga agcgtggcg tttctcaatg ctacgctgt aggtatctca gttcgggtgta	3743
ggctggtcgc tccaagctgg gctgtgtgca cgaaccccc gttcagcccg accgctgcgc	3803
cttatccggt aactatcgtc ttgagtc aa cccgtaaga cagacttat cgccactggc	3863
agcagccact ggtaacagga ttagcagagc gaggtatgta ggcgggtgcta cagagttctt	3923
gaagtgggtg cctaactacg gctacactag aaggacagta tttgggtatct gcgctctgct	3983
gaagccagtt accttcggaa aaagagttgg tagctcttga tccggcaaac aaaccaccgc	4043
tggtagcggg ggtttttttg tttgcaagca gcagattacg cgcagaaaaa aaggatctca	4103
agaagatcct ttgatctttt ctacggggtc tgacgctcag tggaaacgaaa actcacgtta	4163
agggattttg gtcattgagat tatcaaaaag gatcttcacc tagatccttt taaattaaaa	4223
atgaagtttt aaatcaatct aaagtatata tgagtaaact tgggtctgaca gttaccaatg	4283
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aatgataccg cgagaccac gctcaccggc tccagattta tcagcaataa accagccagc	4463
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ggcgtcaata cgggataata ccgcgccaca tagcagaact ttaaaagtgc tcatcattgg	4943
aaaacgttct tcggggcgaa aactctcaag gatcttaccg ctgttgagat ccagttcgat	5003
gtaaccact cgtgcaccca actgatcttc agcatctttt actttcacca gcgtttctgg	5063
gtgagcaaaa acaggaaggc aaaatgccgc aaaaaaggga ataaggcgca cacggaaatg	5123
ttgaatactc atactcttcc tttttcaata ttatgaagc atttatcagg gttattgtct	5183
catgagcggg tacatatttg aatgtattta gaaaaataaa caaatagggg ttccgcgcac	5243
atttccccga aaagtccac ctgacgtc	5271

<210> 24

<211> 681

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 24

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Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
 1           5           10           15
Val Val Ile Ala Gly Thr Ser Ala Val Thr Leu Val Arg Lys Asn Arg
      20           25           30
Trp Leu Leu Leu Asn Val Thr Ser Glu Asp Leu Gly Lys Thr Phe Ser
      35           40           45
Val Gly Thr Gly Asn Cys Thr Thr Asn Ile Leu Glu Ala Lys Tyr Trp
 50           55           60
Cys Pro Asp Ser Met Glu Tyr Asn Cys Pro Asn Leu Ser Pro Arg Glu
65           70           75           80
Glu Pro Asp Asp Ile Asp Cys Trp Cys Tyr Gly Val Glu Asn Val Arg
      85           90           95
Val Ala Tyr Gly Lys Cys Asp Ser Ala Gly Arg Ser Arg Arg Ser Arg
      100          105          110
Arg Ala Ile Asp Leu Pro Thr His Glu Asn His Gly Leu Lys Thr Arg
      115          120          125
Gln Glu Lys Trp Met Thr Gly Arg Met Gly Glu Arg Gln Leu Gln Lys
      130          135          140
Ile Glu Arg Trp Phe Val Arg Asn Pro Phe Phe Ala Val Thr Ala Leu
145          150          155          160
Thr Ile Ala Tyr Leu Val Gly Ser Asn Met Thr Gln Arg Val Val Ile
      165          170          175
Ala Leu Leu Val Leu Ala Val Gly Pro Ala Tyr Ser Ala His Cys Ile
      180          185          190
Gly Ile Thr Asp Arg Asp Phe Ile Glu Gly Val His Gly Gly Thr Trp
      195          200          205
Val Ser Ala Thr Leu Glu Gln Asp Lys Cys Val Thr Val Met Ala Pro
      210          215          220
Asp Lys Pro Ser Leu Asp Ile Ser Leu Glu Thr Val Ala Ile Asp Arg
225          230          235          240
Pro Ala Glu Val Arg Lys Val Cys Tyr Asn Ala Val Leu Thr His Val
      245          250          255
Lys Ile Asn Asp Lys Cys Pro Ser Thr Gly Glu Ala His Leu Ala Glu
      260          265          270
Glu Asn Glu Gly Asp Asn Ala Cys Lys Arg Thr Tyr Ser Asp Arg Gly
      275          280          285
Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Ser Ile Val Ala Cys
      290          295          300
Ala Lys Phe Thr Cys Ala Lys Ser Met Ser Leu Phe Glu Val Asp Gln
305          310          315          320
Thr Lys Ile Gln Tyr Val Ile Arg Ala Gln Leu His Val Gly Ala Lys
      325          330          335
Gln Glu Asn Trp Thr Thr Asp Ile Lys Thr Leu Lys Phe Asp Ala Leu
      340          345          350
Ser Gly Ser Gln Glu Val Glu Phe Ile Gly Tyr Gly Lys Ala Thr Leu
      355          360          365
Glu Cys Gln Val Gln Thr Ala Val Asp Phe Gly Asn Ser Tyr Ile Ala
      370          375          380
Glu Met Glu Thr Glu Ser Trp Ile Val Asp Arg Gln Trp Ala Gln Asp
385          390          395          400
Leu Thr Leu Pro Trp Gln Ser Gly Ser Gly Gly Val Trp Arg Glu Met
      405          410          415

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His His Leu Val Glu Phe Glu Pro Pro His Ala Ala Thr Ile Arg Val
 420 425 430
 Leu Ala Leu Gly Asn Gln Glu Gly Ser Leu Lys Thr Ala Leu Thr Gly
 435 440 445
 Ala Met Arg Val Thr Lys Asp Thr Asn Asp Asn Asn Leu Tyr Lys Leu
 450 455 460
 His Gly Gly His Val Ser Cys Arg Val Lys Leu Ser Ala Leu Thr Leu
 465 470 475 480
 Lys Gly Thr Ser Tyr Lys Ile Cys Thr Asp Lys Met Phe Phe Val Lys
 485 490 495
 Asn Pro Thr Asp Thr Gly His Gly Thr Val Val Met Gln Val Lys Val
 500 505 510
 Ser Lys Gly Ala Pro Cys Arg Ile Pro Val Ile Val Ala Asp Asp Leu
 515 520 525
 Thr Ala Ala Ile Asn Lys Gly Ile Leu Val Thr Val Asn Pro Ile Ala
 530 535 540
 Ser Thr Asn Asp Asp Glu Val Leu Ile Glu Val Asn Pro Pro Phe Gly
 545 550 555 560
 Asp Ser Tyr Ile Ile Val Gly Arg Gly Asp Ser Arg Leu Thr Tyr Gln
 565 570 575
 Trp His Lys Glu Gly Ser Ser Ile Gly Lys Leu Phe Thr Gln Thr Met
 580 585 590
 Lys Gly Val Glu Arg Leu Ala Val Met Gly Asp Thr Ala Trp Asp Phe
 595 600 605
 Ser Ser Ala Gly Gly Phe Phe Thr Ser Val Gly Lys Gly Ile His Thr
 610 615 620
 Val Phe Gly Ser Ala Phe Gln Gly Leu Phe Gly Gly Leu Asn Trp Ile
 625 630 635 640
 Thr Lys Val Ile Met Gly Ala Val Leu Ile Trp Val Gly Ile Asn Thr
 645 650 655
 Arg Asn Met Thr Met Ser Met Ser Met Ile Leu Val Gly Val Ile Met
 660 665 670
 Met Phe Leu Ser Leu Gly Val Gly Ala
 675 680

<210> 25

<211> 35

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature

<222> 1-35

<223> POW 454

<400> 25

aaaagaaaaa gcgctaccac catccaccgg gacag

35

<210> 26

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature

<222> 1-41
 <223> CPOW 2417

<400> 26
 actgttaccc tcaaccccggt actcgccggc gaaaaagaaa a 41

<210> 27
 <211> 24
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<223> Modified JE Signal

<400> 27
 Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser Leu Ala
 1 . 5 10 15
 Val Val Ile Ala Gly Thr Ser Ala
 20

<210> 28
 <211> 36
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<221> misc_feature
 <222> 1-36
 <223> YF 482

<400> 28
 aaaagaaaaa gcgctgtgac cttggtgcgg aaaaac 36

<210> 29
 <211> 41
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<221> misc_feature
 <222> 1-41
 <223> CYF 2433

<400> 29
 acagagatcc tcaaccccggt actcgccggc gaaaaagaaa a 41

<210> 30
 <211> 41
 <212> DNA
 <213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature

<222> 1-41

<223> SLE 463

<400> 30

aaaagaaaaa gcgctttgca gttatcaacc tatcagggga a

41

<210> 31

<211> 40

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> misc_feature

<222> 1-40

<223> CSLE 2477

<400> 31

accgttggtc gcacgttcgg actcgccggc gaaaaagaaa

40

<210> 32

<211> 39

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 32

Leu	Asp	Thr	Ile	Asn	Arg	Arg	Pro	Ser	Lys	Lys	Arg	Gly	Gly	Thr	Arg
1				5				10						15	
Ser	Leu	Leu	Gly	Leu	Ala	Ala	Leu	Ile	Gly	Leu	Ala	Ser	Ser	Leu	Gln
			20					25					30		
Leu	Leu	Ser	Thr	Tyr	Gln	Gly									
			35												

<210> 33

<211> 24

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 33

Met	Trp	Leu	Ala	Ser	Leu	Ala	Val	Val	Ile	Ala	Cys	Ala	Gly	Ala	Met
1				5					10					15	
Lys	Leu	Ser	Asn	Phe	Gln	Gly	Lys								
			20												

<210> 34

<211> 30

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 34

Met	Asn	Glu	Gly	Ser	Ile	Met	Trp	Leu	Ala	Ser	Leu	Ala	Val	Val	Ile
1				5				10					15		
Ala	Cys	Ala	Gly	Ala	Met	Lys	Leu	Ser	Asn	Phe	Gln	Gly	Lys		
			20				25						30		

<210> 35

<211> 39

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 35

Met	Gly	Arg	Lys	Gln	Asn	Lys	Arg	Gly	Gly	Asn	Glu	Gly	Ser	Ile	Met
1				5				10					15		
Trp	Leu	Ala	Ser	Leu	Ala	Val	Val	Ile	Ala	Cys	Ala	Gly	Ala	Met	Lys
			20				25						30		
Leu	Ser	Asn	Phe	Gln	Gly	Lys									
			35												

<210> 36

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 36

Met	Ser	Lys	Lys	Arg	Gly	Gly	Ser	Glu	Thr	Ser	Val	Leu	Met	Val	Ile
1				5				10					15		
Phe	Met	Leu	Ile	Gly	Phe	Ala	Ala	Ala	Leu	Lys	Leu	Ser	Asn	Phe	Gln
			20				25						30		
Gly	Lys														

<210> 37

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 37

Met	Gly	Lys	Arg	Ser	Ala	Gly	Ser	Ile	Met	Trp	Leu	Ala	Ser	Leu	Ala
1				5				10					15		
Val	Val	Ile	Ala	Cys	Ala	Gly	Ala	Val	Thr	Leu	Ser	Asn	Phe	Gln	Gly
			20				25						30		
Lys															

<210> 38
 <211> 46
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<400> 38
 Met Asn Val Leu Arg Gly Phe Arg Lys Glu Ile Gly Arg Met Leu Asn
 1 5 10 15
 Ile Leu Asn Arg Arg Arg Thr Ala Gly Met Ile Ile Met Leu Ile
 20 25 30
 Pro Thr Val Met Ala Phe His Leu Thr Thr Arg Asn Gly Glu
 35 40 45

<210> 39
 <211> 40
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<400> 39
 Met Val Gly Leu Gln Lys Arg Gly Lys Arg Arg Ser Ala Thr Asp Trp
 1 5 10 15
 Met Ser Trp Leu Leu Val Ile Thr Leu Leu Gly Met Thr Leu Ala Ala
 20 25 30
 Thr Val Arg Lys Glu Arg Gly Asp
 35 40

<210> 40
 <211> 24
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<400> 40
 Met Gly Trp Leu Leu Val Val Val Leu Leu Gly Val Thr Leu Ala Ala
 1 5 10 15
 Thr Val Arg Lys Glu Arg Gly Asp
 20

<210> 41
 <211> 24
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of artificial sequence; note =
 synthetic construct

<400> 41

Met Ser Trp Leu Leu Val Ile Thr Leu Leu Gly Met Thr Ile Ala Ala
 1 5 10 15
 Thr Val Arg Lys Glu Arg Gly Asp
 20

<210> 42

<211> 5292

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
 synthetic construct

<221> CDS

<222> (910)...(2964)

<400> 42

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gacggatcgg gagatctccc gatccccctat ggtgcactct cagtacaatc tgctctgatg      60
ccgcatagtt aagccagtat ctgctccctg cttgtgtggt ggaggtcgct gagtagtgcg      120
cgagcaaaat ttaagctaca acaaggcaag gcttgaccga caattgcatg aagaatctgc      180
ttagggtttag gcgttttgcg ctgcttcgcg atgtacgggc cagatatacg cgttgacatt      240
gattattgac tagttattaa tagtaatcaa ttacgggggc attagttcat agcccatata      300
tggagttccg cgttacataa cttacggtaa atggcccggc tggctgaccg cccaacgacc      360
cccgcccatt gacgtcaata atgacgtatg ttcccatagt aacgccataa gggactttcc      420
attgacgtca atgggtggag tatttacggt aaactgcccc cttggcagta catcaagtgt      480
atcatatgcc aagtacgccc cctattgacg tcaatgacgg taaatggccc gcctggcatt      540
atgcccaagta catgacctta tgggactttc ctacttggca gtacatctac gtattagtca      600
tcgctattac catggtgatg cgggttttggc agtacatcaa tgggcgtgga tagcggtttg      660
actcacgggg atttccaagt tcaccacca ttgacgtcaa tgggagtttg ttttggcacc      720
aaaatcaacg ggactttcca aaatgtcgta acaactccgc cccattgacg caaatgggcg      780
gtaggcgtgt acggtgggag gtctatataa gcagagctct ctggctaact agagaacca      840
ctgcttactg gcttatcgaa attaatcaga ctcactatag ggagacccaa gcttgggtacc      900
gcccgcgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc      951
      Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser
            1                5                10

```

```

ttg gca gtt gtc ata gct tgt gca ggc gcc ttc cat tta acc aca cgt      999
Leu Ala Val Val Ile Ala Cys Ala Gly Ala Phe His Leu Thr Thr Arg
  15                20                25                30

```

```

aac gga gaa cca cac atg atc gtc agc aga caa gag aaa ggg aaa agt      1047
Asn Gly Glu Pro His Met Ile Val Ser Arg Gln Glu Lys Gly Lys Ser
            35                40                45

```

```

ctt ctg ttt aaa aca gag gat ggc gtg aac atg tgt acc ctc atg gcc      1095
Leu Leu Phe Lys Thr Glu Asp Gly Val Asn Met Cys Thr Leu Met Ala
            50                55                60

```

```

atg gac ctt ggt gaa ttg tgt gaa gac aca atc acg tac aag tgt ccc      1143
Met Asp Leu Gly Glu Leu Cys Glu Asp Thr Ile Thr Tyr Lys Cys Pro
            65                70                75

```

```

ctt ctc agg cag aat gag cca gaa gac ata gac tgt tgg tgc aac tct      1191
Leu Leu Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys Asn Ser
            80                85                90

```

```

acg tcc acg tgg gta act tat ggg acg tgt acc acc atg gga gaa cat      1239
Thr Ser Thr Trp Val Thr Tyr Gly Thr Cys Thr Thr Met Gly Glu His
  95                100                105                110

```

aga aga gaa aaa aga tca gtg gca ctc gtt cca cat gtg gga atg gga	1287
Arg Arg Glu Lys Arg Ser Val Ala Leu Val Pro His Val Gly Met Gly	
115 120 125	
ctg gag aca cga act gaa aca tgg atg tca tca gaa ggg gcc tgg aaa	1335
Leu Glu Thr Arg Thr Glu Thr Trp Met Ser Ser Glu Gly Ala Trp Lys	
130 135 140	
cat gtc cag aga att gaa act tgg atc ttg aga cat cca ggc ttc acc	1383
His Val Gln Arg Ile Glu Thr Trp Ile Leu Arg His Pro Gly Phe Thr	
145 150 155	
atg atg gca gca atc ctg gca tac acc ata gga acg aca cat ttc caa	1431
Met Met Ala Ala Ile Leu Ala Tyr Thr Ile Gly Thr Thr His Phe Gln	
160 165 170	
aga gcc ctg att ttc atc tta ctg aca gct gtc act cct tca atg aca	1479
Arg Ala Leu Ile Phe Ile Leu Leu Thr Ala Val Thr Pro Ser Met Thr	
175 180 185 190	
atg cgt tgc ata gga atg tca aat aga gac ttt gtg gaa ggg gtt tca	1527
Met Arg Cys Ile Gly Met Ser Asn Arg Asp Phe Val Glu Gly Val Ser	
195 200 205	
gga gga agc tgg gtt gac ata gtc tta gaa cat gga agc tgt gtg acg	1575
Gly Gly Ser Trp Val Asp Ile Val Leu Glu His Gly Ser Cys Val Thr	
210 215 220	
acg atg gca aaa aac aaa cca aca ttg gat ttt gaa ctg ata aaa aca	1623
Thr Met Ala Lys Asn Lys Pro Thr Leu Asp Phe Glu Leu Ile Lys Thr	
225 230 235	
gaa gcc aaa cag cct gcc acc cta agg aag tac tgt ata gag gca aag	1671
Glu Ala Lys Gln Pro Ala Thr Leu Arg Lys Tyr Cys Ile Glu Ala Lys	
240 245 250	
cta acc aac aca aca aca gaa tct cgc tgc cca aca caa ggg gaa ccc	1719
Leu Thr Asn Thr Thr Thr Glu Ser Arg Cys Pro Thr Gln Gly Glu Pro	
255 260 265 270	
agc cta aat gaa gag cag gac aaa agg ttc gtc tgc aaa cac tcc atg	1767
Ser Leu Asn Glu Glu Gln Asp Lys Arg Phe Val Cys Lys His Ser Met	
275 280 285	
gta gac aga gga tgg gga aat gga tgt gga cta ttt gga aag gga ggc	1815
Val Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Gly	
290 295 300	
att gtg acc tgt gct atg ttc aga tgc aaa aag aac atg gaa gga aaa	1863
Ile Val Thr Cys Ala Met Phe Arg Cys Lys Lys Asn Met Glu Gly Lys	
305 310 315	
gtt gtg caa cca gaa aac ttg gaa tac acc att gtg ata aca cct cac	1911
Val Val Gln Pro Glu Asn Leu Glu Tyr Thr Ile Val Ile Thr Pro His	
320 325 330	
tca ggg gaa gag cat gca gtc gga aat gac aca gga aaa cat ggc aag	1959
Ser Gly Glu Glu His Ala Val Gly Asn Asp Thr Gly Lys His Gly Lys	
335 340 345 350	

gaa atc aaa ata aca cca cag agt tcc atc aca gaa gca gaa ttg aca Glu Ile Lys Ile Thr Pro Gln Ser Ser Ile Thr Glu Ala Glu Leu Thr 355 360 365	2007
ggg tat ggc act gtc aca atg gag tgc tct cca aga acg ggc ctc gac Gly Tyr Gly Thr Val Thr Met Glu Cys Ser Pro Arg Thr Gly Leu Asp 370 375 380	2055
ttc aat gag atg gtg ttg ttg cag atg gaa aat aaa gct tgg ctg gtg Phe Asn Glu Met Val Leu Leu Gln Met Glu Asn Lys Ala Trp Leu Val 385 390 395	2103
cac agg caa tgg ttc cta gac ctg ccg tta cca tgg ttg ccc gga gcg His Arg Gln Trp Phe Leu Asp Leu Pro Leu Pro Trp Leu Pro Gly Ala 400 405 410	2151
gac aca caa ggg tca aat tgg ata cag aaa gag aca ttg gtc act ttc Asp Thr Gln Gly Ser Asn Trp Ile Gln Lys Glu Thr Leu Val Thr Phe 415 420 425 430	2199
aaa aat ccc cat gcg aag aaa cag gat gtt gtt gtt tta gga tcc caa Lys Asn Pro His Lys Lys Gln Asp Val Val Val Leu Gly Ser Gln 435 440 445	2247
gaa ggg gcc atg cac aca gca ctt aca ggg gcc aca gaa atc caa atg Glu Gly Ala Met His Thr Ala Leu Thr Gly Ala Thr Glu Ile Gln Met 450 455 460	2295
tca tca gga aac tta ctc ttc aca gga cat ctc aag tgc agg ctg aga Ser Ser Gly Asn Leu Leu Phe Thr Gly His Leu Lys Cys Arg Leu Arg 465 470 475	2343
atg gac aag cta cag ctc aaa gga atg tca tac tct atg tgc aca gga Met Asp Lys Leu Gln Leu Lys Gly Met Ser Tyr Ser Met Cys Thr Gly 480 485 490	2391
aag ttt aaa gtt gtg aag gaa ata gca gaa aca caa cat gga aca ata Lys Phe Lys Val Val Lys Glu Ile Ala Glu Thr Gln His Gly Thr Ile 495 500 505 510	2439
gtt atc aga gtg caa tat gaa ggg gac ggc tct cca tgc aag atc cct Val Ile Arg Val Gln Tyr Glu Gly Asp Gly Ser Pro Cys Lys Ile Pro 515 520 525	2487
ttt gag ata atg gat ttg gaa aaa aga cat gtc tta ggt cgc ctg att Phe Glu Ile Met Asp Leu Glu Lys Arg His Val Leu Gly Arg Leu Ile 530 535 540	2535
aca gtc aac cca att gtg aca gaa aaa gat agc cca gtc aac ata gaa Thr Val Asn Pro Ile Val Thr Glu Lys Asp Ser Pro Val Asn Ile Glu 545 550 555	2583
gca gaa cct cca ttc gga gac agc tac atc atc ata gga gta gag ccg Ala Glu Pro Pro Phe Gly Asp Ser Tyr Ile Ile Ile Gly Val Glu Pro 560 565 570	2631
gga caa ctg aag ctc aac tgg ttt aag aaa gga agt tct atc ggc caa Gly Gln Leu Lys Leu Asn Trp Phe Lys Lys Gly Ser Ser Ile Gly Gln 575 580 585 590	2679

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Met Phe Glu Thr Thr Met Arg Gly Ala Lys Arg Met Ala Ile Leu Gly	
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Asp Thr Ala Trp Asp Phe Gly Ser Leu Gly Gly Val Phe Thr Ser Ile	
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Trp Ile Gly Met Asn Ser Arg Ser Thr Ser Leu Ser Val Thr Leu Val	
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<210> 43

<211> 685

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
 synthetic construct

<400> 43

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Glu	Pro	His	Met	Ile	Val	Ser	Arg	Gln	Glu	Lys	Gly	Lys	Ser	Leu	Leu
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Phe	Lys	Thr	Glu	Asp	Gly	Val	Asn	Met	Cys	Thr	Leu	Met	Ala	Met	Asp
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Leu	Gly	Glu	Leu	Cys	Glu	Asp	Thr	Ile	Thr	Tyr	Lys	Cys	Pro	Leu	Leu
65				70					75					80	
Arg	Gln	Asn	Glu	Pro	Glu	Asp	Ile	Asp	Cys	Trp	Cys	Asn	Ser	Thr	Ser
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Thr	Trp	Val	Thr	Tyr	Gly	Thr	Cys	Thr	Thr	Met	Gly	Glu	His	Arg	Arg
		100					105						110		
Glu	Lys	Arg	Ser	Val	Ala	Leu	Val	Pro	His	Val	Gly	Met	Gly	Leu	Glu
		115				120					125				
Thr	Arg	Thr	Glu	Thr	Trp	Met	Ser	Ser	Glu	Gly	Ala	Trp	Lys	His	Val
	130				135						140				
Gln	Arg	Ile	Glu	Thr	Trp	Ile	Leu	Arg	His	Pro	Gly	Phe	Thr	Met	Met
145				150					155					160	
Ala	Ala	Ile	Leu	Ala	Tyr	Thr	Ile	Gly	Thr	Thr	His	Phe	Gln	Arg	Ala
			165					170					175		
Leu	Ile	Phe	Ile	Leu	Leu	Thr	Ala	Val	Thr	Pro	Ser	Met	Thr	Met	Arg
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Cys	Ile	Gly	Met	Ser	Asn	Arg	Asp	Phe	Val	Glu	Gly	Val	Ser	Gly	Gly
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Ser	Trp	Val	Asp	Ile	Val	Leu	Glu	His	Gly	Ser	Cys	Val	Thr	Thr	Met
	210				215						220				
Ala	Lys	Asn	Lys	Pro	Thr	Leu	Asp	Phe	Glu	Leu	Ile	Lys	Thr	Glu	Ala
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Lys	Gln	Pro	Ala	Thr	Leu	Arg	Lys	Tyr	Cys	Ile	Glu	Ala	Lys	Leu	Thr
			245					250					255		
Asn	Thr	Thr	Thr	Glu	Ser	Arg	Cys	Pro	Thr	Gln	Gly	Glu	Pro	Ser	Leu
		260				265						270			
Asn	Glu	Glu	Gln	Asp	Lys	Arg	Phe	Val	Cys	Lys	His	Ser	Met	Val	Asp
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Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys	Gly	Gly	Ile	Val
	290				295						300				
Thr	Cys	Ala	Met	Phe	Arg	Cys	Lys	Lys	Asn	Met	Glu	Gly	Lys	Val	Val
305				310					315					320	
Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Thr	Ile	Val	Ile	Thr	Pro	His	Ser	Gly
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Glu	Glu	His	Ala	Val	Gly	Asn	Asp	Thr	Gly	Lys	His	Gly	Lys	Glu	Ile
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Lys	Ile	Thr	Pro	Gln	Ser	Ser	Ile	Thr	Glu	Ala	Glu	Leu	Thr	Gly	Tyr
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Gly	Thr	Val	Thr	Met	Glu	Cys	Ser	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Asn
	370				375						380				

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 Gln Trp Phe Leu Asp Leu Pro Leu Pro Trp Leu Pro Gly Ala Asp Thr
 405 410 415
 Gln Gly Ser Asn Trp Ile Gln Lys Glu Thr Leu Val Thr Phe Lys Asn
 420 425 430
 Pro His Ala Lys Lys Gln Asp Val Val Val Leu Gly Ser Gln Glu Gly
 435 440 445
 Ala Met His Thr Ala Leu Thr Gly Ala Thr Glu Ile Gln Met Ser Ser
 450 455 460
 Gly Asn Leu Leu Phe Thr Gly His Leu Lys Cys Arg Leu Arg Met Asp
 465 470 475 480
 Lys Leu Gln Leu Lys Gly Met Ser Tyr Ser Met Cys Thr Gly Lys Phe
 485 490 495
 Lys Val Val Lys Glu Ile Ala Glu Thr Gln His Gly Thr Ile Val Ile
 500 505 510
 Arg Val Gln Tyr Glu Gly Asp Gly Ser Pro Cys Lys Ile Pro Phe Glu
 515 520 525
 Ile Met Asp Leu Glu Lys Arg His Val Leu Gly Arg Leu Ile Thr Val
 530 535 540
 Asn Pro Ile Val Thr Glu Lys Asp Ser Pro Val Asn Ile Glu Ala Glu
 545 550 555 560
 Pro Pro Phe Gly Asp Ser Tyr Ile Ile Ile Gly Val Glu Pro Gly Gln
 565 570 575
 Leu Lys Leu Asn Trp Phe Lys Lys Gly Ser Ser Ile Gly Gln Met Phe
 580 585 590
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 595 600 605
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 610 615 620
 Ala Leu His Gln Val Phe Gly Ala Ile Tyr Gly Ala Ala Phe Ser Gly
 625 630 635 640
 Val Ser Trp Thr Met Lys Ile Leu Ile Gly Val Ile Ile Thr Trp Ile
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<210> 44

<211> 5293

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> CDS

<222> (910)...(2964)

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gattattgac	tagttattaa	tagtaataca	ttacgggggc	attagttcat	agcccatata	300
tggagttccg	cgttacataa	cttacggtaa	atggcccggc	tggtcgaccg	cccaacgacc	360
cccggccatt	gacgtcaata	atgacgtatg	ttcccatagt	aacgccaata	gggactttcc	420
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gccgccgcc atg ggc aag agg tcc gcc ggc tca atc atg tgg ctc gcg agc	951
Met Gly Lys Arg Ser Ala Gly Ser Ile Met Trp Leu Ala Ser	
1 5 10	
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Leu Ala Val Val Ile Ala Cys Ala Gly Ala Phe His Leu Thr Thr Arg	
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Asn Gly Glu Pro Lys His Met Ile Val Ser Arg Gln Glu Lys Gly Lys Ser	
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Leu Leu Phe Lys Thr Glu Asp Gly Val Asn Met Cys Thr Leu Met Ala	
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Met Asp Leu Gly Glu Leu Cys Glu Asp Thr Ile Thr Tyr Lys Cys Pro	
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Leu Leu Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys Asn Ser	
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Leu Glu Thr Arg Thr Glu Thr Trp Met Ser Ser Glu Gly Ala Trp Lys	
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Met Met Ala Ala Ile Leu Ala Tyr Thr Ile Gly Thr Thr His Phe Gln	
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aga gcc ctg att ttc atc tta ctg aca gct gtc act cct tca atg aca	1479
Arg Ala Leu Ile Phe Ile Leu Leu Thr Ala Val Thr Pro Ser Met Thr	
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Gly Gly Ser Trp Val Asp Ile Val Leu Glu His Gly Ser Cys Val Thr	
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aataaggcg acacggaaat gttgaatact catactcttc ctttttcaat attattgaag 5184
catttatcag gggtattgtc tcatgagcgg atacatattt gaatgtattt agaaaaataa 5244
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<210> 45

<211> 685

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 45

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          20           25           30
Glu Pro His Met Ile Val Ser Arg Gln Glu Lys Gly Lys Ser Leu Leu
          35           40           45
Phe Lys Thr Glu Asp Gly Val Asn Met Cys Thr Leu Met Ala Met Asp
          50           55           60
Leu Gly Glu Leu Cys Glu Asp Thr Ile Thr Tyr Lys Cys Pro Leu Leu
65           70           75           80
Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys Asn Ser Thr Ser
          85           90           95
Thr Trp Val Thr Tyr Gly Thr Cys Thr Thr Met Gly Glu His Arg Arg
          100          105          110
Glu Lys Arg Ser Val Ala Leu Val Pro His Val Gly Met Gly Leu Glu
          115          120          125

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Gln	Arg	Ile	Glu	Thr	Trp	Ile	Leu	Arg	His	Pro	Gly	Phe	Thr	Met	Met
145					150					155					160
Ala	Ala	Ile	Leu	Ala	Tyr	Thr	Ile	Gly	Thr	Thr	His	Phe	Gln	Arg	Ala
				165					170					175	
Leu	Ile	Phe	Ile	Leu	Leu	Thr	Ala	Val	Thr	Pro	Ser	Met	Thr	Met	Arg
			180					185					190		
Cys	Ile	Gly	Met	Ser	Asn	Arg	Asp	Phe	Val	Glu	Gly	Val	Ser	Gly	Gly
	195						200					205			
Ser	Trp	Val	Asp	Ile	Val	Leu	Glu	His	Gly	Ser	Cys	Val	Thr	Thr	Met
	210					215					220				
Ala	Lys	Asn	Lys	Pro	Thr	Leu	Asp	Phe	Glu	Leu	Ile	Lys	Thr	Glu	Ala
225					230					235					240
Lys	Gln	Pro	Ala	Thr	Leu	Arg	Lys	Tyr	Cys	Ile	Glu	Ala	Lys	Leu	Thr
				245					250					255	
Asn	Thr	Thr	Thr	Glu	Ser	Arg	Cys	Pro	Thr	Gln	Gly	Glu	Pro	Ser	Leu
			260					265					270		
Asn	Glu	Glu	Gln	Asp	Lys	Arg	Phe	Val	Cys	Lys	His	Ser	Met	Val	Asp
	275						280					285			
Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys	Gly	Gly	Ile	Val
	290					295					300				
Thr	Cys	Ala	Met	Phe	Arg	Cys	Lys	Lys	Asn	Met	Glu	Gly	Lys	Val	Val
305					310					315					320
Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Thr	Ile	Val	Ile	Thr	Pro	His	Ser	Gly
			325						330					335	
Glu	Glu	His	Ala	Val	Gly	Asn	Asp	Thr	Gly	Lys	His	Gly	Lys	Glu	Ile
			340					345					350		
Lys	Ile	Thr	Pro	Gln	Ser	Ser	Ile	Thr	Glu	Ala	Glu	Leu	Thr	Gly	Tyr
	355						360					365			
Gly	Thr	Val	Thr	Met	Glu	Cys	Ser	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Asn
	370					375					380				
Glu	Met	Val	Leu	Leu	Gln	Met	Glu	Asn	Lys	Ala	Trp	Leu	Val	His	Arg
385					390					395					400
Gln	Trp	Phe	Leu	Asp	Leu	Pro	Leu	Pro	Trp	Leu	Pro	Gly	Ala	Asp	Thr
			405						410					415	
Gln	Gly	Ser	Asn	Trp	Ile	Gln	Lys	Glu	Thr	Leu	Val	Thr	Phe	Lys	Asn
			420					425					430		
Pro	His	Ala	Lys	Lys	Gln	Asp	Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly
	435						440					445			
Ala	Met	His	Thr	Ala	Leu	Thr	Gly	Ala	Thr	Glu	Ile	Gln	Met	Ser	Ser
	450					455					460				
Gly	Asn	Leu	Leu	Phe	Thr	Gly	His	Leu	Lys	Cys	Arg	Leu	Arg	Met	Asp
465					470					475					480
Lys	Leu	Gln	Leu	Lys	Gly	Met	Ser	Tyr	Ser	Met	Cys	Thr	Gly	Lys	Phe
			485						490					495	
Lys	Val	Val	Lys	Glu	Ile	Ala	Glu	Thr	Gln	His	Gly	Thr	Ile	Val	Ile
			500					505					510		
Arg	Val	Gln	Tyr	Glu	Gly	Asp	Gly	Ser	Pro	Cys	Lys	Ile	Pro	Phe	Glu
	515						520					525			
Ile	Met	Asp	Leu	Glu	Lys	Arg	His	Val	Leu	Gly	Arg	Leu	Ile	Thr	Val
	530					535					540				
Asn	Pro	Ile	Val	Thr	Glu	Lys	Asp	Ser	Pro	Val	Asn	Ile	Glu	Ala	Glu
545					550					555					560
Pro	Pro	Phe	Gly	Asp	Ser	His	Ile	Ile	Ile	Gly	Val	Glu	Pro	Gly	Gln
			565						570					575	
Leu	Lys	Leu	Asn	Trp	Phe	Lys	Lys	Gly	Ser	Ser	Ile	Gly	Gln	Met	Phe
			580					585					590		
Glu	Thr	Thr	Met	Arg	Gly	Ala	Lys	Arg	Met	Ala	Ile	Leu	Gly	Asp	Thr
	595						600					605			

Ala Trp Asp Phe Gly Ser Leu Gly Gly Val Phe Thr Ser Ile Gly Lys
 610 615 620
 Ala Leu His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly
 625 630 635 640
 Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met
 645 650 655
 Gly Val Asn Ala Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu Ala Thr
 660 665 670
 Gly Gly Val Leu Val Phe Leu Ala Thr Asn Val His Ala
 675 680 685

<210> 46

<211> 5293

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<221> CDS

<222> (910)...(2964)

<400> 46

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cgagcaaaat	ttaagctaca	acaaggcaag	gcttgaccga	caattgcatg	aagaatctgc	180
ttaggggttag	gcgtttttgcg	ctgcttcgcg	atgtacgggc	cagatatacg	cgttgacatt	240
gattattgac	tagttattaa	tagtaatcaa	ttacgggggc	attagttcat	agcccatata	300
tggagttccg	cgttacataa	cttacggtaa	atggcccggc	tggtcgaccg	cccaacgacc	360
ccgcccatt	gacgtcaata	atgacgtatg	ttcccatagt	aacgcccaata	gggactttcc	420
attgacgtca	atgggtggac	tatttacggg	aaactgcccc	cttggcagta	catcaagtgt	480
atcatatgcc	aagtacgcc	cctattgacg	tcaatgacgg	taaatggccc	gcttggcatt	540
atgcccagta	catgacctta	tgggactttc	ctacttggca	gtacatctac	gtattagtca	600
tcgctattac	catggtgatg	cggttttggc	agtacatcaa	tgggcgtgga	tagcggtttg	660
actcacgggg	atttccaagt	ctccacccca	ttgacgtcaa	tgggagtttg	ttttggcacc	720
aaaatcaacg	ggactttcca	aaatgtcgta	acaactccgc	cccattgacg	caaatgggcg	780
gtaggcgtgt	acgggtgggag	gtctatataa	gcagagctct	ctggctaact	agagaacca	840
ctgcttactg	gcttatcgaa	attaatacga	ctcactatag	ggagacccaa	gcttggtacc	900
gccgcgcgcc	atg ggc aag	agg tcc gcc	ggc tca atc	atg tgg ctc	gcg agc	951
	Met Gly Lys	Arg Ser Ala	Gly Ser Ile	Met Trp Leu	Ala Ser	
	1	5		10		

ttg gca gtt gtc	ata gct tgt	gca ggc gcc	ttc cat tta	acc aca	cgt	999
Leu Ala Val Val	Ile Ala Cys	Ala Gly Ala	Phe His Leu	Thr Thr	Arg	
15	20		25		30	

aac gga gaa cca	cac atg atc	gtc agc aga	caa gag aaa	ggg aaa	agt	1047
Asn Gly Glu Pro	His Met Ile	Val Ser Arg	Gln Glu Lys	Gly Lys	Ser	
	35	40		45		

ctt ctg ttt aaa	aca gag gat	ggc gtg aac	atg tgt acc	ctc atg gcc		1095
Leu Leu Phe Lys	Thr Glu Asp	Gly Val Asn	Met Cys Thr	Leu Met Ala		
	50	55		60		

atg gac ctt ggt	gaa ttg tgt	gaa gac aca	atc acg tac	aag tgt ccc		1143
Met Asp Leu Gly	Glu Leu Cys	Glu Asp Thr	Ile Thr Tyr	Lys Cys Pro		
	65	70		75		

ctt ctc agg cag aat gag cca gaa gac ata gac tgt tgg tgc aac tct	1191
Leu Leu Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys Asn Ser	
80 85 90	
acg tcc acg tgg gta act tat ggg acg tgt acc acc atg gga gaa cat	1239
Thr Ser Thr Trp Val Thr Tyr Gly Thr Cys Thr Thr Met Gly Glu His	
95 100 105 110	
aga aga gaa aaa aga tca gtg gca ctc gtt cca cat gtg gga atg gga	1287
Arg Arg Glu Lys Arg Ser Val Ala Leu Val Pro His Val Gly Met Gly	
115 120 125	
ctg gag aca cga act gaa aca tgg atg tca tca gaa ggg gcc tgg aaa	1335
Leu Glu Thr Arg Thr Glu Thr Trp Met Ser Ser Glu Gly Ala Trp Lys	
130 135 140	
cat gtc cag aga att gaa act tgg atc ttg aga cat cca ggc ttc acc	1383
His Val Gln Arg Ile Glu Thr Trp Ile Leu Arg His Pro Gly Phe Thr	
145 150 155	
atg atg gca gca atc ctg gca tac acc ata gga acg aca cat ttc caa	1431
Met Met Ala Ala Ile Leu Ala Tyr Thr Ile Gly Thr Thr His Phe Gln	
160 165 170	
aga gcc ctg att ttc atc tta ctg aca gct gtc act cct tca atg aca	1479
Arg Ala Leu Ile Phe Ile Leu Leu Thr Ala Val Thr Pro Ser Met Thr	
175 180 185 190	
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Met Arg Cys Ile Gly Met Ser Asn Arg Asp Phe Val Glu Gly Val Ser	
195 200 205	
gga gga agc tgg gtt gac ata gtc tta gaa cat ggg agc tgt gtg acg	1575
Gly Gly Ser Trp Val Asp Ile Val Leu Glu His Gly Ser Cys Val Thr	
210 215 220	
acg atg gca aaa aac aaa cca aca ttg gat ttt gaa ctg ata aaa aca	1623
Thr Met Ala Lys Asn Lys Pro Thr Leu Asp Phe Glu Leu Ile Lys Thr	
225 230 235	
gaa gcc aaa cag cct gcc acc cta agg aag tac tgt ata gag gca aag	1671
Glu Ala Lys Gln Pro Ala Thr Leu Arg Lys Tyr Cys Ile Glu Ala Lys	
240 245 250	
cta acc aac aca aca aca gaa tct cgc tgc cca aca caa ggg gaa ccc	1719
Leu Thr Asn Thr Thr Thr Glu Ser Arg Cys Pro Thr Gln Gly Glu Pro	
255 260 265 270	
agc cta aat gaa gag cag gac aaa agg ttc gtc tgc aaa cac tcc atg	1767
Ser Leu Asn Glu Glu Gln Asp Lys Arg Phe Val Cys Lys His Ser Met	
275 280 285	
gta gac aga gga tgg gga aat gga tgt gga cta ttt gga aag gga ggc	1815
Val Asp Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Gly	
290 295 300	
att gtg acc tgt gct atg ttc aga tgc aaa aag aac atg gaa gga aaa	1863
Ile Val Thr Cys Ala Met Phe Arg Cys Lys Lys Asn Met Glu Gly Lys	
305 310 315	

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tca ggg gaa gag cat gca gtc gga aat gac aca gga aaa cat ggc aag Ser Gly Glu Glu His Ala Val Gly Asn Asp Thr Gly Lys His Gly Lys 335 340 345 350	1959
gaa atc aaa ata aca cca cag agt tcc atc aca gaa gca gaa ttg aca Glu Ile Lys Ile Thr Pro Gln Ser Ser Ile Thr Glu Ala Glu Leu Thr 355 360 365	2007
ggc tat ggc act gtc aca atg gag tgc tct cca aga acg ggc ctc gac Gly Tyr Gly Thr Val Thr Met Glu Cys Ser Pro Arg Thr Gly Leu Asp 370 375 380	2055
ttc aat gag atg gtg ttg ttg cag atg gaa aat aaa gct tgg ctg gtg Phe Asn Glu Met Val Leu Leu Gln Met Glu Asn Lys Ala Trp Leu Val 385 390 395	2103
cac agg caa tgg ttc cta gac ctg ccg tta cca tgg ttg ccc gga gcg His Arg Gln Trp Phe Leu Asp Leu Pro Leu Pro Trp Leu Pro Gly Ala 400 405 410	2151
gac aca caa ggg tca aat tgg ata cag aaa gag aca ttg gtc act ttc Asp Thr Gln Gly Ser Asn Trp Ile Gln Lys Glu Thr Leu Val Thr Phe 415 420 425 430	2199
aaa aat ccc cat gcg aag aaa cag gat gtt gtt gtt tta gga tcc caa Lys Asn Pro His Ala Lys Lys Gln Asp Val Val Val Leu Gly Ser Gln 435 440 445	2247
gaa ggg gcc atg cac aca gca ctt aca ggg gcc aca gaa atc caa atg Glu Gly Ala Met His Thr Ala Leu Thr Gly Ala Thr Glu Ile Gln Met 450 455 460	2295
tca tca gga aac tta ctc ttc aca gga cat ctc aag tgc agg ctg aga Ser Ser Gly Asn Leu Leu Phe Thr Gly His Leu Lys Cys Arg Leu Arg 465 470 475	2343
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aag ttt aaa gtt gtg aag gaa ata gca gaa aca caa cat gga aca ata Lys Phe Lys Val Val Lys Glu Ile Ala Glu Thr Gln His Gly Thr Ile 495 500 505 510	2439
gtt atc aga gtg caa tat gaa ggg gac ggc tct cca tgc aag atc cct Val Ile Arg Val Gln Tyr Glu Gly Asp Gly Ser Pro Cys Lys Ile Pro 515 520 525	2487
ttt gag ata atg gat ttg gaa aaa aga cat gtc tta ggt cgc ctg att Phe Glu Ile Met Asp Leu Glu Lys Arg His Val Leu Gly Arg Leu Ile 530 535 540	2535
aca gtc aac cca att gtg aca gaa aaa gat agc cca gtc aac ata gaa Thr Val Asn Pro Ile Val Thr Glu Lys Asp Ser Pro Val Asn Ile Glu 545 550 555	2583

gca gaa cct cca ttc gga gac agc tac atc atc ata gga gta gag ccg	2631
Ala Glu Pro Pro Phe Gly Asp Ser Tyr Ile Ile Ile Gly Val Glu Pro	
560 565 570	
gga caa ctg aag ctc aac tgg ttt aag aaa gga agc acg ctg ggc aag	2679
Gly Gln Leu Lys Leu Asn Trp Phe Lys Lys Gly Ser Thr Leu Gly Lys	
575 580 585 590	
gcc ttt tca aca act ttg aag gga gct caa aga ctg gca gcg ttg ggc	2727
Ala Phe Ser Thr Leu Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly	
595 600 605	
gac aca gcc tgg gac ttt ggc tct att gga ggg gtc ttc aac tcc ata	2775
Asp Thr Ala Trp Asp Phe Gly Ser Ile Gly Gly Val Phe Asn Ser Ile	
610 615 620	
gga aaa gcc gtt cac caa gtg ttt ggt ggt gcc ttc aga aca ctc ttt	2823
Gly Lys Ala Val His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe	
625 630 635	
ggg gga atg tct tgg atc aca caa ggg cta atg ggt gcc cta ctg ctc	2871
Gly Gly Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu	
640 645 650	
tgg atg ggc gtc aac gca cga gac cga tca att gct ttg gcc ttc tta	2919
Trp Met Gly Val Asn Ala Arg Asp Arg Ser Ile Ala Leu Ala Phe Leu	
655 660 665 670	
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Ala Thr Gly Gly Val Leu Val Phe Leu Ala Thr Asn Val His Ala	
675 680 685	
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catttatcag ggttattgtc tcatgagcgg atacatattt gaatgtattt agaaaaataa 5244
acaaataggg gttccgcgca catttccccg aaaagtgcc a cctgacgtc 5293

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<210> 47

<211> 685

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of artificial sequence; note =
synthetic construct

<400> 47

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Glu Pro His Met Ile Val Ser Arg Gln Glu Lys Gly Lys Ser Leu Leu
 35          40          45
Phe Lys Thr Glu Asp Gly Val Asn Met Cys Thr Leu Met Ala Met Asp
 50          55          60
Leu Gly Glu Leu Cys Glu Asp Thr Ile Thr Tyr Lys Cys Pro Leu Leu
 65          70          75          80
Arg Gln Asn Glu Pro Glu Asp Ile Asp Cys Trp Cys Asn Ser Thr Ser
 85          90          95
Thr Trp Val Thr Tyr Gly Thr Cys Thr Thr Met Gly Glu His Arg Arg
100          105          110
Glu Lys Arg Ser Val Ala Leu Val Pro His Val Gly Met Gly Leu Glu
115          120          125
Thr Arg Thr Glu Thr Trp Met Ser Ser Glu Gly Ala Trp Lys His Val
130          135          140
Gln Arg Ile Glu Thr Trp Ile Leu Arg His Pro Gly Phe Thr Met Met
145          150          155          160
Ala Ala Ile Leu Ala Tyr Thr Ile Gly Thr Thr His Phe Gln Arg Ala
165          170          175
Leu Ile Phe Ile Leu Leu Thr Ala Val Thr Pro Ser Met Thr Met Arg
180          185          190
Cys Ile Gly Met Ser Asn Arg Asp Phe Val Glu Gly Val Ser Gly Gly
195          200          205
Ser Trp Val Asp Ile Val Leu Glu His Gly Ser Cys Val Thr Thr Met
210          215          220
Ala Lys Asn Lys Pro Thr Leu Asp Phe Glu Leu Ile Lys Thr Glu Ala
225          230          235          240
Lys Gln Pro Ala Thr Leu Arg Lys Tyr Cys Ile Glu Ala Lys Leu Thr
245          250          255
Asn Thr Thr Thr Glu Ser Arg Cys Pro Thr Gln Gly Glu Pro Ser Leu
260          265          270
Asn Glu Glu Gln Asp Lys Arg Phe Val Cys Lys His Ser Met Val Asp
275          280          285
Arg Gly Trp Gly Asn Gly Cys Gly Leu Phe Gly Lys Gly Gly Ile Val
290          295          300
Thr Cys Ala Met Phe Arg Cys Lys Lys Asn Met Glu Gly Lys Val Val
305          310          315          320

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Gln Pro Glu Asn Leu Glu Tyr Thr Ile Val Ile Thr Pro His Ser Gly
 325 330 335
 Glu Glu His Ala Val Gly Asn Asp Thr Gly Lys His Gly Lys Glu Ile
 340 345 350
 Lys Ile Thr Pro Gln Ser Ser Ile Thr Glu Ala Glu Leu Thr Gly Tyr
 355 360 365
 Gly Thr Val Thr Met Glu Cys Ser Pro Arg Thr Gly Leu Asp Phe Asn
 370 375 380
 Glu Met Val Leu Leu Gln Met Glu Asn Lys Ala Trp Leu Val His Arg
 385 390 395 400
 Gln Trp Phe Leu Asp Leu Pro Leu Pro Trp Leu Pro Gly Ala Asp Thr
 405 410 415
 Gln Gly Ser Asn Trp Ile Gln Lys Glu Thr Leu Val Thr Phe Lys Asn
 420 425 430
 Pro His Ala Lys Lys Gln Asp Val Val Leu Gly Ser Gln Glu Gly
 435 440 445
 Ala Met His Thr Ala Leu Thr Gly Ala Thr Glu Ile Gln Met Ser Ser
 450 455 460
 Gly Asn Leu Leu Phe Thr Gly His Leu Lys Cys Arg Leu Arg Met Asp
 465 470 475 480
 Lys Leu Gln Leu Lys Gly Met Ser Tyr Ser Met Cys Thr Gly Lys Phe
 485 490 495
 Lys Val Val Lys Glu Ile Ala Glu Thr Gln His Gly Thr Ile Val Ile
 500 505 510
 Arg Val Gln Tyr Glu Gly Asp Gly Ser Pro Cys Lys Ile Pro Phe Glu
 515 520 525
 Ile Met Asp Leu Glu Lys Arg His Val Leu Gly Arg Leu Ile Thr Val
 530 535 540
 Asn Pro Ile Val Thr Glu Lys Asp Ser Pro Val Asn Ile Glu Ala Glu
 545 550 555 560
 Pro Pro Phe Gly Asp Ser Tyr Ile Ile Ile Gly Val Glu Pro Gly Gln
 565 570 575
 Leu Lys Leu Asn Trp Phe Lys Lys Gly Ser Thr Leu Gly Lys Ala Phe
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 Ser Thr Thr Leu Lys Gly Ala Gln Arg Leu Ala Ala Leu Gly Asp Thr
 595 600 605
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 610 615 620
 Ala Val His Gln Val Phe Gly Gly Ala Phe Arg Thr Leu Phe Gly Gly
 625 630 635 640
 Met Ser Trp Ile Thr Gln Gly Leu Met Gly Ala Leu Leu Leu Trp Met
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<213> Artificial Sequence

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<213> Artificial Sequence

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33

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<211> 11

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<213> Artificial Sequence

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<223> Description of artificial sequence; note =
synthetic construct

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Asn Trp Lys Lys Gly Ser Thr Leu Gly Lys Ala

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5

10

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/10764

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : Please See Extra Sheet.

US CL : 435/5, 7.1, 69.1; 530/387.1; 424/218.1; 536/23.72

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/5, 7.1, 69.1; 530/387.1; 424/218.1; 536/23.72

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

MEDLINE, BIOSIS, WPIDS, USPATFUL

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,494,671 A (LAI, C., et al.) 27 February 1996, see entire document.	1-43
Y	US 6,074,865 A (KELLY, E. P. and A. D. KING) 13 June 2000, see entire document.	1-43
Y	US 6,165,477 A (IVY, J., et al.) 26 December 2000, see entire document.	1-43

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principles or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"G"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search 18 JUNE 2002	Date of mailing of the international search report 18 JUL 2002
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-9230	Authorized officer J. S. PARKIN Telephone No. (703) 308-1234

INTERNATIONAL SEARCH REPORT .

International application No.
PCT/US02/10764

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (7):

C12Q 1/70; G01N 33/53; C12P 21/06; C07K 16/00; C07H 21/04; A61K 39/12